Introduction

The alarming increase in the bacterial resistance towards the existing chemotherapeutics has paved the way for exploration of new alternatives. One of the options – folk medicines could provide a platform for yielding effective compounds with potent activity against microbes. The last three decades have seen the development of several synthetic drugs, but the resistance towards these drugs is also developing at a faster stroke, as the bacteria can acquire and transmit genes responsible for antibiotic resistance [1]. The main advantage of natural agents is that, the crude extracts contain a mixture of compounds like phenols, acids, esters, aldehydes etc., for which it is difficult to develop resistance by bacteria unlike the synthetic antibiotics that contain a single compound [2]. The world health organization has taken an initiative in the development of plant based health care, making it available to maximum population [3]. Developing countries have fast making use of the traditional medicines with already 1400 preparations in use.

Gymnema sylvestre R. Br belonging to the family Asclepiadaceae, is found in various parts of India and Tropical Africa. Its major active principles are triterpenoid saponins, responsible for the various hepatoprotective [5] and antidiabetic activities [6]. The other important phytoconstituents are flavones, anthroquinones, aconitines, phytin, quercitol, lupeol, β-amyrin related glycosides, alkaloids and stigmasteryl [7]. The methanol extracts of G. sylvestre were used for evaluating the antimicrobial activity of the aerial and root parts in two pH ranges. The extraction at acidic pH range was much superior in activity towards all the microorganisms in comparison to the neutral range extraction.

Evaluation of Gymnema sylvestre Antimicrobial Activity in Methanol

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Abstract

G. sylvestre is a medicinal plant known for its sugar destroying property, as an anti-diabetic agent. The major phytoconstituents are the triterpenoid saponins, responsible for the various activities. The antimicrobial activity of this plant has been assessed in methanol as the solvent system for the extraction of active principles. The gram positive and gram negative organisms used in the study, have shown susceptibility towards the extracts, with the root extracts at acidic pH, showing higher activity. E. coli and E. cloacae were found to be the most sensitive and Pseudomonas aeruginosa, the resistant type of microorganisms, based on the results obtained from the zones of inhibition. The broad spectrum activity of the plant can be utilised in the development of new antimicrobial drugs.

Key Words: G. sylvestre, Aerial and Root extracts, Antimicrobial activity, Methanol, Antibiotic resistance.

Material and Methods

Materials

Plant material was obtained from A G Biotek. Methanol, Amikacin and bacteriological media (Mueller Hinton Agar (MHA) & Mueller Hinton Broth (MHB)) were procured from Himedia.

Extract preparation

The plants were thoroughly washed and separated in different parts like shoots (aerial parts) and roots. They were oven dried at 60°C for 24h to avoid moisture interference and finely ground using a blender. Four different extracts (aerial & root) were obtained at two different pH ranges: acidic pH (5.5) and neutral pH (7.0). The pH was adjusted using 0.1N HCl.

Microbial cultures

The MTCC cultures – Bacillus subtilis MTCC 2391, Escherichia coli MTCC 1563 and Pseudomonas aeruginosa MTCC 6642 were obtained from IMTECH, Chandigarh. The other test organisms were obtained as clinical isolates from Global hospitals, Hyderabad, India. All the cultures were tested for purity and maintained on Mueller Hinton agar at 4°C. The strains were sub-cultured every fortnight and reactivated in Mueller Hinton broth before the antimicrobial assay.

Antimicrobial assay

The antimicrobial activity of the extracts was assessed using the Agar well diffusion method suggested by Perez et al in 1920 [8]. An inoculum size of 10^6 colony forming units (cfu)/ml of bacteria was used for the pour plate technique. A borer with a diameter of 8mm was used for making the wells in the Müller Hinton agar plate containing antibiotics at a concentration of 100 μg/ml.
the MHA plates. Plant extracts at a concentration of 500 μg in the respective solvents were dispensed into each of the wells. Amikacin, a broad spectrum antibiotic was used at a concentration of 50 μg as the positive reference standard. The zones of inhibition around each well were measured after a 24h incubation period at 37°C. The sensitivity of the pathogens towards the extracts was determined by comparing the inhibitory zones around the well. All the assays were performed in triplicate and expressed.

Statistical analysis
Results were expressed as the means ± standard deviations from the obtained triplicate data. The data was compared by least significant difference test using Statistical Analysis System (ver 9.1).

Results and Discussion
Plants synthesize a wide variety of substances as part of defence mechanism against attack by insects, herbivores and microorganisms [9]. The increasing incidence of antibiotic resistance among microbes is paving the way in search of new drugs from different sources. There is a pressing need to discover new compounds differing in structure as well as activity. One such source is the plant biodiversity, as there is a widespread belief that, green medicines are nontoxic, healthier and safe, in comparison to the synthetic drugs. The antimicrobial activity of plant extracts towards the drug resistant bacteria has been studied by many researchers [10].

<table>
<thead>
<tr>
<th>Name of the Organism</th>
<th>Diameter of the inhibition zone (mm*)</th>
<th>Antibiotic</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Aerial Acidic(5.5)</td>
<td>Neutral(7.0)</td>
</tr>
<tr>
<td>Gram positive</td>
<td></td>
<td></td>
</tr>
<tr>
<td>B. subtilis</td>
<td>15.17±0.30</td>
<td>13.96±0.22</td>
</tr>
<tr>
<td>E. faecalis</td>
<td>12.42±0.01</td>
<td>10.83±0.17</td>
</tr>
<tr>
<td>S. aureus</td>
<td>13.98±0.01</td>
<td>13.32±0.30</td>
</tr>
<tr>
<td>S. epidermis</td>
<td>13.61±0.36</td>
<td>12.70±0.09</td>
</tr>
<tr>
<td>Gram negative</td>
<td></td>
<td></td>
</tr>
<tr>
<td>E. aerogene</td>
<td>15.46±0.13</td>
<td>13.97±0.29</td>
</tr>
<tr>
<td>E. cloacae</td>
<td>15.25±0.03</td>
<td>13.23±0.29</td>
</tr>
<tr>
<td>E. coli</td>
<td>17.64±0.80</td>
<td>15.86±0.45</td>
</tr>
<tr>
<td>K. pneumonia</td>
<td>13.19±0.34</td>
<td>11.14±0.25</td>
</tr>
<tr>
<td>P. aeruginosa</td>
<td>11.87±0.19</td>
<td>10.88±0.24</td>
</tr>
<tr>
<td>S. typhimurium</td>
<td>10.45±0.29</td>
<td>12.64±0.21</td>
</tr>
</tbody>
</table>

*Borer is 8mm

In the present study, methanol has been used for extraction of active principles from the different parts of G. sylvestre. The presence of phytoconstituents were analysed separately in aerial and root parts. The efficient extraction of active principles depends on the competence of the organic solvent and the extraction protocol. Clinically important microorganisms were used to assess the activity of extracts. The methanol extracts have shown good activity towards all the pathogens showing its broad spectrum nature. The activity was more prominent at acidic pH (5.5) in comparison to the neutral pH (7.0). Amikacin served as the positive control, while the solvent (methanol) served as negative control. The impact of pH on the secondary metabolite extraction from plant material has been previously established [11, 12, 2].

Among the aerial and root extracts, the root extracts were much superior to the aerial extracts. The inhibition zones of root extracts at acidic pH were in the range of 20.02 ± 0.36 mm – 12.72 ± 0.24 mm (Table 1). Both the gram negative and gram positive organisms were equally susceptible. The highest zone of inhibition was obtained against E. coli, which is known to play an important role in nosocomial infections. The efficiency of the extracts can be seen from the zones of
inhibition obtained against potent microbes like *S. aureus* (19.01 ± 0.20mm) and *S. typhimurium* (18.86 ± 0.24mm). The most resistant organism in the present study was found to be *P. aeruginosa* (13.11 ± 0.56mm). The permeability of the compounds and the resistance mechanisms displayed by the microbes could be the reason for the variable zones of inhibition exhibited by the organisms.

Earlier reports of strong antimicrobial activity of *G. sylvestre* towards pathogens like *S. aureus* and *S. typhimurium*, have confirmed the choice of the plant in the present study [13, 14, 15]. Gram negative organisms are known to be resistant to many of the drugs as well as the natural agents, as the outer membrane acts as a selective barrier for the passage of molecules in and out of the cell. However, in the present study, the gram negative bacteria have been successfully inhibited by the extracts of *G. sylvestre* owing to the high content of saponins. The antimicrobial activity of plants attributing to the presence of saponins has been established by Soetan et al., 2006 [16]. Also the sterols viz. stigmasterol present in the plant could essay an important role in the activity due their lipophylic nature that helps them in the outer membrane penetration.

**Conclusion**

The exploration of secondary metabolites from plant sources seems to be an excellent choice for the development of new age antimicrobials, given the vast biodiversity in the subcontinent. The methanol extracts of *G. sylvestre* have displayed good activity against both the gram positive and gram negative microorganisms displaying its potential in the development of new phytopharmaceuticals. As these drugs are plant based, they can be considered safe for human consumption.

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**References**


