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Molecular docking study of phytol and its derivatives against COX-2 induced inflammation: A combined density functional study

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ABSTRACT

This study aimed to determine the activity of PYT and its derivatives against COX-2, including 5KIR protein induced inflammation by using the computational tools. PYT and its derivatives have been designed by utilizing density functional theory (DFT) and the performance of the drugs was also evaluated by molecular docking study. Results suggest that the NH₂ derivative of PYT (D-NH₂) showed binding energy -6.4 (Kcal/mol) with protein 5KIR of COX-2 compared to the main drug (D) that showed binding energy -5.1 (Kcal/mol) with the same protein. HOMO and LUMO energy values were also calculated to determine the chemical reactivity of all the modified drugs. Non-covalent interactions of PYT and its derivatives were essential in improving the performance. In conclusion, D-NH₂ showed better preference in inhibiting to the protein 5KIR of COX-2 compared to other modified drugs and it can be claimed that D-NH₂ will be the best conformer for COX-2 induced inflammation.

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KEYWORDS: Phytol, 5KIR, COX-2, inflammation, molecular docking

INTRODUCTION

Phytol (PYT), a chlorophyll-derived diterpene essential oils found abundantly in nature (1). PYT is widely distributed in bacteria, especially cyanobacteria, plants and algae (2). PYTis evident in various important biological activities including anxiolytic (3), antinociceptive (4), antidiabetic (5), anti-inflammatory (6), antiatherogenic, antiteratogenic (7), anticonvulsant (8), antispasmodic (9), antimutagenic (10), anti-protozoal (11), antimicrobial (12), hypolipidemic (13), and immunoadjuvant properties (14). Cyclooxygenase (COX) catalyzes the first committed step in the synthesis of prostanoids, known as prostaglandin (PG) H synthase (15). It has been demonstrated that, through the inhibition of COX enzymatic activity, non-steroidal anti-inflammatory drugs (NSAIDs) exert their anti-inflammatory properties as well as inhibiting PG synthesis (16). COX remains central, as a unique enzyme, to the development of anti-inflammatory treatments of various pathologies, including neurodegenerative and neuroinflammatory diseases and COX produces two types of catalytic activities, firstly it catalyses PGG2 formation from arachidonic acid that is a bis-oxygenase activity (cyclooxygenase) and secondly COX reduces PGG2 to PGH2 which is a peroxidase activity (15). Two types of enzymatic activities include- external factors and interacting sites on the COX molecule can affect them (17) and COX undergoes a conformational rearrangement which gives rise to an inactive enzymatic species leading to an unstable intermediate during the cyclooxygenase activity (15). COX is involved in both enzymatic activities as an integral membrane glycoprotein which in the association of the heme group consisting of a homodimer (17). In many tissues, COX-1 is expressed as a constitutive enzyme, including the intestine and colon, whereas in macrophages, fibroblasts, and other cell types in inflammation COX-2 is expressed an inducible enzyme (18). In inflammatory reactions, COX-2 has emerged as a major player in peripheral tissues and COX-2 also functions in inflammatory and degenerative brain diseases (15). Some NSAIDs, including ibuprofen and mefenamic acid are the competitive inhibitors of COX-1 and -2 isoforms. The objective of this study is to determine the anti-inflammatory activity of PYT and its derivatives against COX-2, including 5KIR protein induced inflammation. The computational tools have been used, for this purpose, to investigate the best ligand and optimized receptor proteins as the best choice for the future scientists. The structure of PYT is modified with -OH, -F, -Cl, -OCH₂ CN and -NH₂ in C-20 position.

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COMPUTATIONAL METHODS

Optimization of Ligands by Quantum Mechanical Calculations

By using quantum mechanical (QM) methods, various types of complicated interactions between ligands and target proteins are interpreted and internal energy calculations were done (19), and all predictions were made by using Gaussian view 09 and Chem3D Pro12.0 program packages (20). The optimized structure of PYT (D) was modified with -OH, -F, -Cl, -OCH₃ CN and -NH₂ groups (Figure 1). Considering Parr and Pearson interpretation with HOMO and LUMO energy (ϵ) (21), hardness (η) and softness (S) of all drugs were also calculated from the energies of frontier HOMOs and LUMOs. Hardness (η) and softness (S) of all the drugs were calculated according to the equation: $\eta = [\epsilon LUMO - \epsilon HOMO]/2$ and S=1/ η (Table 1).

Protein Preparation

From the Protein Data Bank (PDB) database, the crystal structure of the protein (5KIR) of COX-2 of *Homo sapiens* was collected. Swiss-Pdb Viewer software package (version 4.1.0) has been utilized for the energy minimization of crystal structure and by using PyMOI (version 1.7.4.5) all the hetero atoms and water molecules of proteins (Figure 2) were removed before docking. For the analysis of docking results both the proteins and drug structures are taken into PDBQT format finally.

Docking Analysis and Binding Site

In computational drug design, molecular docking is an

Table 1: HOMO-LUMO, gap, hardness and softness

Molecules (Chair)	٤HOMO	٤LUMO	Gap	Hardness (η)	Softness (S)
Phytol (D)	-8.584	2.381	10.965	5.482	0.182
D-0H	-3.878	10.023	13.901	6.950	0.143
D-F	-4.201	10.075	14.276	7.138	0.140
D-CI	-4.640	9.996	14.636	7.318	0.136
D-OCH ₃	-6.620	9.890	16.51	8.225	0.121
D-CN	-6.460	9.504	15.964	7.982	0.125
D-NH ₂	-3.236	9.991	13.227	6.613	0.151

important tool that can predict the predominant binding mode(s) of a ligand with the target protein (22). By using CASTp the prediction of the active binding pocket of COX-2 was done and the docked pose of lowest binding free energy conformer with the respective protein was analyzed by PyMOL Molecular Graphics System (version 1.7.4.5).

RESULTS AND DISCUSSION

HOMO-LUMO, Gap, Hardness (η) and Softness (S) Analysis

Highest occupied molecular orbital (HOMO), lowest unoccupied molecular orbital (LUMO) refers frontier orbitals (FO), and the calculation of the quantity and chemical reaction in which drug molecules bind with the specific receptor have been performed by using these energy values. The structure of FO of PYT (D) and D-NH₂ were given in Figure 3. The energy gap, reveals the chemical stability and kinetic of the drug molecules, between HOMO and LUMO (20). The D-NH₂ exhibited lowest energy gap value and lowest η value and increased S value among the modified derivatives,which indicated that this drug has enhanced chemical reactivity. HOMO-LUMO, gap, η and S values were given in Table 1.

Binding Energy of the Protein-Ligands by Molecular Docking

To predict the stronger binder and virtual screen a database of compounds the docking is essential and D-NH₂ showed binding energy -6.4 (Kcal/mol) with protein 5KIR of COX-2 compared to the D (PYT) that showed binding energy -5.1 (Kcal/mol). The -OH, -F, -Cl, -OCH₃ and -CN group of PYT showed binding energy -6.3, -5.4, -5.4 and -5.7 Kcal/mol, respectively and the D-NH₂ exhibited the lowest binding energy. The binding energy of ligand-proteins were given in Table 2.

Selected Non-Covalent Interactions Among Chair Ligands D, D-NH, and COX-2 (5KIR)

The binding affinity and binding specificity were increased due to the improved hydrogen bonding (23) in D-NH₂. In the D-NH₂-5KIR complex, multiple non-bonded interactions and

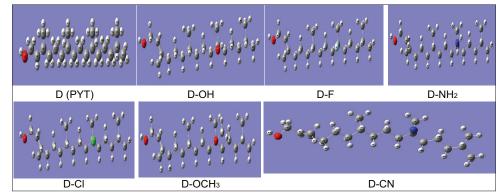


Figure 1: Modified structures of PYT (D)

docked structure were observed. A strong hydrogen bond with CYS 41 (2.67 Å) and GLN 42 (2.59 Å) were observed in D-NH₂-5KIR complex (Figure 4). For enhancing the binding affinity of D-NH₂-5KIR, the strong hydrogen bonding is considered the most significant contributing factor. Several hydrophobic bonds were observed in the D-NH₂-5KIR complex, including LYS 468 (4.52 Å), LEU 152 (5.03 Å), PRO 153 (4.69 Å), CYS 36 (4.01 Å), and TYR 130 (5.44 Å). Non-covalent interactions in D-5KIR complex were stabilized by several hydrophobic bonds, including LEU 93 (4.14 Å), VAL 116 (4.66 Å), ILE 112 (3.84 Å), VAL 89 (3.88 Å), ILE 92 (4.17 Å), and TYR 115 (4.61 Å). Selected non-covalent interactions among chair ligands D, D-NH, and COX-2 (5KIR) were given in Table 3.

Table 2: Free energy of binding values (Kcal/mol) for ligands -COX-2 (5KIR)

Ligands	Binding energy of ligands-proteins (5KIR) (Kcal/mo					
Phytol (D) (chair)	-5.1					
D-0H (chair)	-6.3					
D-F (chair)	-5.4					
D-Cl (chair)	-5.4					
D-OCH ₃ (chair)	-5.4					
D-CN (chair)	-5.7					
D-NH ₂ (chair)	-6.4					

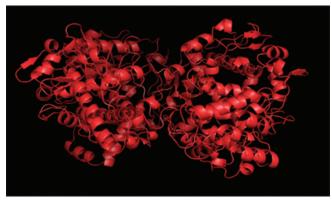


Figure 2: Crystal structure of 5KIR protein prior to docking

Pharmacokinetic Properties of PYT and Its Modified Derivatives

The modified drugs showed low acute oral toxicity, therefore, they are expected to be safe for use. The drugs will act positively, as the human intestinal absorption values of all the drugs were found positive in the bioavailability, drug metabolism and intestinal absorption (24). PYT and its modified derivatives showed weak inhibitory property for the human ether-a-go-go-related gene (hERG). AdmetSAR values of ligands were given in Table 4. Toxicity of all the compounds was predicted by PreADMET suggesting that all the compounds having a lower toxicity (Table 5).

Stoichiometry, Electronic Energy, Enthalpy, Gibb's Free Energy and Dipole Moment of PYT and Its Derivatives

The electronic energy, after modification, which indicates that the structures become more stable (Table 5). The highest Gibb's free energy is observed for D-OCH₃ and D-NH₂. On the other hand, D-Cl showed higher electronic energy and also the D-NH₂ exhibited higher electronic energy than the parent drug. The D-NH₂ showed Gibb's free energy, enthalpy, and dipole moment as 0.520518, 0.615199 1.6838, respectively; and these values would make the drug chemically more stable (Table 5).

CONCLUSION

This study showed that modified PYT drugs interact with 5KIR of COX-2 and some interesting characteristics related to free energy, dipole moment, charge distribution, and molecular orbital of the drug molecules were described by the DFT calculation. The electronic energy, enthalpy, Gibb's free energy and dipole moment of PYT and its derivatives indicate that these compounds are chemically more reactive than the main drug (PYT). The -D-NH₂ showed binding energy -6.4 (Kcal/mol) with the protein 5KIR of COX-2 compared to the main drug (D) that showed binding energy -5.1 (Kcal/mol)

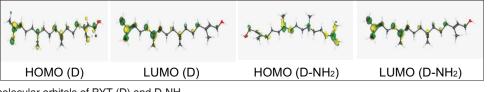


Figure 3: Frontier molecular orbitals of PYT (D) and D-NH,

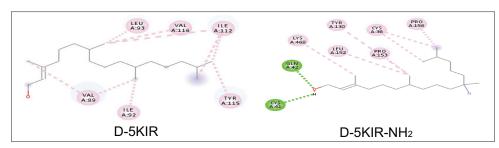


Figure 4: Binding site ligand-protein complexes

Systems	Contacts	Bond distances (Å)	Systems	Contacts	Bond distances (Å)
D-5KIR	Alkyl (LEU 93)	4.14	D-NH ₂ -5KIR	0H (CYS 41)	2.67
	Alkyl (VAL 116)	4.66	2	0H (GLN 42)	2.59
	Alkyl (ILE 112)	3.84		Alkyl (LYS 468)	4.52
	Alkyl (VAL 89)	3.88		Alkyl (LEU 152)	5.03
	Alkyl (ILE 92)	4.17		Alkyl (PRO 153)	4.69
	Pi-alkyl (TYR 115)	4.61		Alkyl (PRO 156)	3.65
	Alkyl (LEU 344)	3.41		Alkyl (CYS 36)	4.01
				Pi-alkyl (TYR 130)	5.44

Table 4: AdmetSAR values of ligands

Parameters	D	D-0H	D-F	D-CI	D-0CH ₃	D-CN	D-NH ₂
Blood Brain Barrier	0.937	0.905	0.984	0.978	0.948	0.954	0.916
Human intestinal absorption	0.984	0.990	0.992	0.991	0.969	0.968	0.984
P-glycoprotein inhibitor	0.886 (NI)	0.863 (NI)	0.839 (NI)	0.888 (NI)	0.661 (NI)	0.829 (NI)	0.893(NI)
Human Ether a-go-go-Related (hERG) Gene Inhibition	0.783 (WI)	0.892 (WI)	0.945 (WI)	0.915 (WI)	0.835 (WI)	0.848 (WI)	0.956 (WI)
Acute Oral Toxicity	0.855	0.761	0.789	0.749	0.762	0.768	0.702

Table 5: Stoichiometry, electronic energy, enthalpy, Gibb's free energy in hartree and dipole moment (Debye) of PYT and its derivatives

Name	Stoichiometry	Electronic Energy	Enthalpy (Hartee)	Gibb's free energy (Hartee)	Dipole moment (Debye)
Phytol (D)	C ₂₀ H ₄₀ O	-910.112	0.495786	0.454336	1.2354
D-0H	C ₂₀ H ₄₀ O ₂	-936.117	0.602311	0.507255	2.2188
D-F	C ₂₀ H ₃₀ FO	-960.152	0.589743	0.494326	1.4337
D-CI	C20H39CIO	-1320.513	0.588849	0.493338	1.6486
D-OCH ₃	Č, H, O,	-975.386	0.632000	0.534108	2.3188
D-CN	C , H , NO	-953.144	0.597589	0.501483	2.6719
D-NH ₂	C ₂₀ H ₄₁ NO	-916.237	0.615199	0.520518	1.6838

and the docking results revealed that D-NH₂ shows the best performance on inhibiting human COX-2 (5KIR). The nonbonding interactions, help to develop new drug which can effectively target the COX-2. Pharmacokinetic calculation predicts that all the modified drugs are non-carcinogenic. D-NH₂ for COX-2 (5KIR) will be the best conformer for 5KIRinduced inflammation in animals.

COMPETING INTEREST

None declared.

REFERENCES

- McGinty D, Letizia CS, Api AM 2010, Review fragrance material review on phytol. Food Chem Toxicol. 48(3):59-63.
- Ishibashi Y, Nagamatsu Y, Miyamoto T, Matsunaga N, Okino N, Yamaguchi K, Ito M. 2014, A novel ether-linked phytol containing digalactosyl glycero lipid in the marine green alga, *Ulva pertusa* Biochem Biophys Res Commun. 52:873-80.
- Costa JP, de Oliveira GAL, de Almeida AAC, Islam MT, de Sousa DP, Freitas RM. 2014, Anxiolytic-like effects of phytol: possible involvement of GABAergic transmission. Brain Res. 1547:34-42.
- Santos CC, Salvadori MS, Mota VG, Costa LM, de Almeida AA, de Oliveira GA, et al. 2013, Antinociceptive and antioxidant activities of phytol *in vivo* and *in vitro* models. Neurosci J. 2013:949452.
- Elmazar MM, El-Abhar HS, Schaalan MF, Farag NA. 2013, Phytol/ phytanic acid and insulin resistance: potential role of phytanic acid proven by docking simulation and modulation of biochemical alterations. PLoS One 8:45638.
- Silva RO, Sousa FB, Damasceno SR, Carvalho NS, Silva VG, Oliveira FR, et al. 2014, Phytol, a diterpene alcohol, inhibits the inflammatory response by reducing cytokine production and oxidative

stress. Fundam Clin Pharmacol. 28:455-64.

- Arnhold T, Elmazar MMA, Nau H. 2002, Prevention of vitamin A teratogenesis by phytol or phytanic acid results from reduced metabolism of retinol to the teratogenic metabolite, all-transretinoic acid. Toxicol Sci. 66:274-82. 8. Costa JP, Ferreira PB, De Sousa DP, Jordan J, Freitas RM. 2012, Anticonvulsant effect of phytol in a pilocarpine model in mice. Neurosci Lett.523:115-8.
- Pongprayoon U, Baeckstr€om P, Jacobsson U, Lindstr€om M, Bohlin L. 1992, Antispasmodic activity of beta-damascenone and E-PHY isolated from Ipomoea pes-caprae. Planta Med. 58:19-21.
- Kagoura M, Matsui C, Morohashi M. 1993, Carcinogenicity study of phytol (3,7,11,15-tetramethyl-2-hexadecen-1-ol) in ICR mice. J Investig Dermatol. 101:460.
- Bero J, Beaufay C, Hannaert V, Hérent MF, Michels PA, Quetin-Leclercq J. 2013, Antitrypanosomal compounds from the essential oil and extracts of *Keetia leucantha* leaves with inhibitor activity on *Trypanosoma brucei* glyceraldehyde-3-phosphate dehydrogenase. Phytomedicine. 20:270-4.
- 12. Inoue Y, Hada T, Shiraishi A, Hirose K, Hamashima H, et al. 2005, Biphasic effects of geranylgeraniol, teprenone, and phytol onthe growth of *Staphylococcus aureus*. Antimicrob Chemother Agents Chemother. 49:1770-4. 13. Takahashi N, Kawada T, Goto T, Yamamoto T, Taimatsu A, Matsui N, et al. 2002, Dual action of isoprenols from herbal medicines on both PPARg and PPARa in 313-L1 adioocvtes and HepG2 hepatocvtes. FEBS Lett. 514:315-22.
- Lim SY, Bauermeister A, Kjonaas RA, Ghosh SK. 2006, Phytolbased novel adjuvants in vaccine formulation: 2. Assessment of efficacy in the induction of protective immune responses to lethal bacterial infections in mice. J Immune Based Ther Vacc.4:5.
- Minghetti L. 2004, Cyclooxygenase-2 (COX-2) in inflammatory and degenerative brain diseases. J Neuropathol Exp Neurol. 63:901-10.
- Vane JR. 1971, Inhibition of prostaglandin synthesis as a mechanism of action for aspirin-like drugs. Nat New Biol. 231:232-5.
- Smith WL, Garavito RM, DeWitt DL. 1996, Prostaglandin endoperoxide H synthases (cyclooxygenases)-1 and -2. J Biol Chem. 271:33157-60.
- 18. Eberhart CE, Dubois RN. 1995, Eicosanoids and the gastrointestinal

tract. Gastroenterology. 109:285-301.

- Gleeson MP, Gleeson D. 2009, QM/MM calculations in drug discovery: a useful method for studying binding phenomena? J Chem Information Modeling. 49:670-7.
- Rahman A, Ali MT, Shawan MMAK, Sarwar MG, Khan MA, Halim MA. 2016, Halogen-directed drug design for Alzheimer's disease: a combined density functional and molecular docking study. SpringerPlus. 5:1346. 21. Parr RG, Yang W. 1989, Density-Functional Theory of Atoms and Molecules (New York: Oxford

University Press).

- Morris GM, Lim-Wilby M. 2008, Molecular docking. Methods Mol Biol. 443:365-82.
- Bissantz C, Kuhn B, Stahl M. 2010, A medicinal chemist's guide to molecular interactions. J Med Chem. 53:5061-84.
- Rehman S, Nabi B, Fazil M, Khan S, Bari NK, Singh R, Ali J. Role of P-2017, Glycoprotein Inhibitors in the Bioavailability Enhancement of Solid Dispersion of Darunavir. BioMed Res Int. doi: 10.1155/2017/8274927.