

Short Communication

Role of salicylic acid on conifer somatic embryogenesis

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This review paper highlights about the role of salicylic acid on somatic embryogenesis and also recent updates on cloning mature trees of conifers using salicylic acid were discussed. Salicylic acid (SA) is a mobile molecule, which is capable of acting as a cell signal that senses, amplifies, and transmit information from a cell and might help in programming towards embryogenesis during cloning. Very recently incorporation of 1.0 mg⁻¹ SA in the DCR induction medium was found to be optimum for all the genotypes of *P. roxburghii* in terms of increasing the percentage of somatic embryogenesis compared against control. Therefore, salicylic acid can be used as growth regulator in conifer somatic embryogenesis and its use might help to solve the low initiation frequencies of many other recalcitrant pines. However, the mechanism of salicylate-induced differentiation in plants is not known, although salicylate is a signal molecule implicated in eliciting many physiological functions in plants.

Key words: Cloning, conifers, mature trees, forestry, India, signaling

Abbreviations: ASA- acetylsalicylic acid; DCR- Durzan and Gupta medium; H₂O₂. Hydrogen peroxide; PGR- Plant growth regulators; SA-salicylic acid; TDZ- thidiazuron; tTCLs-transverse thin cell layers;

Overview

The plant growth regulator, salicylic acid (SA), when applied to plants, affects diverse physiological processes (Malabadi *et al.* 2008a, 2008b; Cleland and Ajami, 1974; Cleland *et al.* 1982; Conrath *et al.* 1995; Dean and Delaney, 2008; Durner and Klessig, 1995; Gutierrez-Coronado *et al.* 1998; Hao *et al.* 2006; Hutchinson and Saxena, 1996; Komaraiah *et al.* 2004; Larque-Saavedra, 1978, 1979; Leslie

and Ramani, 1988; Luo *et al.* 2001). In African violet, the application of SA at low concentrations affects plant size, and the number of leaves and flowers. Moreover, application of SA to the growth medium induced flowering in several species of Lemnaceae. SA has also been reported to increase the activity of superoxide dismutase, and inhibit activities of ascorbate peroxidase, and catalase, thus leading to endogenous

H₂O₂ accumulation (Larque-Saavedra, 1978, 1979; Leslie and Ramani, 1988; Luo *et al.* 2001). Thus, SA inhibits the decomposition of H₂O₂ produced in plants. SA is an important signaling molecule involved in plant defense responses to pathogens and abiotic stress, as well as in plant growth and development. There are several reports describing exogenous SA and acetylsalicylic acid (ASA) enhancing somatic embryogenesis in plants *viz.* carrot (*Daucus carota*), pearl millet (*Pennisetum americanum*), geranium (*Pelargonium × hortorum* Bailey), *Astragalus adsurgens* Pall, *Plumbago rosea* L., naked oat (*Avena nuda*), and *Pinus roxburghii*. On the other hand salicylic acid also promotes induction of flowering in duckweeds, increased cell division, and growth during somatic embryogenesis in tissue culture of *Coffea arabica*, stomatal closure in *Phaseolus vulgaris*, increase in the accumulation of nitrates in roots of *Pinus patula*, inhibition of the biosynthesis of ethylene in cell suspension cultures of pears and seed germination in species, and root growth stimulation of soybean (Malabadi *et al.* 2008a, 2008b; Cleland and Ajami, 1974; Cleland *et al.* 1982; Conrath *et al.* 1995; Dean and Delaney, 2008; Durner and Klessig, 1995; Gutierrez-Coronado *et al.* 1998; Hao *et al.* 2006; Hutchinson and Saxena, 1996; Komaraiah *et al.* 2004; Larque-Saavedra, 1978, 1979; Leslie and Ramani, 1988; Luo *et al.* 2001; Martin-Max *et al.* 2005; San Miguel *et al.* 2002; Raskin, 1992; Quiroz *et al.* 2001; Ping *et al.* 2001).

Role of salicylic acid on somatic embryogenesis

SA is an important signaling molecule not only involved in defense responses but also in somatic embryogenesis of many plant species. SA is a mobile molecule, which is capable of acting as a cell signal that senses, amplifies, and transmit information from a cell and might help in programming towards embryogenesis during cloning. Secondly, SA is involved (together with nitrogen oxide,

hydrogen peroxide, and other metabolites) in the function of several signal systems, unifying them into an intricate network of regulatory interactions. Perhaps embryo differentiation may sharing some of the intermediates in the salicylate signal pathway (Malabadi *et al.* 2008a, 2008b; Cleland and Ajami, 1974; Cleland *et al.* 1982; Conrath *et al.* 1995; Dean and Delaney, 2008; Durner and Klessig, 1995; Gutierrez-Coronado *et al.* 1998; Hao *et al.* 2006; Hutchinson and Saxena, 1996; Komaraiah *et al.* 2004; Larque-Saavedra, 1978, 1979; Leslie and Ramani, 1988; Luo *et al.* 2001; Martin-Max *et al.* 2005; San Miguel *et al.* 2002; Raskin, 1992; Quiroz *et al.* 2001; Ping *et al.* 2001).

In a recent study, it was observed that SA acts as a signaling molecule during cloning mature trees of *P. roxburghii*. Therefore, SA can be used as growth regulator in conifer somatic embryogenesis and its use might help to solve the low initiation frequencies of many other recalcitrant pines. Embryogenic cultures have been generated for most of the conifers. For the most part, however, even the best of these systems lack commercial viability for two reasons: first, a low frequency of regeneration for many of the most desirable clones; and secondly, unproven genotypes, as starting material for the cultures is derived from seeds or seedlings. Therefore, the current approach of cloning mature trees of conifers using SA has many practical applications particularly in clonal forestry schemes. Use of SA might be helpful in solving many problems of conifer somatic embryogenesis. The pre-treatment of tTCLs from any of the 10 genotypes of *P. roxburghii* with 0.1, 0.2 and 0.4 mg⁻¹ SA could not effectively increasing the percentage of somatic embryogenesis when compared to the control (Malabadi *et al.* 2008a, 2008b; Malabadi and Teixeira da Silva, 2011). Pretreatment of explants with higher concentrations (2.0-5.0 mg⁻¹) of SA might have had a toxic effect and resulted in the browning of explants without callus

formation in all 10 genotypes. Incorporation of 1.0 mg⁻¹ SA in the induction medium was optimum for all 10 genotypes by increasing the percentage of somatic embryogenesis compared to the control. The addition of 1.0 mg⁻¹ SA to the induction medium was very beneficial since in the control this genotype failed to induce somatic embryogenesis. This clearly indicates the positive role of SA as a signaling molecule during cloning of mature *P. roxburghii* trees. In this study SA alone (i.e. without PGRs) did not induce somatic embryogenesis and resulted in the browning of explants and callus. Therefore, SA when combined with 22.6 µM 2, 4-D, 26.8 µM NAA, 8.9 µM BAP in the induction medium, improved the percentage of somatic embryogenesis. Hence, the combination of SA with other PGRs such as 2,4-D/NAA/BA might be beneficial in inducing somatic embryogenesis in Chir pine. In geranium (*Pelargonium x hortorum* Bailey), thidiazuron (TDZ) effectively induced somatic embryogenesis in cultured hypocotyls explants during only a 3-day period of induction. The presence of acetylsalicylic acid (ASA) during this period caused a two-fold increase in the number of somatic embryos an enhanced synchronization of embryo development compared to the TDZ treatment alone. However, in the same study, SA was ineffective in modulating similar embryogenic responses as ASA in geranium. Enhanced somatic embryogenesis and plant regeneration have been obtained using young leaf bases of naked oat (*Avena nuda*) as explants by including 0.5 mM SA and carrot embryogenic callus extracts in MS media. An improvement was achieved in somatic embryogenesis and plant regeneration on the corresponding media supplemented with 0.5 mM SA and carrot embryogenic callus extracts as compared to control. Somatic embryogenesis was induced from suspension cultures derived from leaf callus of an important medicinal plant, *Plumbago rosea* L. in which 8.32 µM ASA alone induced

embryogenesis, but IAA, NAA or IBA alone failed to elicit a similar response. Optimal embryogenic response per culture (216 embryos per culture) was observed in MS medium containing a combination of ASA (8.32 µM) and IAA (5.06 µM), i.e. a similar synergistic response as observed in our study between SA/ASA and other PGR(s) in the medium. It was also observed that by increasing the concentration of ASA alone (without auxin) in the medium (up to 11.09 µM) the number of somatic embryos formed per culture increased. The interactive effect of ASA and IAA appears to be essential for enhanced production of embryos per culture since no embryogenesis was noticed when IAA alone was added in *Plumbago rosea* L. SA is endogenously produced in many plants in normal as well as during stress conditions. However, the mechanism of salicylate-induced differentiation in plants is not known, although salicylate is a signal molecule implicated in eliciting many physiological functions in plants. It was also reported that the inclusion of SA to differentiation medium below 200 µmol/L significantly enhanced somatic embryogenesis in *Astragalus adsurgens* Pall., callus cultures, the highest frequency of somatic embryogenesis occurring at 150 µmol/L SA. They also reported that enhanced somatic embryogenesis by SA was accompanied by an increase in the endogenous hydrogen peroxide (H₂O₂) level compared to controls (Malabadi et al. 2008a, 2008b; Cleland and Ajami, 1974; Cleland et al. 1982; Conrath et al. 1995; Dean and Delaney, 2008; Durner and Klessig, 1995; Gutierrez-Coronado et al. 1998; Hao et al. 2006; Hutchinson and Saxena, 1996; Komaraiah et al. 2004; Larque-Saavedra, 1978, 1979; Leslie and Ramani, 1988; Luo et al. 2001; Martin-Max et al. 2005; San Miguel et al. 2002; Raskin, 1992; Quiroz et al. 2001; Ping et al. 2001).

This increased endogenous H₂O₂ level was related to the inhibition of ascorbate peroxidase and catalase activities. Although

the promoting effect of exogenous H₂O₂ was significantly lower than that of exogenous SA on the development of somatic embryos, the pre-treatment of callus cultures of *A. adsurgens* with dimethylurea (a trap for H₂O₂) significantly inhibited somatic embryogenesis, even if callus was cultured on the differentiation medium supplemented with 150 µmol/L SA, suggesting that endogenous H₂O₂ was required for SA-enhanced somatic embryogenesis in *A. adsurgens* (Malabadi et al. 2008a, 2008b; Cleland and Ajami, 1974; Cleland et al. 1982; Conrath et al. 1995; Dean and Delaney, 2008; Durner and Klessig, 1995; Gutierrez-Coronado et al. 1998; Hao et al. 2006; Hutchinson and Saxena, 1996; Komaraiah et al. 2004; Larque-Saavedra, 1978, 1979; Leslie and Ramani, 1988; Luo et al. 2001; Martin-Max et al. 2005; San Miguel et al. 2002; Raskin, 1992; Quiroz et al. 2001; Ping et al. 2001).

Therefore, one possible link between oxidative stress and plant regeneration in tissue culture could be H₂O₂. SA is endogenously produced in many plants in normal as well as during stress conditions. SA also inhibited ethylene biosynthesis in cell suspension cultures of carrot. It is well known that ethylene inhibits differentiation in plants. Therefore, SA may be promoting embryo development by inhibiting ethylene biosynthesis. Another hypothesis is that SA has been reported to increase the activity of superoxide dismutase, and inhibits the activities of ascorbate peroxidase and catalases, thus leading to endogenous H₂O₂ accumulation in *Arabidopsis thaliana*.

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