

IN SILICO PREDICTION OF 3D STRUCTURE OF PECTATELYASE FROM

FUSARIUM OXYSPORUM F. SP. LYCOPERSICI

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Introduction

MICROBIOLOGY

Fusariumoxysporum f. sp. lycopersici, is a soil borne plant pathogenic fungus, causes Fusarium wilt specifically in tomato. This disease is of worldwide importance and is particularly severe in countries with warm climate (1). The fungus enters the host roots directly through penetration using hyphae and colonize the cortex by intracellular and intercellular growth. Once it reaches the vascular tissue, the pathogen spreads rapidly upward through the xylem vessels, provoking the characteristic wilt symptoms (2). During root penetration and host plant colonization, F. oxysporumsecretes an array of enzymes such as pectatelyase, polygalacturonases and xylanases that may contribute to the degradation of the structural barriers constituted by plant cell walls (3, 4, 5, 6, 7, 8). Among these enzymes, Pectatelyase (PL) is most important in breaking the cell wall of host plant and brings about maceration of parenchymatous tissue (9).

Pectatelyases (PL, EC 4.2.2.2), otherwise known as pectatetranseliminases, catalyse the eliminative cleavage of de-esterified pectin, which is a major component of primary cell walls of many higher plants (10). The backbone of pectic polysaccharides is built from blocks of β -1,4 linked polygalactosyluronic acid residues interspersed with regions of alternating galactosyluronic acid and rhamnosyl residues (11). Cleavage of pectin by PL generates oligosaccharides with unsaturated galacturonosyl residues at their nonreducing ends.

The structural details of the enzyme are helpful in developing efficient inhibitors. Even though *F. oxysporum* is a dangerous plant pathogen and PL is one of the potential weapons used by this pathogen to invade the plant system, the structural details of PL are not available in www.rcsb.org.In the absence of experimental data on the structure of PL1 of *F. oxysporum* f.sp.*lycopersici*, homology modeling approach with its ability to derive its reasonable 3D structure based on sequence identity among various proteins of same class, offers reasonable alternative. By considering these points this work was conducted to

develop the 3D structure of pectatelyase from *Fusariumoxysporum* f. sp. *Lycopersici*.

Materials and Methods

Sequence search and Analysis

Complete sequence information of PL enzyme (GenBank: AAC64368.1) of Fusariumoxysporum f. sp. lycopersiciretrieved from NCBI (12) was submitted to PSI blast tool (http://www.ebi.ac.uk/Tools/blastpgp/). PSI-BLAST is a tool that produces a position-specific scoring matrix constructed from a multiple alignment of top scoring BLAST responses to a given guery sequence. The degree of similarity is given in terms of a scoring parameter called the E-value (13). The conserved amino acid residues in PL protein and in the selected PDB templates were identified by submitting the sequence information to CLUSTAL W (14), a general purpose multiple sequence alignment program for DNA or proteins. All parameters were set at default values. This program produces biologically meaningful multiple sequence alignments of divergent sequences. It calculates the best match for the selected sequence and lines them up so that the identities, similarities and differences can be seen.

Homology modeling, Structure prediction and External Validation

Based on these data homology modeling was done using SWISS-MODEL homology-modeling server (15) (http://swissmodel.expasy.org/workspace).The predicted model for PL 1 of *F. oxysporum* f. sp. *lycopersici* was evaluated using additional structure assessment tools like PROCHECK (16) (Ramachandran plot analysis and G-value), Verify 3D (17), and WHAT_CHECK (18).

Results and Discussion

In the past, several studies have been conducted to understand the 3D structure and function of many proteins from *Fusarium* (19-21).However, 3D structure of PL from *F. oxysporum* f. sp. *lycopersici*, which is considered to be one of the potential weapons used by

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this pathogen to invade the plant system, has not been determined experimentally. In the present study the authors have attempted to predict the structure of PL. The results pertaining to the tertiary structure prediction of PL enzyme (GenBank: AAC64368.1) of *F. oxysporum* f. sp. *lycopersici* is presented and discussed in this chapter.

Template search using PSI blast

As the model accuracy is expected to increase with the use of multiple templates (22), the present study focused on generating PL models using several PDB template combinations. One of the main advantages of comparative modeling program is that it can combine information from multiple template structures in two ways. In the first way multiple template structures may be aligned with different domains of the target with little overlap between them wherein the modeling procedure can construct a homology based model of the whole target sequence. In the second waySecond the template structures may be aligned with same part of the targetand themodeling procedure is likely to automatically select the best template (sali). The search for potential templates of PL enzyme (GenBank: AAC64368.1) from *Fusariumoxysporum* f. sp. *lycopersici*using PSI blast tool revealed many PDB templates sharing similar sequence homology. Three of these templates having high sequence identity and better E valuesthan threshold were selected for generating 3D model (Table 1).

Sequence alignment and Model Building

Multiplesequence alignment of PL using CLUSTAL W tool revealed that Leu⁴⁴, Ala⁴⁸, Asn⁵², Ile⁵⁵, Ala⁵⁷, Asn⁷⁵, Val⁷⁶, Trp⁷⁸, Asp⁸⁰, Glu⁸³, Asp⁸⁴, Ala⁸⁵, Thr⁸⁷, Cys⁷¹, T hr72,Cys66,His65,Ala63,Ala57,Ile55,Asn52,Ala48,Leu44,Asn11 ¹,Gln¹⁰⁹,Lys¹⁰⁶,Asp⁹⁸,Ala⁹⁶,Ala⁹³,Gly⁹²,Gly⁹¹,Ile⁸⁹,Thr⁸⁶, Gly¹⁶⁹, Gly¹³⁵, Arg¹³², Lys¹³⁰ and Gly¹²⁹ were completely conserved in all the three PDB templates...... 3D models of PL enzyme (GenBank: AAC64368.1) of Fusariumoxysporum f. sp. lycopersiciwhich were first obtained using different multiple template combinations from the selected three templates. However, only the template 3B4N A was chosen for model building as it had the lowest E value (2e-7)(Table-1).

Fig.1: a. Clustal W analysis of pectatelyase sequence with other related sequence; b. Homology modeledpectatelyase from Fusariumoxysporum f. sp. Lycopersici

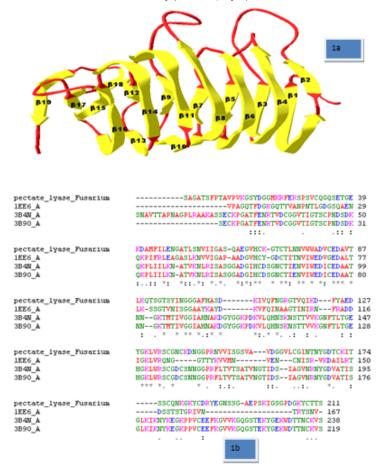


Table 1: Summary of the template sequence profile that was generated at the end of 20th iteration of PSI blast analysis using Pectatelyase (AAC64368) of *Fusariumoxysporum* f. sp. *lycopersici*as query. Threshold PSI blast E-value = 0.001

Sequences with patterr	n at position and E-value BETTER	than threshold	
Template	Score	E- value	
pdb 3B4N A	261	2e-70	
pdb 3B90 A	288	2e-60	
pdb 1EE6 A	206	5e-54	
Sequences with E-value	WORSE than threshold		
pdb 2OJU A	30.1	0.80	
pdb 1WCQ A	27.8	3.8	
pdb 1EUT A	27.8	3.8	
pdb 1W8N A	27.8	4.1	
pdb 1QQE A	27.4	4.2	
pdb 2BER A	27.4	4.5	
pdb 2QE7 D	27.4	5.3	
pdb/1EUR/A	27.4	5.3	
pdb 2BZD A	27.0	5.8	
pdb 1EGZ A	27.0	6.1	

External validation of tertiary structure

Validation of the predicted 3D neurotoxin structures (after loop refinement) by PROCHECK analysis showed that 73.8 % of the residues of PL model were present in the most favoured region followed by 18.6 % in the allowed region, 4.7 % in generously allowed region and 2.9 % in disallowed plot. region of Ramachandran However, Ramachandran plot analysis of the PDB template, 3B4N A showed that 82.9 % of the residues were present in the most favored region followed by 15.5 % of residues in additional disallowed region, 1.6% in generously allowed region and 0.0 % in disallowed region of Ramachandran plot.

References

- Jones, J.P., and Woltz, S.S. 1981. Fusariumincited diseases of tomato and potato and their control. Pages 157-168 in Fusarium: Diseases, Biology, and Taxonomy. P.E. Nelson, T.A. Toussoun, and R.J. Cook, eds. Pennsylvania State University Press, University Park
- Beckman, C.H. 1987. The nature of wilt diseases of plants. Am. Phytopathology. Soc. Press, St. Paul, MN, USA.
- Di Pietro, A., and Roncero, M. I. G. 1996a.Endopolygalacturonase from *Fusarium* oxysporum f. sp. *lycopersici:* Purification, characterization, and production during infection of tomato plants. Phytopathology 86: 1324-1330.
- Di Pietro, A., and Roncero, M. I. G. 1996b. Purification and characterization of an exopolygalacturonase from the tomato vascular wilt pathogen *Fusarium oxysporum* f. sp. *lycopersici*. FEMS Microbiol.Lett. 145:295-299.

- Di Pietro, A., and Roncero, M. I. G. 1998. Cloning, expression, and role in pathogenicity of *pg1* encoding the major extracellular endopolygalacturonase of the vascular wilt pathogen *Fusarium oxysporum*. Mol. Plant-Microbe Interact. 11:91-98.
- García-Maceira, F. I., A. Di Pietro, and M. I. G. Roncero. 2000. Cloning and disruption of *pgx4* encoding an in planta expressed exo polygalacturonase from *Fusarium oxysporum*. Mol. Plant-Microbe Interact. 13:359-365.
- Huertas-González, M. D., Ruiz-Roldán, M. C., García-Maceira, F. I., Roncero, M. I. G., and Di Pietro, A. 1999. Cloning and characterization of *pl1* encoding an in planta-secreted pectatelyase of *Fusarium oxysporum*. Curr.Genet. 35:36-40.
- Ruiz Roldán, M. C., A. Di Pietro, M. D. Huertas-González, and M. I. G. Roncero. 1999. Two xylanase genes of the vascular wilt pathogen *Fusarium oxysporum* are differentially expressed during infection of tomato plants. Mol. Gen. Genet. 261:530-536.
- Pérombelon M.C.M., Kelman A. (1980): Ecology of the soft rot erwinias. Annual Review of Phytopathology, 18: 361–387.
- Carpita NC, Gibeaut DM. 1993. Structural models of primary cell walls in flowering plants: consistency of molecular structure with the physical properties of the walls during growth. *The Plant Journal* 3, 1–30.[Medline]
- Willats WGT, Orfila C, Limberg G, Buchholt HC, Van Alebeek G-JWM, Voragen AGJ,Marcus SE, Christensen TMIE, Mikkelsen JD, Murray BS, Knox JP (2001)Modulation of the degree and pattern of methyl-esterification of pectic homo galacturonanin plant cell walls - Implications for

pectin methyl esterase action, matrix properties, andcell adhesion. J BiolChem276: 19404-19413

- 12. http://www.ncbi.nlm.nih.gov/
- Altschul, S. F., Madden, T. L., Schaffer, A. A., Zhang, J., Zhang, Z., Miller, W., and Lipman, Gapped BLAST and PSI-BLAST: a new generation of protein database search programs, Nucleic Acids Research, 1997, Vol. 25, No. 17 3389-3402
- Thompson, J. D., Higgins, D. G., and Gibson, T. J. 1994 CLUSTAL W: improving the sensitivity of progressive multiple sequence alignment through sequence weighting, position-specific gap penalties and weight matrix choice. *Nuc Acids Res.* 22, 4673-4680.
- 15. Tovchigrechko, A. and Vakser, I. A. 2006. GRAMM-X public web server for protein-protein docking..*Nucleic Acids Research 34.* Web Server issue W310- 314.
- Laskowski, R. A., McArthur, M. W., Moss, M. D., and Thornton, J. M., 1998. "PROCHECK: a program to check the stereochemical quality of protein structures." *JApplCryst2* :283-291.

- Luthy, R., Bowie, J. U., Eisenberg, D. 1992. Assessment of protein models with threedimensional profiles Nature 356: 83-85.
- Hooft, R. W., Vreind, G., Sander, C., and Abola, E.
 E. 1996. Errors in protein structure *Nature 381*, 272-272.
- D'Ovidio, R., Mattei, B., Roberti, S., and Bellincampi, D. 2004.Polygalacturonases, polygalacturonase-inhibiting proteins and pecticoligomers in plant-pathogen interactions. iochim. Biophys. Acta 1696,237-244.
- André-Leroux G, Tessier D. and Bonnin E. (2005).Action pattern of *Fusarium moniliforme* endopolygalacturonase towards pectin fragments: Comprehension and prediction. BiochimBiophysActa 1749, 53-64.
- Cooper R.M. and Wood R.K.S. (1975) Regulation of synthesis of cell wall-degrading enzymes by *Verticilliumalbo-atrum* and *Fusarium oxysporum* f. sp. lycopersici.Physiol Plant Pathol. 5, 135-156.
- Fiser, A. and Sali, A. *Methods in Enzymology*, pp. 374, 463-493. Eds., Carter, C. W. andSweet, R. M. Academic Press, San Diego (2003).