

Diversity, activity, antibiotic and heavy metal resistance of bacteria from petroleum hydrocarbon contaminated soils located in different sites

Jai Godheja, Krishna Dixit and Payal Sharma

Department of Biotechnology, RITEE Raipur, India.

Abstract

The main aim behind this project was to obtain a collection of hydrocarbonoclastic bacteria that might be suitable for bioremediation processes as soil inoculums to remove hydrocarbons in presence of heavy metals. To achieve this soils from different sites have been selected like Hindustan Petroleum of RAIPUR, BILASPUR & BACHELI strongly contaminated solely with hydrocarbons and for comparison non-contaminated soil of RAIPUR. To reveal affect of pollutants on endogenous micro biota and the bacterial shifts resulting as a consequence of different types of pollution. Different molecular biological and microbiological techniques were used. Individual samples were tested:

1. For their ability of degrading different types of hydrocarbons (aliphatic, aromatic & polycyclic aromatic hydrocarbons)
2. For their capability of resisting heavy metals (Cu^{2+} , Zn^{2+} , Pb^{2+})
3. If they proliferate in presence of antibiotics.

Results that were observed was increased activity of hydro carbon clastic bacteria due to the significant CO_2 production and elevated hydrocarbon degrading bacterial counts. And the bacterial communities that are diversified in number of species were influenced by the pollutants present in it was tested by the T-RFLP. Strains of highest antibiotic resistance and heavy metal tolerance were isolated from the sample. Moreover, some test indicated significant correlation between heavy metal tolerance/antibiotic resistance and antibiotic resistance/hydrocarbon degradation ability of the isolates.

Keywords: Polyaromatic Hydrocarbon, Benzene, Toluene, Anthracene.

INTRODUCTION

Soil contamination may take place due to production, refinery, transportation or storage of crude oil or their derivatives by incidents like accidental leakages or such like that with the BTEX compounds (benzene, toluene, ethyl-benzene, & xylene), poly aromatic hydrocarbons (i.e. PAHs). The main sources of contamination are observed to be the oil wells, petroleum plants etc. since trace heavy metals are common constituents of crude oil, petroleum derivatives (Pb^{2+} , leaded gasoline, lubricating oils or greases, Zn^{2+} , Cd^{2+}). The area with an increased long term hydrocarbon polluted, heavy metal contaminations of them are chosen. These compounds in the environment leads to serious health risks, due to carcinogenic and mutagenic effects. Because antibiotic resistant & heavy metal resistant genes are to be found in some mobile genetic element, metal pollution often promotes antibiotic resistance emergence in exposed organisms that also has a growing concern in natural and clinical settings. Thus, remediation of these areas was the great interest.

Multiple treatment methods have been applied. Among them, physical and chemical approaches eliminated a broad spectrum of contaminants, with a drawback of high energy consumption and need of additional chemicals. With physico-chemical treatment, for

example incineration, pollutants may transfer from one environmental compartment to another. As a result, BIO-Remediation was found to be cost-effective, applicable in large fields. In this method, metabolic activity of microorganisms leads to complete breakdown of organic compounds into non-toxic compounds potentially ending in their mineralization. It's a time consuming process and its success depends on many factors such as pH, temperature, availability of O_2 , nutrients. For evaluation of the petroleum contaminated hydrocarbon environment, if a microbial community of the contaminated environment has a metabolic potential to remove the contamination.

MATERIALS AND METHOD

Site description and soil sampling

Soils samples were collected from four different places. They were Hindustan petroleum of Raipur, Bachel, Bilaspur & for comparing non contaminated soil from Raipur as a Control. Among all these places, Raipur is in $22^{\circ}33'N$ to $21^{\circ}14'N$ & $82^{\circ}6'E$ to $81^{\circ}38'E$ in centre of Chhattisgarh. Bilaspur is located in the eastern part of Chhattisgarh, situated within the latitude of $21^{\circ}47'$ to $23^{\circ}8'$ & $81^{\circ}14'$ to $83^{\circ}15'$ longitude and the last Bachel, within latitude of $18^{\circ}42'13.36''$ & longitude of $81^{\circ}14'17.65''$. Hydrocarbon contaminated soil samples were collected in vicinity of above ground or underground. Samples as a negative control is been taken from non-contaminated areas. The top 1 cm of soil was collected using sterile spatula into sterile flasks with cotton plugs for the microbiological analysis.

*Corresponding Author

Gram Staining

This method is also known as differential staining. It was invented in 1884 by Danish scientist Hans Christian Gram. Gram Staining involves basic steps: In this bacteria are stained with Crystal violet. It imparts purple color. Some of the bacteria gain Purple color and some remain unstained. Then it is treated with grams iodine solution. Iodine is used as a mordant. Mordant is a substances which increases it efficacy of stains towards biological specimens. A decolorizer is used to remove excess of stain. Here a mixture of alcohol and acetone is used, to remove the extra stain. This is a differential step. Then another, i.e. secondary stain is added Saffranin. Those bacteriae which are remain unstained will get stained by the secondary stain. Bacteria that retain color are classified a gram's positive and that loses color are classified as gram's negative. On the basis of Gram staining and their respective shapes some plates were selected

Table 1.

S.No.	Serial Dilution	Contaminated Site
1	10 ⁻²	RNCS (Raipur Non Contaminated Site)
2	10 ⁻⁴	RNCS
3	10 ⁻²	BICS (Bilaspur Contaminated Site)
4	10 ⁻⁷	BICS
5	10 ⁻⁴	BCS (Bacheli Contaminated Site)
6	10 ⁻⁵	BCS
7	10 ⁻²	RCS (Raipur contaminated Site)
8	10 ⁻²	RCS

Enrichment purification and culturing of hydrocarbon degrading bacteria

All the microorganisms isolated from various soil samples of different petroleum sites. For this, identification techniques were applied like Gram staining & biochemical tests like :

- Catalase test
- Urease test
- Hydrolysis of gelatin test
- Cellulase production test
- Hydrogen sulfide test
- Starch hydrolysis test
- Carbohydrate fermentation test

- Imvic tests:
 - Indole test
 - Methyl red & Voges proskaeur test
 - Citrate utilization test
- Casein hydrolysis test.

Hydrocarbon degradation of potential isolates

Identified strains were treated with hydrocarbons like Benzene and Toluene. There were two methods applied for degradation of these hydrocarbons. They are:

Dye Method: Microbes have been cultured in BBH media along with filter sterilized benzene / toluene and a indicator dye resazurine. Resazurine is used as a redox indicator. Test media contained 50ml BBH mineral broth supplemented with 0.2µm filtered sterilized of one of the hydrocarbon and resazurine indicator i.e. 10mg/l. For selecting a strain which posses increase hydrocarbon degradation potential the concentration of applied carbon source was set to 0.5g/l. Test solutions were inoculated with 250µl strain cultured solutions prepared from respective plates. Reading were taken at (OD₆₀₀= 0.5nm). Initial hydrocarbon degradation changes the blue color of test solution to colorless via pink. The flasks contains the test solution were incubated for a week in rotary shaker at 145 rpm and 28°C. Samples which shows no degradation activity (blue color) were marked "-", minimum microbial activity (bluish pink color) "+", the medium activity pink samples by "++", while some samples showing increased hydrocarbon degradation activity (colorless) were marked "+++".

Turbidity Method: In this, test solution is prepared without addition of indicator. It is incubated for a 4four weeks and reading were taken at (OD₆₀₀= 0.5nm).

RESULTS

Enrichment purification and culturing of hydrocarbon degrading bacteria

Biochemical tests were done for the identification of microbes present in contaminated soils. From these we come to know that in some of the soil samples *pseudomonas* species were present and in some of the contaminated soils *Enterobacter aerogenes*.

PROPERTIES	RNCS 10 ⁻⁴	RNCS 10 ⁻³	BICS 10 ⁻⁵	BICS 10 ⁻⁷	BCS 10 ⁻⁵	BCS 10 ⁻⁴	RCS 10 ⁻³	RCS 10 ⁻⁶
Gram stain	+	-	+	-	-	+	+	-
Color	white	white	White	White	white	White	white	white
Shape	cocci	cocci	Rod	Rod	cocci	Cocci	cocci	cocci
Starch hydrolysis	+	+	+	+	+	+	+	+
Urease	+	-	-	-	+	-	+	-
Indole	-	-	+	-	+	-	+	+
Methyl red	-	+	-	-	-	+	-	-
Voges Proskauer	-	-	-	-	-	-	-	-
Citrate	+	+	+	+	+	+	+	+
Catalase	+	+	+	+	+	+	+	+
Hydrogen Sulphide	-	-	-	-	-	-	-	-
Cellulase	+	+	+	+	+	+	+	+
Gelatinase	+	+	+	+	+	+	+	+
Glucose	-	+	+	-	-	-	-	-
Sucrose	+	-	-	-	+	-	-	-
Lactose	-	-	-	+	+	-	-	-
Caseinase	-	-	-	-	-	-	-	-

Hydrocarbon degradation of potential isolates

Two methods of checking biodegradation have been employed:

Dye method: Microbes have been cultured in BBH media along with filter sterilized benzene / toluene and an indicator dye resazurine. The color of the dye is observed as it changes from purple to colorless via pink. Eight selected colonies in duplicate have been kept on Oct 15th, 2013 in shaking incubator at 140 rpm. One sample

from Bacheli (10^{-4}) has shown the pink color as on Oct 21st, 2013 and another from Bacheli 10^{-5} shown pink color on 1st Nov, 2013. Rest of the samples is still kept in shaking incubator for further observation.

Turbidity method: All the eight samples in duplicates also have been kept in BBH media along with filter sterilized benzene / toluene at 35°C and turbidity was measured using spectrophotometer at regular intervals. Most of the samples have shown an increase in turbidity.

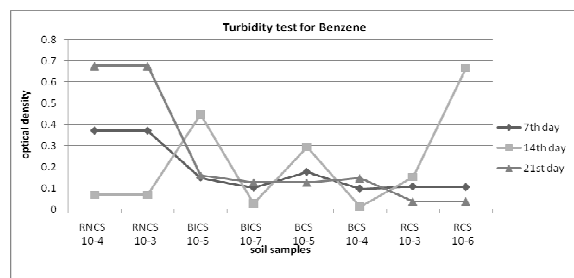
Observation table for Benzene

Soil samples	7 th day	14 th day	21 st day
RNCS 10^{-3}	0.032	0.210	0.474
RNCS 10^{-4}	0.348	0.518	0.552
BICS 10^{-5}	0.530	0.198	0.360
BICS 10^{-7}	0.494	0.405	0.799
BCS 10^{-4}	0.328	0.511	0.414
BCS 10^{-5}	0.330	0.400	0.450
RCS 10^{-3}	0.512	0.530	0.540
RCS 10^{-6}	0.420	0.877	0.563

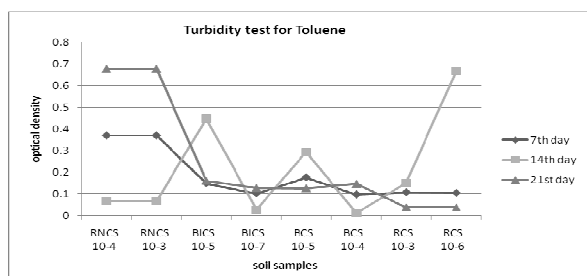
Observation table for Toluene

Soil samples	7 th day	14 th day	21 st day
RNCS 10^{-3}	0.371	0.068	0.678
RNCS 10^{-4}	0.090	0.044	0.167
BICS 10^{-5}	0.150	0.448	0.162
BICS 10^{-7}	0.103	0.028	0.129
BCS 10^{-4}	0.098	0.012	0.148
BCS 10^{-5}	0.177	0.294	0.300
RCS 10^{-3}	0.108	0.152	0.390
RCS 10^{-6}	0.105	0.668	0.581

This shows the increase in turbidity in mid of month and then it declines this just because of the life cycle of bacteria. From the above observation we come to know that bacteriae present in contaminated soil are more potent in degradation of benzene as compared to the toluene.



line Chart for benzene



Line Chart for Toluene

CONCLUSION

Majority of isolated strains were able to successfully degrade the aromatic hydrocarbons. The further degradations of poly aromatic hydrocarbon will be studied as future aspects. The isolated strains were related to pseudomonas species. According to various biochemical test conducted we come to inference that it might be *Enterobacter aerogenes*. In hydrocarbon degradation, it was find that microbes were more potent in degradation of Benzene as compared to the Toluene. One sample from Bacheli (10^{-4}) having benzene have shown the pink color as on Oct 21st, 2013 and another of Bacheli 10^{-4} turn pink on 1st Nov, 2013. From this it was concluded that, according to dye method the microbial strains present in that has partially degraded the aromatic hydrocarbon.

REFERENCES

- [1] Abdullah, M.Z.B., Saat, A.B., Hamzah, Z.B., 2011. Assessment of the impact of petroleum and petrochemical industries to the surrounding areas in Malaysia using mosses as bioindicator supported by multivariate analysis. Environmental Monitoring and Assessment. <http://dx.doi.org/10.1007/s10661-011-2236-y>.
- [2] Altschul, S.F., Madden, T.L., Schäffer, A.A., Zhang, J., Zhang, Z., Miller, W., Lipman, D.J., 1997. Gapped BLAST and PSI-BLAST: a new generation of protein database search programs. Nucleic Acids Research 25, 3389-3402.
- [3] Amáñile-Cuevas, C.F., Piña-Zentella, R., Wah-Laborde, M.E., 1991. Decreased resistance to antibiotics and plasmid loss in

- plasmid-carrying strains of *Staphylococcus aureus* treated with ascorbic acid. *Mutation Research* 264, 119-125.
- [4] Bachoon, D.S., Hodson, R.E., Araujo, R., 2001. Microbial community assessment in oil-impacted salt marsh sediment microcosms by traditional and nucleic acidbased indices. *Journal of Microbiological Methods* 46, 37-49.
 - [5] Baker-Austin, C., Wright, M.S., Stepanauskas, R., 2006. Co-selection of antibiotic and metal resistance. *Trends in Microbiology* 14, 176-182.
 - [6] Balba, M.T., Al-Awadhi, N., Al-Daher, R., 1998. Bioremediation of oil-contaminated soil: microbiological methods for feasibility assessment and field evaluation. *Journal of Microbiological Methods* 32, 155-164.
 - [7] Bartha, R., Pramer, D., 1965. Features of a flask and method for measuring the persistence and biological effects of pesticides in soil. *Soil Science* 100, 68-70.
 - [8] Bauer, A.W., Kirby, W.M.M., Sherris, J.C., Turck, M., 1996. Antibiotic susceptibility testing by a standardized single disc method. *American Journal of Clinical Pathology* 45, 493-496.
 - [9] Ben Said, O., Goñi-Urriza, M.S., El Bour, M., Dellali, M., Aissa, P., Duran, R., 2008. Characterization of aerobic polycyclic aromatic hydrocarbon-degrading bacteria from Bizerte lagoon sediments, Tunisia. *Journal of Applied Microbiology* 104, 987-997.
 - [10] Bossert, I.D., Kosson, D.S., 1997. Methods for measuring hydrocarbon biodegradation in soils. In: Hurst, C.J., Knudsen, G.R., McInerney, M.J., Stetzenbach, L.D., Walter, M.V. (Eds.), *Manual of environmental microbiology*. ASM Press, Washington, pp. 738-745.
 - [11] Braddock, J.F., Catterall, P.H., 1999. A simple method for enumerating gasoline and diesel-degrading microorganisms. *Bioremediation Journal* 3, 81-84.
 - [12] Budo, G.H., Berca, M., Penescu, A., Gavrilut, a, I., Soare, M., Dana, D., Birescu, L., 2003. Global agrochemical indexes of soil fertility. In: Mitchell, N., Zibold, G. (Eds.), *Proceedings of the XXXI annual meeting of ESNA, Crete*, pp. 57-63.
 - [13] Cánovas, D., Cases, I., de Lorenzo, V., 2003. Heavy metal tolerance and metal homeostasis in *Pseudomonas putida* as revealed by complete genome analysis. *Environmental Microbiology* 5, 1242-1256.
 - [14] Cejkova, A., Masák, J., Jirk_u, V., Veselý, M., Pátek, M., Ne_svera, J., 2005. Potential of *Rhodococcus erythropolis* as a bioremediation organism. *World Journal of Microbiology and Biotechnology* 21, 317-321.
 - [15] Chen, X., Shi, J., Chen, Y., Xu, X., Xu, S., Wang, Y., 2006. Tolerance and biosorption of copper and zinc by *Pseudomonas putida* C21 isolated from metal-polluted soil. *Canadian Journal*.
 - [16] Chun, J., Lee, J.H., Jung, Y., Kim, M., Kim, S., Kim, B.K., Lim, Y.W., 2007. EzTaxon: a web-based tool for the identification of prokaryotes based on 16S ribosomal RNA gene sequences. *International Journal of Systematic and Evolutionary Microbiology* 57, 2259-2261.
 - [17] CLSI, 2007. Performance Standards for Antimicrobial Susceptibility Testing. CLSI Approved Standard M100-S17. Clinical and Laboratory Standards Institute, Wayne, PA.
 - [18] Damian, F., Damian, G., L_ac_atus, u, R., Iepure, G., 2008. Heavy metals concentration of the soils around Zlatna and Cops, aMic_a smelters Romania. *Carpathian Journal of Earth and Environmental Sciences* 3, 65-82.
 - [19] Devulder, G., Perriere, G., Baty, F., Flandrois, J.P., 2003. BIBI, a bioinformatics bacterial identification tool. *Journal of Clinical Microbiology* 41, 1785-1787.
 - [20] DIN 38407-9, 1991. German Standard Methods for the Examination of Water, WasteWater and Sludge; Substance Group Analysis (Group F); Determination of Benzene and Some of Its Derivatives by Gas Chromatography (F9).
 - [21] George-Okafor, U., Tasie, F., Muotoe-Okafor, F., 2009. Hydrocarbon degradation potentials of indigenous fungal isolates from petroleum contaminated soils. *Journal of Physical and Natural Sciences* 3, 1-6.
 - [22] Goslee, S.C., Urban, D.L., 2007. The ecodist package for dissimilarity-based analysis of ecological data. *Journal of Statistical Software* 22, 1-19.
 - [23] Haines, J.R., Wrenn, B.A., Holder, E.I., Strohmeier, K.L., Herrington, R.T., Venosa, A.D., 1996. Measurement of hydrocarbon-degrading microbial populations by a 96-well plate most-probable-number procedure. *Journal of Industrial Microbiology* 16, 36-41.
 - [24] Hammer, Ø, Harper, D.A.T., Ryan, P.D., 2001. PAST: Paleontological statistics software package for education and data analysis. *Palaeontologia Electronica* 4, 1-9.
 - [25] Idise, O.E., Ameh, J.B., Yakubu, S.E., Okuofu, C.A., 2010. Modification of *Bacillus cereus* and *Pseudomonas aeruginosa* isolated from a petroleum refining effluent for increased petroleum product degradation. *African Journal of Biotechnology* 9, 3303-3307.
 - [26] John, N.M., 2007. Heavy metals content of crude oil sludge/poultry manure and crude oil sludge/municipal solid waste composts. *Agricultural Journal* 2, 281-284.
 - [27] Kaplan, C.W., Kitts, C., 2004. Bacterial succession in a petroleum land treatment unit. *Applied Environmental Microbiology* 70, 1777-1786.
 - [28] Knapp, C.W., McCluskey, S.M., Singh, B.K., Campbell, C.D., Hudson, G., Graham, D.W., 2011. Antibiotic resistance gene abundances correlate with metal and geochemical conditions in archived Scottish soils. *PLoS One*. <http://dx.doi.org/10.1371/journal.pone.0027300>.
 - [29] Lane, D.J., 1991. 16S/23S rRNA sequencing. In: Stackebrandt, E., Goodfellow, M. (Eds.), *Nucleic acids techniques in bacterial systematic*. Wiley, New York, pp. 115-175.
 - [30] Liu, W.T., Marsh, T.L., Cheng, H., Forney, L.J., 1997. Characterization of microbial diversity by determining terminal restriction fragment length polymorphism of genes encoding - 16S rRNA. *Applied Environmental Microbiology* 63, 4516-4522.
 - [31] Lacatus, u, R., Lacatus, u, A.R., 2008. Vegetable and fruits quality within heavy metals polluted areas Romania. *Carpathian Journal of Earth and Environmental Sciences* 3, 115-129.
 - [32] L_az_aroaie, M.M., 2010. Multiple responses of Gram-positive and Gram-negative bacteria to mixture of hydrocarbons. *Brazilian Journal of Microbiology* 41, 649-667.
 - [33] Marqués, S., Ramos, J.L., 1993. Transcriptional control of the *Pseudomonas putida* TOL plasmid catabolic pathways. *Molecular Microbiology* 9, 923-929.
 - [34] Massol-Deya, A.A., Odelson, D.A., Hickey, R.F., Tiedje, J.M., 1995. Bacterial community fingerprinting of amplified 16S and 16-23S ribosomal DNA gene sequences and restriction endonuclease analysis (ARDRA). In: Akkermans, A.D.L., van Elsas, J.D., de Bruijn, F. (Eds.), *Molecular microbial ecology*

- manual. Kluwer Academic Press, Dordrecht, The Netherlands, pp. 1-8.
- [35] Mishra, S., Dwivedi, S.P., Singh, R.B., 2010. A review on epigenetic effect of heavy metal carcinogens on human health. *The Open Nutraceuticals Journal* 3, 188-193.
- [36] Mrozik, A., Piotrowska-Seget, Z., 2009. Bioaugmentation as a strategy for cleaning up of soils contaminated with aromatic compounds. *Microbiological Research* 165, 363-375.
- [37] MSZ 21470-84, 2002. Environmental Protection. Soil Analysis. Part 84. Determination of Polycyclic Aromatic Hydrocarbon (PAH) Content. Gas Chromatographic mass Spectrometric Method. Hungarian Standard Institution, Budapest.
- [38] MSZ 21470-92, 1997. Environmental Protection. Soil Analysis. Determination of Volatile Aromatic Hydrocarbons. Hungarian Standard Institution, Budapest.
- [39] MSZ 21470-94, 2001. Environmental Protection. Soil Analysis. Determination of Extractable Petroleum Hydrocarbon Content in the Boiling Range 160-520
- [40] Gas Chromatographic Method. Hungarian Standard Institution, Budapest. 2002.
- [41] Muntean, A., Rusu, T., 2011. The nitrogen regime in arable soils under the influence of tailings from mining landfills in the urban perimeter of Baia Mare. *Bulletin UASVM Agriculture* 68, 101-107.
- [42] Osuji, L.C., Onojake, C.M., 2004. Trace heavy metals associated with crude oil: a case study of Ebocha-8 oil-spill-polluted site in Niger Delta, Nigeria. *Chemistry and Biodiversity* 11, 1708-1715.
- [43] Pardo, R., Herguades, M., Barado, E., Vega, M., 2003. Biosorption of cadmium, copper, lead and zinc by inactive biomass of *Pseudomonas putida*. *Analytical and Bioanalytical Chemistry* 376, 26-32.
- [44] Polz, M.F., Cavanaugh, C.M., 1998. Bias in template-to-product ratios in multitemplate PCR. *Applied and Environmental Microbiology* 64, 3724-3730.
- [45] Popp, N., Schlömn, M., Mau, M., 2006. Bacterial diversity in the active stage of a bioremediation system for mineral oil hydrocarbon-contaminated soils. *Microbiology* 152, 3291-3304.
- [46] Roy, S., Hens, D., Biswas, D., Kumar, R., 2002. Survey of petroleum degrading bacteria in coastal waters of Sunderban Biosphere Reserve. *World Journal of Microbiology and Biotechnology* 18, 575-581.
- [47] Sarkar, D., Ferguson, M., Datta, R., Birnbaum, S., 2004. Bioremediation of petroleum hydrocarbons in contaminated soils: comparison of biosolids addition, carbon supplementation, and monitored natural attenuation. *Environmental Pollution* 136, 187-195.
- [48] Sevgi, E., Coral, G., Gizir, A.M., Sangün, M.K., 2010. Investigation of heavy metal resistance in some bacterial strains isolated from industrial soils. *Turkish Journal of Biology* 34, 423-431.
- [49] Singh, S.K., Tripathi, V.R., Jain, R.K., Vikram, S., Garg, S.K., 2010. An antibiotic, heavy metal resistant and halotolerant *Bacillus cereus* SIU1 and its thermoalkaline protease. *Microbial Cell Factories* 9. <http://dx.doi.org/10.1186/1475-2859-9-59>.
- [50] Smith, C.J., Danilowicz, B.S., Clear, A.K., Costello, F.J., Wilson, B., Meijer, W.G., 2005. TAlign, a web-based tool for comparison of multiple terminal restriction fragment length polymorphism profiles. *FEMS Microbiological Ecology* 54, 375-380.
- [51] Stancu, M.M., Grifoll, M., 2011. Multidrug resistance in hydrocarbon-tolerant Gram-positive and Gram-negative bacteria. *Journal of General and Applied Microbiology* 57, 1-18.
- [52] Tamura, K., Dudley, J., Nei, M., Kumar, S., 2007. MEGA4: Molecular Evolutionary Genetics Analysis (MEGA) software version 4.0. *Molecular Biology and Evolution* 24, 1596-1599.
- [53] Thavamani, P., Malik, S., Beer, M., Megharaj, M., Naidu, R., 2012. Microbial activity and diversity in long-term mixed contaminated soils with respect to polycyclic aromatic hydrocarbons and heavy metals. *Journal of Environmental Management* 99, 10-17.
- [54] Trindade, P.V.O., Sobral, L., Rizzo, A.C.L., Leite, S.G.F., Soriano, A., 2004. Bioremediation of a weathered and a recently oil-contaminated soils from Brazil: a comparison study. *Chemosphere* 58, 515-522.
- [55] Wasi, S., Tabrez, S., Ahmad, M., 2010. Isolation and characterization of *Pseudomonas fluorescens* strain tolerant to major Indian water pollutants. *Journal of Bioremediation and Biodegradation*. <http://dx.doi.org/10.4172/2155-6199.1000101>.
- [56] Wolicka, D., Suszek, A., Borkowski, A., Bielecka, A., 2009. Application of aerobic microorganisms in bioremediation in situ of soil contaminated by petroleum products. *Bioresource Technology* 100, 3221-3227.
- [57] Yumoto, I., Nakamura, A., Iwata, H., Kojima, K., Kusumoto, K., Nodasaka, Y., Matsuyama, H., 2002. *Dietzia psychrophila* sp. nov., a novel, facultatively psychrophilic alkaliphile that grows on hydrocarbons. *International Journal of Systematic and Evolutionary Microbiology* 52, 85-90.
- [58] Zhou, H.W., Wong, A.H.Y., Yu, R.M.K., Park, Y.D., Wong, Y.S., Tam, N.F.Y., 2009. Polycyclic aromatic hydrocarbon-induced structural shift of bacterial communities in mangrove sediment. *Microbiological Ecology* 58, 153-160.