

Pharmacognostical Studies and Preliminary Phytochemical Investigations on Roots of *Sophora interrupta* Bedd., Fabaceae

Panthati Murali Krishna*, T. Rajeshwar, P. Sai Kumar, S.Sandhya, K.N.V. Rao* and David Banji

Nalanda College of Pharmacy, Hyderabad main road, Cherlapally, Nalgonda, Andhra Pradesh-508001, India

Article Info	Summary
Article History Received : 15-05-2011 Revised : 03-09-2011 Accepted : 07-09-2011	<i>Sophora interrupta</i> Bedd., is a woody perennial shrub which belongs to the family Fabaceae. Many species of this genus like <i>S. flavescens</i> Ait., and <i>S. japonica</i> L. are used in traditional Chinese medicine. Several phytochemical investigations on this genus revealed the presence of many bioactive constituents like matrine and oxymatrine alkaloids, flavonoids, glycosides and polysaccharides which has medicinal importance. In view of its allied species, their importance in the Chinese medicine and in the absence of its scientifically reported pharmacognostical parameters, the present study attempts to undertake the study of qualitative and quantitative microscopic evaluation of the root along with physicochemical parameters and fluorescence analysis of root powder which helps to establish diagnostic characters and quality parameters for the identification of powdered form of root. Phytochemical evaluation of roots revealed the presence of flavonoids, alkaloids, saponins, glycosides and carbohydrates. TLC and HPTLC profile of Benzene extract was performed for flavonoids.
*Corresponding Author Tel : +91-8106995849 Email: raokav@yahoo.co.in	
©ScholarJournals, SSR	

Key Words: *Sophora interrupta* Bedd., *S. flavescens* Ait., *S. japonica* L., matrine, oxymatrine, flavonoids and polysaccharides

Introduction

Sophora is a genus of about 40 species in the family Fabaceae of herbaceous (*Sophora flavescens*) to trees (*Sophora japonica*). More than 15 species in this genus have a long history of use in traditional Chinese medicine [1]. Several phytochemical researches, in vivo and in vitro experiments and clinical practices have demonstrated that *Sophora* contains many phyto-constituents like matrine, oxymatrine type of alkaloids [2,3], flavonoids [4,5], saponins and polysaccharides [6] which possess wide-reaching pharmacological actions, including anti oxidant [7], anti cancer [8], anti asthmatic, anti neoplastic, antimicrobial [9], anti viral, antidote, anti pyretic, cardiotonic, anti inflammatory, diuretic and in the treatment of skin diseases like eczema, colitis and psoriasis [10].

Sophora interrupta Bedd., is a woody perennial shrub which grows endemically in seshachalam hill ranges, seshatheertham and kumaradhara theertham in Tirumala, India. Leaves are odd-pinnate, leaflets sub opposite, broadly ovate, pubescent below emarginated mucronate. Flowers are golden yellow in axillaries and terminal racemes. Calyx tube widely campanulate, oblique at mouth, teeth short and deltoid. Corolla much exerted petals 5 clawed, obtuse, stamens 10, and free anthers versatile. Ovary stipulate, many ovules, style incurved, stigma terminal and minute, pods 4 winged constricted between seeds. Seeds are 3-6, obovoid or globose and strophiole [11] (Figure 1).

Materials and Methods

Plant Material

The whole plant (*Sophora interrupta* Bedd., Fabaceae) was collected from surroundings of Seshachalam hill ranges of Tirumala, Tirupathi, Andhra Pradesh, India in the month of

November. The plant material was identified and authenticated by Mr. A. Laxma Reddy, Retired lecturer (Botany), Nagarjuna Government College, (Affiliated to Osmania University) Nalgonda. The plant specimen was prepared and submitted in the Department of Pharmacognosy under the voucher no: NCOP-NLG/Ph'cog/10-11/040.

Chemicals and Instruments

All the chemicals and reagents like chloral hydrate, phloroglucinol, hydrochloric acid, nitric acid, potassium hydroxide, picric acid, lead acetate etc., used were of analytical grade.

Sisco muffle furnace (3003137), Stage micrometer and eye piece micrometer.

Macroscopic and Microscopic Analysis

The macroscopy and microscopy of the plant was studied according to the methods of [12, 13], the cross sections were prepared and stained by using phloroglucinol and hydrochloric acid in the ratio 1:1 was used as a stain and mounted on a glass slide and focused under a microscope. The microscopic analysis of root powder was done according to the method of [14] and Kokate et al. (1986).

Physico-Chemical Analysis

Air dried plant material was used for the quantitative determination of ash and extractive values [15]. Fluorescence analysis of the extract(s) was carried out by the method of [16, 17].

Preliminary Phytochemical Screening

Preliminary phytochemical screening was carried out by using standard procedure described by [18, 19]. 100 gm of powdered root material was subjected to successive solvent extraction using petroleum ether benzene, chloroform, ethylacetate, acetone, alcohol and water based on increasing polarity using soxhalet apparatus. The extracts were concentrated under vacuum using rotary vacuum evaporator, dried and weighed. Each extract was tested for the presence of phytoconstituents viz. flavonoids, alkaloids, glycosides, saponins and carbohydrates. The TLC pattern of benzene extract was carried using precoated silica gel plates (Merck). The plates were developed using Benzene-Acetone-Ethyl acetate (8: 1: 1) mobile phase. The developed plate was sprayed with respective spray reagent 1% ethanolic aluminium chloride for flavonoids and dried at 100°C in Hot air oven and the spots were observed in UV 366 nm, it is followed by HPTLC studies using CAMAG LINOMAT 5 instrument, photodocumentation chamber (CAMAG REPROSTAR 3), CAMAG TLC SCANNER 3. An alluminum plate (2x10 cm) precoated Silica gel 60F₂₅₄ was used as adsorbent. The plates were developed using Benzene-Acetone-Ethyl acetate (8: 1: 1) mobile phase. The developed plate was sprayed with respective spray reagent 1% Ethanolic Aluminium chloride for flavonoids and dried at 100°C in Hot air oven. The plate was photo-documented at Day light and UV 366nm mode using Photo-documentation (CAMAG REPROSTAR 3) chamber. After derivatization, the plate was fixed in scanner stage (CAMAG TLC SCANNER 3) and scanning was done at UV

365nm. The Peak table, Peak display and Peak densitogram were noted.

Results and Discussion

Macroscopic Characters

The roots are woody, tuberous, and perennial, about 4 to 8 cm in diameter, light brownish yellow in colour with characteristic odour and highly bitter taste. Fractured surface is fibrous. Inner tuber is whitish cream coloured. The tubers show many rootlets (Figure 1).

Microscopic Characters

Cork cells are arranged in 12-14 layers with lignified, suberised rectangular shaped cells. Cortex composed of several layers of loosely arranged thin walled parenchymatous cells. In cortex the cells are arranged without any intercellular spaces. Calcium oxalate crystals are seen in this region. Vascular bundles are radially arranged, xylem and phloem constitutes xylem bundles, phloem bundles respectively and present alternatively on different radii. Uniseriate medullary rays are seen in between the vascular bundles, they are formed of radially arranged thin walled parenchymatous cells from centre to cortex (Figure 2)

Powder Characters

Powder of root material showed the presence of xylem vessels with annular and scalariform thickenings, cork cells, starch grains and calcium oxalate crystals. These powder characteristics can be used for diagnostic purposes of crude drug (Figure 3). Quantitative analysis of root powder was also done and the results were shown in the Table 1.

Table 1. Quantitative microscopy

Parameter	Length	Width
Phloem fibres	250-562.5-1250 μ	12.5-18.75-40μ
Xylem vessels	-----	37.5-62.5-87.5 μ
Starch grains	-----	12.5-18.75-31.25 μ

Table 2. Extractive values

S.No.	Parameters	Value(% w/w±SD)
	Water soluble	17.6±0.08
	Alcohol soluble	12.8 ±0.12
	Benzene soluble	16.9±0.09

Results are mean±S.D of 3 replicates.

Table 3. Ash values

S.No.	Parameters	Value(% w/w±SD)
	Total ash	3.95±0.12
	Acid insoluble	0.6±0.04
	Water soluble	7.6±0.152
	Sulphated ash	3.56±0.13

Results are mean±S.D of 3 replicates.



Figure 1. *Sophora interrupta* Bedd.

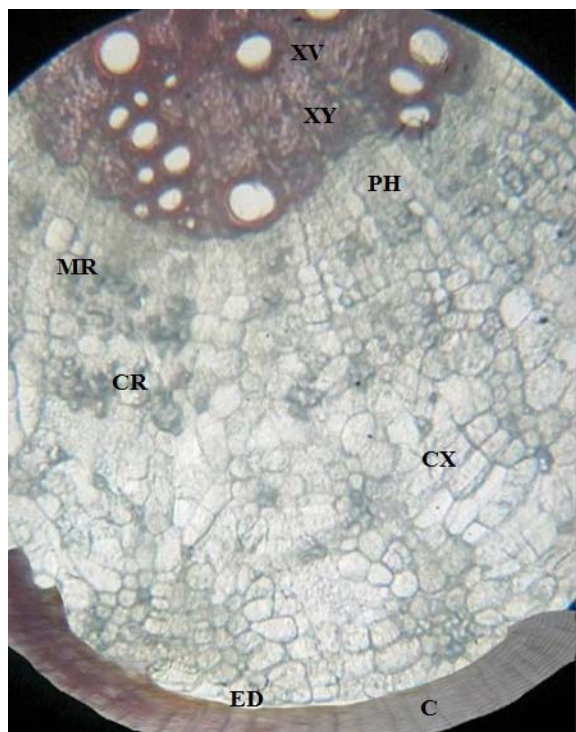


Figure 2. Transverse section of root: *S. interrupta* Bedd. (XV-Xylem vessel, XY-Xylem, PH-phloem, MR, Medullary rays, CR- Calcium oxalate crystals, CX-cortex, ED-Epidermis, and C-Cork)

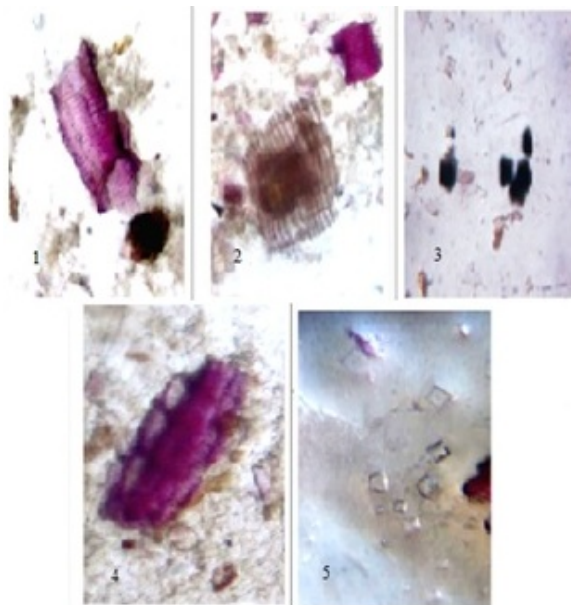


Figure 3. Powder characters of *S. interrupta* Bedd. root. (1. Scalariform xylem vessel, 2. Cork, 3. Starch grains, 4. Scalariform vessel with annular thickenings, 5. Calcium oxalate crystals.)

Physicochemical Analysis

Air dried root material was made into fine powder and used for quantitative determination of physicochemical values. Total, acid insoluble and water soluble ash (Table 2) was determined in triplicate and its mean+SD was calculated. Similarly, benzene, alcohol and water soluble extractives were determined in triplicate and its mean+SD was calculated (Table 3). Alcohol and water extractive values were determined as per WHO recommendations while benzene soluble extractive was determined due to the medicinal attributes of the extract. Water extractive was found to be very high when compared to other extractable matter in the powder. Fluorescence analysis of powder was carried out with different chemical reagents and observed under UV short, long and visible light (Table 4). The physical properties and nature of extracts prepared by successive extraction method are recorded (Table 5). Evaluation of physicochemical parameters is prerequisite for the preparation of drug monograph, thus ash values, extractive values, loss on drying, total fibre content and fluorescence characteristics can be used for standardizing of crude drugs and establishment of its quality parameters.

Table 4. Colour & Consistency of *S. interrupta* Bedd., root extracts

S.No	Extracts	Colour	Consistency	Yield % w/w
	Petroleum ether	Yellow	Sticky	7.4
	Benzene	Orange	Flakes	18.2
	Chloroform	Brown	Flakes	13.0
	Ethyl acetate	Brown	Amorphous	16.3
	Acetone	Red	Amorphous	11.6
	Alcohol	Reddish brown	Amorphous	12.5
	Water	Reddish brown	Amorphous	17.9

Table 5. Behaviour of root powder on treatment with different chemical reagents.

S.No.	Reagent	Long (366 nm)	Short (265 nm)	Day light
	Powder+50% H ₂ SO ₄	Black	Black	yellow
	Powder+50%HNO ₃	Black	Pale green	Yellow
	Powder+5% NaOH	Fluorescent green	Green	Yellowish brown
	Powder+1N Me NaOH	Yellowish brown	Green	Pale green
	Powder+1N KOH	Dark green	Green	Yellowish brown
	Powder+5% KOH	Black	green	Yellowish brown
	Powder+FeCl ₃	Black	Green	Yellow
	Powder+Methanol	-----	White	Yellow
	Powder+Conc HCl	-----	Green	Yellow
	Powder+Conc H ₂ SO ₄	-----	Black	Orange
	Powder+Ammonia	Pale green	Green	orange
	Powder+Conc HNO ₃	-----	Green	Yellowish orange

Table 6. Preliminary phytochemical screening.

Test	Pet. ether	Benzene	Chloroform	Ethylacetate	Acetone	Alcohol	Water
Carbohydrates	-	-	-	-	-	+	+
Amino acid	-	-	-	-	-	-	-
Protiens	-	+	-	+	-	-	-
Alkaloids	-	+	+	+	+	-	-
Tannins	-	-	-	-	-	-	-
Steroids	+	+	-	-	-	-	-
Flavonoids	+	+	+	+	+	-	-
Saponins	-	-	-	+	+	+	+
Glycosides	-	-	+	+	+	-	-

Table 7. Peak profile of benzene extract of root for flavonoids.

Peak	Rf	Height	Area	Assigned substance
1	0.02	199.6	3368.3	Flavonoid 1
2	0.10	172.8	5433.7	Flavonoid 2
3	0.16	163.7	9159.9	Flavonoid 3
4	0.21	117.2	3508.8	Flavonoid 4
5	0.26	84.2	2792.4	Unknown
6	0.34	110.3	6316.6	Unknown
7	0.40	106.2	4983.0	Flavonoid 5
8	0.53	487.0	26733.1	Flavonoid 6
9	0.58	132.8	4478.6	Flavonoid 7
10	0.66	31.8	901.8	Flavonoid 8
11	0.72	227.2	8946.3	Flavonoid 9
12	0.81	34.9	1009.4	Unknown
13	0.87	29.0	1109.4	Unknown

Preliminary Phytochemical Screening

Preliminary phytochemical analysis revealed the presence of flavonoids, alkaloids, glycosides, saponins and carbohydrates (Table 6)

Chromatographic Studies

TLC Profile

Yellow coloured fluorescent zones at UV 366 nm in benzene extract of *S. interrupta* Bedd., was observed in the TLC chromatogram after derivatization with spraying reagent for flavonoids four spots were observed with Rf values 0.40, 0.56, 0.64 and 0.69.

HPTLC Finger Print Profile

Yellow coloured fluorescent zones at UV 366nm in benzene extract of *S. interrupta* Bedd., track observed in the chromatogram after derivatization (Figure 4) which may be the presence of flavonoids in the given sample. It revealed the presence of 13 phytoconstituents, out of these 9 compounds are assigned to be flavonoids with Rf values 0.02, 0.10, 0.16, 0.21, 0.40, 0.53, 0.58, 0.66 and 0.72 with a most pronounced spot of maximum area at Rf 0.53 (Figure 5; Table 7). HPTLC developed profiles used for the detection, isolation and standardization of phytoconstituents profile in the extracts and it is useful in quality control of herbs and its extracts.

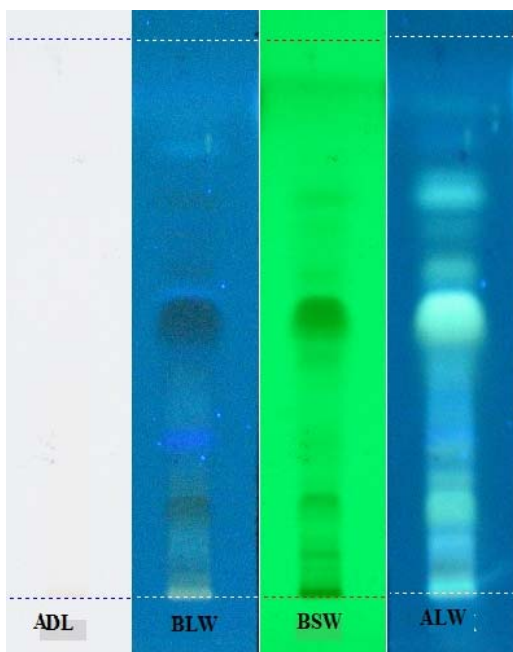


Figure 4. HPTLC chromatogram of benzene extract. ADL-After derivatization day light, BLW-before derivatization long wave length (UV 366 nm), BSW-before derivatization short wavelength (UV 254 nm), ALW-After derivatization long wave length (UV 366 nm).

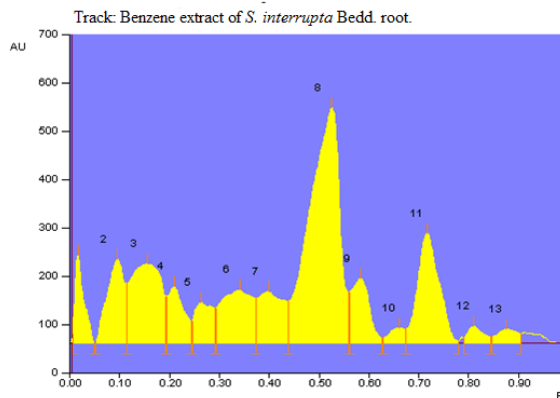


Figure 5. Benzene extract of *S. interrupta* Bedd., root Peak densitogram display (Scanned at 365nm)

Conclusion

The macro and microscopical characters along with physicochemical and fluorescence characters of root powder of *Sophora interrupta* Bedd., is used to establish the pharmacognostical standards and qualitative parameters as per pharmacopoeia and WHO guide lines. Preliminary phytochemical screening of different plant extracts revealed the presence of alkaloids, flavonoids, saponins, glycosides and carbohydrates. TLC profile of benzene extract was confirmed the presence of flavonoids followed by HPTLC finger print profile has given further conformation for the presence of flavonoids, thus further research on *Sophora interrupta* Bedd., is necessary for isolation and characterization of important bioactive constituents which have wide medicinal values in view of its allied species like *S. flavescens*, *S. japonica* and *S. alopecuroides* which has importance in traditional Chinese medicine [1].

Acknowledgement

We are highly grateful to the Management of Nalanda College of Pharmacy for providing necessary help and laboratory facilities in performing these studies and Mr. A. Laxma Reddy for authenticating the plant specimen.

References

- [1] Xiao, P., Kubo, H., Komiya, H., Higashiyama, K., Yan, Y.N., Li, J.S. and Ohmiya, S., 1999. Lupin alkaloids from seeds of *Sophora viciifolia*. *Phytochemistry* 50: 189-193.
- [2] Liu, M., Liu, X.Y., Cheng, J.F. 2003. Advance in the pharmacological research on matrine. *Zhongguo Zhong Yao Za Zhi*. 28: 801-804.
- [3] Zhang, Y.F Wang, S.Z., Li Y.Y., Xiao, Z.Y., Hu, Z.L. and Zhang, J.P. 2008. Sophocarpine and matrine inhibit the production of TNF and IL-6 in murine macrophages and prevent cachexia-related symptoms induced by colon 26 adenocarcinoma in mice. *International Immunopharmacol.* 8: 1767-1772.
- [4] Minhaj, N., Khan, H. and Zeman, A. 1976. Unanisoflavan, a new isoflavan from *Sophora secundiflora* DC. *Tetrahedron Lett.* 27: 2391-2394.
- [5] Minhaj, N., Khan, H. and Zaman, A., 1977. Secondifloran, a novel isoflavanone from *Sophora secundiflora* DC. *Tetrahedron Lett.* 13: 1145-1148.

- [6] Ohyama M, Tanaka T and Inuma M. 1994. Two novel resveratrol trimers, Leachianols A and B, from *Sophora leachiana*. Chem. Pharm. Bull. 42: 2117-2120.
- [7] Ding, P.L., Liao, Z.X., Huang, H., Zhou, P., and Chen, D.F. 2006. (+)-12 α -Hydroxysophocarpine, a new quinolizidine alkaloid and related anti-HBV alkaloids from *Sophora flavescens*. Bioorganic & Medicinal Chemistry Letters. 16: 1231–1235.
- [8] Tse, W.P., Che, C.T., Liu, K. and Lin, Z.X. 2006. Evaluation of the anti-proliferative properties of selected psoriasis-treating Chinese medicines on cultured HaCaT cells. Journal of Ethnopharmacology. 108: 133–141.
- [9] Sato, M., Tsuchiya, H., Takase, I., Kureshiro, H., Tanigaki, S. and Inuma, M. 1995. Antibacterial activity of flavanone isolated from *Sophora exigua* against methicillin-resistant *Staphylococcus aureus* and its combination with antibiotics. Phytotherapy Research. 9: 509–512.
- [10] Kinghorn, A.D., Balandrin, M.F. and Pelletier, S.W., (Eds.) 1984. Alkaloids: Chemical and Biological Perspectives, vol. 2, Wiley, New York, p.105-106.
- [11] Madhava, K.C., Sivaji, K. and Tulasi, K.R. 2008. Flowering Plants of Chittoor Dist. A.P. India, Students Offset Printers, Tirupati, p.141.
- [12] Easu, K. 1964. Plant Anatomy. New York: John Wiley and Sons, p. 767.
- [13] Brain, K.R. and Turner, T.D. 1975a. The Practical Evaluation of Phytopharmaceuticals, Wright-Scientific, Bristol, p 4-9.
- [14] Brain, K.R., and Turner, T.D. 1975b. In: The Practical Evaluation of Phytopharmaceuticals, Wright-Scientific, Bristol, p.36-45.
- [15] WHO/QCMMPM 1992. Quality Control Methods for Medicinal Plant Material, Organisation Mondiale De La Sante, Geneva, p.22-34.
- [16] Chase, C.R. and Pratt, R. 1949. Fluorescence of powdered vegetable drugs with particular reference to development of a system of identification, J. Am. Pharmacol. Assoc. 38: 324-331.
- [17] Kokoski, C.J., Kokoski, R.J and Slama, F.J. 1958. Fluorescence of powdered vegetable drugs under ultraviolet radiation. J. Am. Pharm. Assoc. 47: 715-717.
- [18] Kokate, C.K. 1986. Practical Pharmacognosy. Vallabh Prakashan, New Delhi, p. 111.
- [19] Harbone, J.B. 1998. Method of extraction and isolation, In: Phytochemical methods, Chapman & Hall, London, p.60-66.