

# Curvularia linata as bioherbicide for management of Xanthium strumarium L., an abnoxious weed - A Critical Evualation

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### Abstract

Xanthium strumarium L. is an exotic plant responsible for several agricultural, environmental and health problems in India. Due to non-acceptability of conventional methods of control, the possibilities of its management through an indigenous strain of Curvularia lunata had been explored. In the present study, the organism was recovered in pure culture from the infected/infested parts of the weed, seedling and detached leaf bioassays were employed to evaluate the effect of different epidemiological factors on efficacy of mycoherbicidal agent. Significant effect on herbicidal potential of C. lunata was recorded when subjected to different conditions. Maximum efficacy of the pathogen was observed at 30°C, 80% RH, 2-3 leaves stage when sprayed @ of 4.5 x 105 Sp. /ml. The present study experimentally emphasizes on the development of Curvularia lunata as a mycoherbicide in the management of the weed, Xanthium strumarium L.

**Keywords:** Curvularia lunata, mycoherbicide, pathogen, Xanthium strumarium L.

### INTRODUCTION

Xanthium strumarium L. popularly known as Common cocklebur an exotic plant responsible for several agricultural, environmental and health problems in India. Due to non-acceptability of conventional methods of control, the possibilities of its management through an indigenous strain of Curvularia lunata had been explored. Preliminary evaluation studies viz. Pathogenicity, herbicidal potential, safety to non-target organisms etc. carried out in laboratory conditions and the pathogen was found to have excellent mycoherbicidal potential against this weed. However, herbicidal potentials of mycoherbicidal agents are known to be influenced by environmental conditions and growth stage of the weeds.

Bioherbicides are biological agents applied to plants in similar ways as chemical herbicides to control the propagation of weeds. The active ingredient in a bioherbicide is the living microorganisms which are applied in doses of propagules. The most commonly used microorganism is a fungus and the propagules are the spore or fragments of mycelium. In the present study the bioherbicide is referred as a Mycoherbicide as it is a fungus. Xanthium strumarium is a coarse annual herb, belongs to family Asteraceae and is popularly known as common cocklebur, banokra, Gokhru or Chota Datura. It is an extremely competitive weed creating several serious problems in agriculture and rangelands [1]. It grows luxuriantly and seriously in infested paddy, sorghum and other kharif annual cropfields in Andhra Pradesh, Maharashtra, Rajasthan and Madhya

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**Bioassay** 

The efficacy of the fungus Curvularia lunata was evaluated by detached leaf bioassay [10] and intact plant bioassay [11-13].

Pradesh [2, 3]. The weed is considered as one of the world's worst

weed [4]. All the parts of the weed are highly toxic and allergic to

humans and animals [5, 6]. The major toxic substance in Xanthium is

carboxyatractyloside which is capable of killing hogs, cattle, goats,

horses, sheep and poultry. Though the seed and seedlings contain

the highest quantity of toxin, the whole plant can also be toxic [7].

The allelochemicals produced from different parts of the weed also

inhibit the seed germination and seedling growth of many crops viz.

Wheat, maize, pearl millet, chickpea, rapeseed, tobacco and lettuce

[8]. Therefore the present investigation was carried out to critically

evaluate the possibility of using Curvularia lunata as bioherbicide

Systematic periodical and thorough survey of various

localities of Jabalpur was conducted and diseased specimens of the

fungus Curvularia lunata were collected. The organism was

recovered in pure cultures as per Agarwal et al. [9].

# **Mycoherbicidal Potential**

against Xanthium strumarium L.

**MATERIALS AND METHODS** 

Recovery of Pathogen

Effect of temperature on spore germination & incidence of disease:

The inoculum was prepared and seedlings at 2-3 leaf stage were inoculated, incubated at 80-100%, RH humidity and different temperatures viz., 15, 20, 25, 30, 35°C. Efficacy of test fungus was evaluated as mentioned earlier.

Relative humidity: Seedlings at 2-3 leaf stage were inoculated with test fungus, RH conditions ranging from 33% to

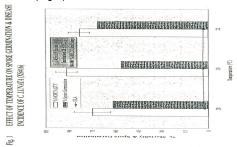
100% were provided in air tight desiccators' using various chemicals I14. 151.

Weed growth stage: Seedlings of X. strumarium from cotyledonary to pre-inflorescence stage were inoculated with test fungus. Plants were subjected to 24hrs incubation of dew period and moved to green house and the disease severity was evaluated as mentioned earlier.

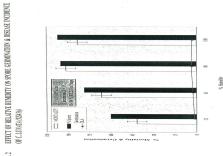
*Inoculum Concentration*: Different concentrations of inoculums were prepared in sterile distilled water. Spore suspension was sprayed by using atomizer on 2<sup>nd</sup> -3<sup>rd</sup> leaf stage of the weed.

### **RESULTS**

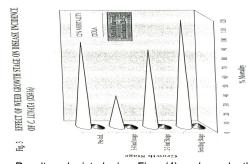
The temperature of 28-30°C supported maximum spore germination of test fungus, which was directly related with disease development (Fig 1). The maximum mortality percent of weed was observed at 30°C (Fig 1).



The results presented in Fig (2) clearly indicate that relative humidity had significant impact on the disease severity. Maximum incidence of disease was recorded at 80-100% More than 91.6% seedlings were died at these humidity.

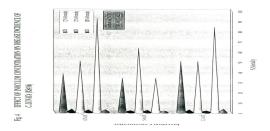


From data presented in the Fig (3), it is evident that, the weed was highly susceptible to all the growth stages; however, it was maximum at the cotyledonary or 1st -2nd true leaf stage.



Results depicted in Fig (4), showed that inoculum concentration had significant influence on disease development, Maximum disease severity was observed at 6.7 x  $10^5$  sp/ml and below 3. 7x  $10^5$  sp/ml pathogen had no significant effect on mortality

of the weed.



### **DISCUSSION**

The development of a fungus as potential mycoherbicide needs the knowledge of its interaction with the environment as well as the host plant. Significant influence of various environmental factors on biological activities at the test pathogen was also observed during the course of the present investigation. A range of temperatures i.e. 28-32°C supported maximum spore germination and percent mortality of the weed. Similar results have also been reported by many other workers [16-18] while evaluating the potential of Alternaria crassa for biological control of Jimson weed. Although the spore germinated optimally between 20- 30°C the infectivity of *Datura strumarium* was reduced at temperatures between 20-30°C, but the temperature between 20-30°C favored epidemics of Alternaria cucumerina on cucurbits in Florida [19]. Maximum incidence of disease was recorded at 80-100% RH. More that 91.6% seedlings died at this humidity. Similar results were observed while working with Colletotrichum lindemuthianum, dematium and Curvularia lunata [17, 18, 20]. It is also a known fact that spore of the pathogen germinate rapidly under higher humidity conditions, while they tend to desiccate in the absence of moisture [21, 22]. The weed was highly susceptible at the cotyledonary or 1-2 leaf stage. Increase in infection from the youngest to oldest leaves or plants due to Alternaria have also been reported by [13, 23, 24]. Similar variation in susceptibility has also been reported on same plants that plant age and inoculum concentration played a major role in the development of Aschocyta blight of Chickpea [25-27]. The pathogen was capable to produce a disease after two days of treatment and reached its maxima after 14 days of treatment. Similar results regarding inoculum concentration versus growth stage for different bio-control agents and their respective hosts were earlier reported [28-30]. Thus it can be boldly concluded that the mycoherbicidal agents can be applied in the field conditions for the biological control of weeds.

## **REFERENCES**

- [1]. Vargas, R. 1984. Weed Management system for cotton. pp 52-56. In: Proceedings 36th Annual California Weed Conference.164 pp.
- [2]. Deshpande, K. 1982. Biocontrol of *Parthenium hysterophorus* L. and *Xanthium strumarium* L. through phytopathogens. In: Abstracts of paper's Annual conference of Indian society of weed science. pp. 48.
- [3]. Kaul,V. 1965. Physiologicalecology of *X. strumarium* L.1. Seasonal, morphological variants and distribution. *Tropical Ecology*. 6: 72 87.

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- [4]. Holm, L. G., D. L. Pluncknett, I. V. Pancho, and J.P. Herberger. 1977. The world's worst weeds. University Press of Hawaii, Honolulu. pp. 609.
- [5]. Kings bury, J. M. 1964. Poisonous plants of the U. S. and Canada. Prentice- Hall, Inc. Engle wood, Cliffs N. I. pp. 626.
- [6]. Parsons, W. T. 1973. Noxious weeds of Victoria. Inkata Press, Ltd., Melbourne, Australia, pp. 300.
- [7]. Hatch, R. C., A. V. Jain, R. Weiss, J. D. Clark. 1982. Toxicological study of Carboxyatractyloside (active principle in cocklebur *X. strumarium* in rats treated with enzyme inducers and inhibitors and glutathione precursor and depletor. *American*. *J. Vet. Res.* 43: 111 - 116.
- [8]. Cutler, H. G. 1983. Carboxyatractyloside: a compound from *Xanthium strumarium* and *Atraetylis gummifera* with plant growth inhibiting properties. The probable "Inhibitor A". *J. Natural Products.* 46: 609-613.
- [9]. Agarwal, G.P. and S. K. Hasija. 1986. Microorganisms in the laboratory. A laboratory guide for Mycology, Microbiology and Plant Pathology. Print House, Lucknow (India). pp.155.
- [10]. Chiang, M.Y., C. G. Van Dyke and K. J. Leonard. 1989. Evaluation of endemic foliar fungi for potential biological control of Johnsongrass (Sorghum halepense): screening and host range tests. Plant Diseases. 73: 459-464.
- [11]. Boyette, C. D. and H. K. Abbas. 1995. Weed control with mycoherbicides and phytotoxins. In Allelopathy Organisms, Process and Applications (Indrajit K., K.M. M. Dakshin and Einhellig eds.) A.C.S. Symp. Ser. pp. 281- 299.
- [12]. Farkya, S. 1994. Evaluation of *Fusarium* sp. for management of *Parthenium hysterophorus* L., Ph. D., Thesis, R. D. University Jabalpur, M. P., India.
- [13]. Pandey, A. K. 1999. Herbicidal potential of microorganisms: Present status and Future prospects. In: Microbial Biotechnology For sustainable development and productivity. Prof. S. K. Hasija Festschrift vol. I (ed. R. C. Rajak) Scientific Publishers, Jodhpur pp. 85 - 105.
- [14]. Johnson, C. 1940.The maintenance of high atmospheric humidity for entomological work - Glycerol water mixture. Ann. Appl. Biol. 22: 295- 299.
- [15]. Mclean, R C. and W. R. I. Cook. 1952. Plant Science formulae. Mac Milan & Co. Ltd. London. pp.205.
- [16]. Boyette, C. D. and H. L. Walker. 1985. Factors influencing biocontrol of Velvetleaf (Abutilon theophrasti) and Prickly sida (Sida sp ioasa) with Fusarium latiritium. Weed Sci. 33: 209 -211.

[17]. Sharma, K. K. and V. K. Gupta. 1993. Germination and germ tube length of *Podosphaera leucotricha* on apple leaves. *Indian Phytopath*. 46: 408-410.

- [18] Thakur, M. P. and M. N. Khare. 1993. Factor's affecting sporulation and conidial germination of two species of Colletotrichum from mung bean. Indian J. Mycol. Plant. Pathol. 23: 188-190
- [19]. Jackson, C. R. 1959. Symptoms and host parasite relations of the *Alternaria* leaf spot disease of cucurbits. *Phytopathol.* 49: 731 733.
- [20]. Bhale, M. S. and M. N. Khare. 1984. Some factor's affecting Growth and spore germination of *Curvularia lunata* associated with Sorghum seeds. *Proc. Nat. Acad. Sci. India. B* 54: 252-255.
- [21]. Cartwright, D. K. and G. E. Templeton. 1992. Preliminary assessment of *Colletotrichum capsici* as a potential mycoherbicide for control of pitted morning glory. *Plant Dis.* 76: 995- 998.
- [22]. Gupta, D. and K. G. Nema. 1979. Effect of different temperature and relative humidity on the development of fruit rots of papaya caused by *Botryodiplodia theobromae* and *Colletotrichum* papaya. Indian Phytopath. 32:106-107.
- [23]. Brian, C. L. and M. K. D. Owen. 1995. Effect of moisture stress and leaf age on bentazon absorption in common cocklebur (X. strumarium and velvet leaf (Abutilion theophrasti). Weed Sci. 43: 7-12.
- [24]. Doworth, C. E. 1995. Biological control of red alder (*Alnus rubra*) with fungus, *Neetria ditissima. Weed Technol.* 1: 243-248.
- [25]. Srivastava, S. K. and S. U. Khan. 1993. Impact of host age at infection time on the severity of Myrothecium leaf spot disease of soybean. *Indian Phytopath*. 46: 190-191.
- [26]. Trapero-Casas, A. and W. J. Kaiser. 1992. Influence of temperature, wetness period plant age and inoculum concentration on infection and development of Ascochyta blight of chickpea. *Phytopathol*. 82: 589-596.
- [27]. Walker, H. L. and C. D. Boyette. 1985. Bio-control of Sickle pod (Cassia obtusifolia) in soybeans (Glycine max) with Alternaria cassiae. Weed Sci. 33: 212-215.
- [28]. Makowski, R M. D. 1993. Effect of inoculum concentration, temperature dew period and plant growth stage on disease of round leaved Mallow and velvet leaf by Colletotrichum gloeosporiodes of sp. malvae. Phytopathal. 83: 1229 - 1234.
- [29] Pandey, A. K., R. C. Rajak and S. K. Hasija. 1997. Application of Biotechnology in Development of Eco-friendly Mycoherbicides. In: New trends and prospects in biotechnology

(Eds. D. K. Maheshwari and R. C. Dubey) Gurukul Kangri University, Haridwar.

[30]. Winder, R. S. and K. Watson. 1994. A potential microbial control for fireweed (*Epilobium angustefolium*). *Phytoprotection*. 75: 19-23. 10.