

Integrated analysis of *Tinospora cordifolia* germplasm: Nutritional, qualitative, quantitative, and phytochemical traits

B C Akhilraj^{1*}, J Suresh², K Rajamani³, M Kumar⁴ & R Gnanam⁵

^{1*}Tamil Nadu Agricultural University, Coimbatore, Tamil Nadu

²Horticultural College and Research Institute for Women, Tamil Nadu Agricultural University, Aliyar Nagar, Pollachi, Tamil Nadu

³Tamil Nadu Agricultural University, Coimbatore, Tamil Nadu

⁴ICAR- Krishi Vigyan Kendra, Tamil Nadu Agricultural University, Tindivanam, Villupuram, Tamil Nadu

⁵Tamil Nadu Agricultural University, Coimbatore, Tamil Nadu

*Email: bcakhilraj@gmail.com

Received 28 October 2023; Revised 21 December 2024; Accepted 21 December 2024

Abstract

Tinospora cordifolia, often known as "Guduchi" is a deciduous climber and is considered valuable in terms of its immense medicinal properties. This study evaluated 48 *Tinospora cordifolia* germplasm accessions collected from Tamil Nadu for their nutritional, qualitative, quantitative, and phytochemical properties. Significant variations were observed among the accessions. TC-22 exhibited the highest phenol (122.73 mg/g) and flavonoid content (18.84 mg/g), while TC-30 recorded the highest tannin content (234.56 mg/g). TC-20 demonstrated superior protein (13.85%), carbohydrate (86.92 mg/g), and antioxidant activity (261.74%) levels. Additionally, TC-20 displayed elevated potassium (1.45%), calcium (0.35%), and iron (48.74%) levels, with the highest zinc content (15.67 µg/g) among accessions. TC-30 had the highest fat content (8.81%), while TC-22 recorded the maximum cardiac glycosides (2.27%). The study identified TC-35 as having the highest fiber content (13.22%) and energy value (99.66 Kcal/100g). Vitamin C (68.96 mg/100g) and vitamin A (0.44 mg/100g) were most abundant in TC-33. These findings provide valuable insights into the biochemical diversity of *T. cordifolia* germplasm, highlighting its potential for nutritional and therapeutic applications. Findings unmistakably demonstrated the significance of ecological condition variables in determining the quantitative and qualitative parameters of diverse accessions from the different places of Tamil Nadu.

Keywords: Germplasm, *Tinospora cordifolia*, Tamil Nadu, qualitative, quantitative

Introduction

The recent surge in the use of herbal medicines has led to a significant rise in Herbal drugs production facilities, driven by the growing demand for herbal raw materials. India's herbal product exports have risen in recent years, with \$ 628.25 million exported in 2022-23 and \$ 612.1 million in 2021-22. Top exports include ashwagandha, turmeric, and ginger, reflecting growing global demand for Ayurvedic and herbal products (Basai, 2023). *Tinospora cordifolia* Willd Miers ex Hook. F. and Thoms, often known as "Guduchi" is a diploid ($2n = 22$), deciduous climber in the family Menispermaceae. It spreads over the tropical and subtropical Indian subcontinent as well as China reaching heights of upto an altitude of about 300 m. Ancient Ayurveda writings describe *T. cordifolia* as an useful herb to cure general senility, dyspepsia, and urinary disorders (Ahmad et al., 2009).

Talbot et al. (2006) reported that it is pertinent to note that people's faith on natural products is on surge but at the same time, the need to safeguard the value chain to ensure quality of the raw material produced is to be realized which requires focus on research in identifying genotypes rich in bioactive constituents. India holds advantage of its rich plant biodiversity which is distributed in various habitats. Among the various medicinal plants used in AYUSH, *Tinospora cordifolia* is considered valuable in terms of its immense medicinal properties. Plant extracts of *T. cordifolia* are used for the rehabilitation of fever, diabetes, allergies, immune system, gout, skin

inflammation, snake and scorpion bites, respiratory tract infections, tuberculosis, rheumatoid arthritis, cancer, etc.

The stem and root of plant play an important role in water purification by sedimentation, flocculation or antibiosis. In addition, the leaves of the plant also contribute significantly to the treatment of respiratory ailments and serve as a source of vitamins A and C. Stem juice also shows strong antimicrobial, antimalarial and antiviral properties. Fruits of the plant are edible and are rich in ascorbic acid. Its kernel are rich source of carotenoids, lecithin and steroids. The plant alkaloids closely resemble like action to the antibiotics which are useful to treat asthma (Ahmed et al., 2006; Puratchimani and Jha, 2007; Srinivasan et al., 2008; Gahlaut et al., 2012; Tanwar et al., 2012 and Choudhary et al., 2014). *Tinospora* is cultivated commercially in India due to its significant demand in the Ayurvedic, pharmaceutical, and herbal product industries. Its cultivation is particularly prominent in states like Rajasthan, Madhya Pradesh, Uttar Pradesh, and Maharashtra. Farmers often grow *Tinospora* in agroforestry systems or as an intercrop with other species (Basai, 2023). In the current study, biochemical properties of *T. cordifolia* accessions that were gathered from various parts of Tamil Nadu were studied.

Materials and methods

Sample collection

Healthy and younger stem samples of 48 *Tinospora* wild populations were collected from different parts of Tamil Nadu during

2021-23 (Table 1). The collection was based on phenotypic evaluation of various traits, including leaf arrangement, color, size, length, and breadth; stem color, size, and internodal distance; as well as flower count, color, and size. The collected populations were established in the medicinal garden of the Horticultural College and Research

Institute, Tamil Nadu Agricultural University, following a RCBD (Randomized Complete Block Design) layout with three replications. The plants were propagated through stem cuttings and field planted, with a spacing of 2 m × 3 m, and cultural practices were carried out as per the scheduled guidelines (NMPB).

Table 1. Accessions of *T. cordifolia* collected from several sites in Tamil Nadu

Accession	Place	District	Latitude	Longitude	Altitude (MSL)
TC-01	Dharmathupatti	Theni	9°59'38"N	77°20'56"E	351 m
TC-02	Narasingapuram	Theni	9°56'18"N	77°36'01"E	341 m
TC-03	Palani	Dindigul	10°26'57"N	77°30'58"E	378 m
TC-04	Villacheri	Madurai	9°53'45"N	78°03'39"E	143 m
TC-05	Srivilliputhur	Virudhunagar	9°30'25"N	77°38'02"E	150 m
TC-06	Thalaivasal	Salem	11°35'10"N	78°45'30"E	162 m
TC-07	Thantheeni	Karur	10°55'50"N	78°05'49"E	131 m
TC-08	Samathuvapuram	Salem	11°49'32"N	77°55'09"E	360 m
TC-09	Kaveripattinam	Krishnagiri	12°25'05"N	78°13'05"E	464 m
TC-10	Nallampalli	Dharmapuri	12°03'33"N	78°06'49"E	485 m
TC-11	Papparapatti	Dharmapuri	12°12'25"N	78°03'26"E	494 m
TC-12	Perundurai	Erode	11°16'15"N	77°34'57"E	278 m
TC-13	Natrampalli	Thirupathur	12°35'07"N	78°30'29"E	443 m
TC-14	Kalavai	Ranipet	12°46'01"N	79°25'01"E	132 m
TC-15	Sendamangalam	Namakkal	11°16'51"N	78°14'15"E	180 m
TC-16	Somandargudi	Kallakurichi	11°45'36"N	78°55'56"E	135 m
TC-17	Cheyyar	Thiruvannamalai	12°39'12"N	79°32'39"E	90 m
TC-18	Thiruvetriyur	Ramanathapuram	9°42'29"N	78°57'02"E	11 m
TC-19	Gopalasamudram	Thirunelveli	8°40'22"N	77°37'49"E	55 m
TC-20	Thovalai	Kanyakumari	8°13'47"N	77°30'02"E	49 m
TC-21	Thally	Krishnagiri	12°35'09"N	77°39'09"E	910 m
TC-22	Nemili	Ranipet	12°36'07"N	78°31'00"E	435 m
TC-23	Gudiyatham	Vellore	12°56'12"N	78°52'46"E	268 m
TC-24	Pallikonda	Vellore	12°54'22"N	78°56'17"E	251 m
TC-25	Ashok Nagar	Chennai	13°02'09"N	80°12'43"E	10 m
TC-26	Virudhachalam	Cuddalore	11°30'52"N	79°19'31"E	33 m
TC-27	Neyveli	Cuddalore	11°32'07"N	79°28'46"E	35 m

TC-28	Kalpakkam	Kanchipuram	12°31'19"N	80°09'26"E	2 m
TC-29	Padappai	Kanchipuram	12°52'43"N	80°01'50"E	28 m
TC-30	Acharavakkam	Chengalpattu	12°41'44"N	79°58'41"E	32 m
TC-31	Pallipattu	Thiruvallur	13°19'59"N	79°26'34"E	158 m
TC-32	Vanthavasi	Thiruvannamalai	12°29'58"N	79°35'56"E	71 m
TC-33	Tindivanam	Villupuram	12°13'21"N	79°38'44"E	37 m
TC-34	Palaiyur	Mayiladuthurai	10°46'28"N	79°49'01"E	4 m
TC-35	Courtallam	Tenkasi	8°55'27"N	77°16'40"E	194 m
TC-36	Thirupuvanam	Sivagangai	9°49'25"N	78°15'15"E	106 m
TC-37	Thirukalukundram	Chengalpattu	12°36'18"N	80°03'50"E	33 m
TC-38	Mathampatti	Coimbatore	10°58'11"N	76°51'33"E	428 m
TC-39	Saravanampatti	Coimbatore	11°04'36"N	77°00'02"E	418 m
TC-40	Kinethukadavu	Coimbatore	10°48'39"N	77°01'20"E	308 m
TC-41	Kunnathur	Thirupur	11°15'43"N	77°25'01"E	313 m
TC-42	Kannaivadi	Thirupur	10°48'40"N	77°47'04"E	201 m
TC-43	Bhavanisagar	Erode	11°28'37"N	77°08'13"E	257 m
TC-44	Nilakottai	Dindigul	10°09'51"N	77°51'08"E	226 m
TC-45	Vedasandur	Dindigul	10°31'50"N	77°56'54"E	212 m
TC-46	Kallupatti	Madurai	9°43'04"N	77°51'13"E	142 m
TC-47	Karaiyipatti	Madurai	10°06'52"N	78°26'21"E	129 m
TC-48	Mohanur	Namakkal	11°03'18"N	78°08'39"E	117 m

Table 2 presents the methods used for qualitative phytochemical analysis, detailing the specific techniques employed to evaluate various biochemical compounds and nutrients in plant samples.

Results and discussion

Biochemical studies were conducted in the leaf samples of 48 *Tinospora* accessions, which revealed statistically significant differences among them for various parameters. The results are presented in Tables 3 and 4. Accession TC-22 exhibited the highest phenol content (122.73 mg/g), while TC-01 had the lowest content (46.56

mg/g). Previous studies have reported total phenolic contents in various plant species, such as *Bryophyllum pinnatum* (18.4 mg/g - leaves), *Ipomea aquatica* (18.8 mg/g - leaves), *Oldenlandia corymbosa* (11.6 mg/g - whole plant), *Ricinus communis* (29.2 mg/g - roots), *Terminalia bellerica* (29.6 mg/g - leaves), *Tinospora cordifolia* (40.8 mg/g - leaves), *Tinospora cordifolia* (12.8 mg/g - stem), and *Xanthium strumarium* (71.6 mg/g - leaves) as reported by Yadav and Agarwala (2011). Additionally, Kumar et al. (2013) observed a total phenol content of 17.3 ± 0.4 mg/g in *Tinospora* leaves.

Table 2. Methods of qualitative phytochemical analysis

Parameter	Methodology	Reference
Phenol	Bromination method	Ingberman (1958)
Flavonoids content (mg/g)	Aluminium chloride spectrometric technique	Chang <i>et al.</i> , (2002)
Antioxidant activity (%)	Stabilized DPPH radicals to measure free radical scavenging capacity	Koleva <i>et al.</i> , (2002)
Saponin	Determined using Obadoni and Ochuko technique	Obadoni and Ochuko (2001)
Cardiac glycosides	Quantitatively measured with minor adjustments to the described method	Solich <i>et al.</i> , (1992)
Protein (Bradford Method)	Bradford test procedure	Bradford (1976)
Carbohydrate content (mg/g)	Extractable carbohydrates measured using Anthrone method	Yemm and Willis (1954)
Tannin content	Modified AOAC procedure	AOAC (1980)
Nitrogen (%)	Determined using Kjeldahl technique	Kirk (1950)
Phosphorus (%)	Colorimetric technique with malachite green	Majed <i>et al.</i> , (2012)
Potassium (%)	Flame photometry	Bhandal and Malik (1988)
Calcium (%)	Titration with standardised EDTA solutions	Armstrong and Kirkby (1979)
Iron (ppm)	Spectroscopy	Jones (1991)
Magnesium (%)	Complexometric approach	Sahrawat, K. L. (1987)
Zinc (ppm)	Quantification via atomic-absorption spectroscopy	David (1958)
Manganese (ppm)	Determined using atomic absorption spectrometry	Tinggi <i>et al.</i> , (1997)
Copper (ppm)	Measured calorimetrically	Middleton (1965)
Fat content (%)	Recovery using a lipophilic solvent	Stien <i>et al.</i> , (2007)
Reducing sugar (%)	Quantified using Benedict's reagent	Thomas and Dutcher (1924)
Fibre (%)	Measured gravimetrically	Van Soest and McQueen (1973)
Energy	Total energy content estimated using FAO method	FAO (2003)
Vitamin C	Spectrometry for ascorbic acid quantification	Hassan and Hassan (2008)
Vitamin A	Measured using spectrophotometric techniques	Lopez-Teros <i>et al.</i> , (2017)

The present study recorded significantly higher protein content (13.85%) in TC-20. Tyagi and Chauhan (2020) reported a protein content of 6.26% in *Tinospora cordifolia* stem powder. Aranha et al. (2012) isolated Guduchi protein with a yield of 29 mg/100 g fresh stem or 7.0% in dried leaf powder. Among the accessions, TC-20 exhibited the highest carbohydrate content (86.92 mg/g) in our research. The presence of carbohydrates in *Tinospora* plants has been reported by Yadav and Agarwala (2011). Kumar et al. (2013) observed the presence of carbohydrates at a level of 84.1 ± 5.2 mg/g. Its high protein and carbohydrate content contributes to the provision of vital energy. In our current study, TC-22 showed the highest total flavonoid content (18.84 mg/g). Similarly, Yadav and Agarwal (2011) reported the presence of flavonoids in extract of various plants, including *Bryophyllum pinnatum* (8.4 mg/g - leaves), *Ipomea aquatica* (37.6 mg/g - leaves), *Oldenlandia corymbosa* (4.4 mg/g - whole plant), *Ricinus communis* (6 mg/g - roots), *Terminalia bellerica* (42.8 mg/g - leaves), *Tinospora cordifolia* (18 mg/g - leaves, 6 mg/g - stem), and *Xanthium strumarium* (28.8 mg/g - leaves). In another study, Kumar et al. (2013) identified flavonoid contents of 6.5 ± 0.2 mg/g in *Tinospora* leaves.

In our study, the highest tannin content of 234.56 mg/g was observed in accession TC-30, while the lowest content of 142.74 mg/g was recorded in TC-35. Choudhry et al. (2014) in a similar study reported tannin content of 154.1 mg/g in male and 162.5 mg/g in female *Tinospora* plants. Kumar et al. (2013) reported a tannin content of 133.8 ± 0.5 mg/g. Khan et al., (2020) discovered a

substantial quantity of tannin (204.39 ± 19.66 mg) in the methanol extract of *Tinospora*. According to Uadia et al., (2017), tannins are present in the leaf, stem, and root of *Vernonia amygdalina* and the tannin content in the root (33.90 mg) was lower compared to the leaf (69.33 mg) and the stem (50.70 mg).

The DPPH technique, commonly used to assess antioxidant activity, was employed to evaluate the extracts' ability to scavenge free radicals. In our study, the highest antioxidant activity was observed in TC-20 at 261.74%, while the lowest activity was found in TC-1 at 48.56%. Contrary to the findings of our study, Upadhyay et al. (2014) reported the best antioxidant activity of 84.62 mg/g in *Tinospora* ethanolic extract. According to Premanath and Lakshmidevi (2010), ethanolic and methanolic leaf extracts of *T. cordifolia* demonstrated higher efficacy in scavenging DPPH radicals compared to the aqueous stem extract, with an EC₅₀ value of 0.5 mg/mL for ethanol and 0.9 mg/mL for methanol. Accession TC-22 exhibited the highest levels of nitrogen (1.06%), phosphorus (1.17%), and magnesium (7.55%) in our study. However, Shivraj and Khobragade (2009) reported nitrogen content of 0.45%, phosphorus content of 0.57%, and magnesium content of 6.41% in *Tinospora* leaves.

In the present study, TC-20 exhibited the highest levels of potassium (1.45%), calcium (0.35%), and iron (48.74%). Consistent with these findings, Shivraj and Khobragade (2009) reported potassium content of 0.845%, calcium content of 0.13%, and iron content of 28.00% in *Tinospora* herbage. Our

research recorded significantly higher zinc content of 15.67 µg/g in TC-20. Similarly, Tyagi and Chauhan (2020) reported a good quantity of zinc (10.35 µg/g) in *Tinospora* leaf powder, and Shivraj and Khobragade (2009) also reported 12.00 µg/g of zinc in the same crop. TC-22 exhibited the highest manganese content of 17.76 ppm in our study, while Garg *et al.* (2022) reported manganese content of 56.7 ± 1.5 µg/g in the same plant. TC-30 recorded the highest copper content of 6.19 µg/g. Tyagi and Chauhan (2020) reported copper content of 10.01 µg/g, and Shivraj and Khobragade (2009) found 3.10 µg/g of copper in *Tinospora* leaf samples.

TC-30 showed the highest fat content of 8.81%. Modi *et al.* (2020) observed a fat content of 3.1% in *Tinospora* leaves. TC-20 exhibited the highest saponin content of 3.88% in our study, while Shervani and Mishra (2019) reported saponin content of 2.59% (Sample I), 2.34% (Sample II), and 2.10% (Sample III) in *Tinospora* cultivated in different soil conditions. TC-22 had the highest cardiac glycoside content of 2.27% in our study, whereas Pradhan *et al.* (2013) reported cardiac glycoside content of $1.52 \pm$

0.03 (Chhindwara), 0.56 ± 0.02 (Jabalpur), and 0.72 ± 0.02 (Seoni) in different districts of Madhya Pradesh.

TC-14 exhibited the highest reducing sugar content of 0.26% in our study. Similarly, Khan *et al.* (2011) observed reducing sugar content of 55 mg/g in freshly ripened *Tinospora* fruits. TC-35 showed significantly higher fiber content of 13.22% in our study, while Tyagi and Chauhan (2020) reported crude fiber content of 4.69% in *Tinospora* leaf powder. TC-35 had the highest energy content of 99.66 Kcal/100g in our study, whereas Shivraj and Khobragade (2009) reported an energy content of 292.54 Cal/100g in the same crop. TC-33 exhibited the highest vitamin C content of 68.96 mg/100g and vitamin A content of 0.44 mg/100g in our study. Gupta and Sharma (2011) reported vitamin C content of 41.36 mg/100g in *T. cordifolia* leaves. The conservation and utilization strategy involves establishing a field gene bank at the Medicinal Department of HC&RI, TNAU, where all 48 accessions will be cultivated and maintained under controlled conditions.

Table 3. Biochemical characterization of *Tinospora cordifolia* from diverse accessions

Accession	Phe. (mg/g)	Pro. (%)	Car. (mg/g)	Fla. (mg/g)	Tan. (mg/g)	AO (%)	N (%)	P (%)	K (%)	Ca (%)	Mg (%)	Fe(ppm)
TC-01	48.56	7.62	63.98	10.53	177.45	191.51	0.48	0.79	0.53	0.20	5.15	25.61
TC-02	67.34	5.98	49.94	9.86	201.75	209.43	0.72	0.91	1.01	0.14	4.83	29.48
TC-03	71.85	8.51	56.77	11.47	164.41	215.67	0.69	0.80	0.64	0.14	6.24	30.06
TC-04	86.76	6.88	72.58	8.66	210.94	187.56	0.57	0.58	0.98	0.10	3.95	27.91
TC-05	64.64	7.98	54.56	11.48	212.58	175.98	0.71	0.82	0.52	0.22	4.37	24.42
TC-06	98.12	10.76	73.01	10.56	192.56	210.56	0.82	0.69	0.56	0.16	5.02	22.57
TC-07	71.67	6.42	71.22	9.08	208.71	162.77	0.75	0.98	0.76	0.18	4.86	30.56
TC-08	95.61	9.23	63.47	10.77	149.74	181.05	0.62	0.58	1.01	0.17	4.98	27.41
TC-09	94.86	8.43	54.67	8.94	195.86	197.18	0.80	0.61	0.76	0.13	6.41	27.64
TC-10	86.74	9.76	50.51	12.56	165.71	179.86	0.75	0.82	0.93	0.25	6.24	23.87
TC-11	68.18	7.99	63.81	9.81	209.59	192.56	0.68	0.78	0.83	0.14	4.84	26.41
TC-12	91.76	5.71	74.47	8.56	193.56	174.86	0.74	0.75	0.79	0.19	5.39	29.56
TC-13	74.86	8.92	53.81	8.41	184.76	201.56	0.64	0.96	0.79	0.16	5.83	25.64
TC-14	75.84	5.57	69.78	10.66	180.62	210.56	0.74	0.64	0.82	0.17	5.92	28.42
TC-15	100.07	9.16	71.48	11.91	156.67	221.87	0.76	1.00	0.68	0.18	5.63	26.92
TC-16	89.78	7.85	63.54	10.64	200.56	200.02	0.61	0.71	1.11	0.13	4.16	21.57
TC-17	103.77	9.78	79.43	12.95	215.57	240.86	0.83	0.59	0.64	0.19	5.76	31.87
TC-18	79.56	5.47	76.09	8.25	173.85	209.14	0.58	0.69	1.02	0.16	5.63	28.37
TC-19	63.51	7.81	64.85	10.41	192.56	173.56	0.71	0.89	0.79	0.18	3.67	27.86
TC-20	116.84	13.85	86.92	15.58	213.58	261.74	0.85	1.06	1.45	0.35	6.39	38.74
TC-21	84.86	5.90	71.56	8.76	192.51	206.61	0.59	0.69	0.90	0.13	6.46	31.86
TC-22	122.73	12.93	85.66	18.84	227.51	256.47	1.06	1.17	1.29	0.29	7.55	35.55
TC-23	109.76	7.39	81.66	13.56	204.47	225.61	0.56	0.74	0.65	0.15	4.57	28.16
TC-24	92.54	6.50	73.59	12.41	187.34	214.76	0.66	0.63	0.77	0.26	5.69	29.13
TC-25	88.81	9.81	59.64	9.65	212.58	217.86	0.52	0.78	0.73	0.21	5.92	28.57
TC-26	110.49	12.68	81.54	17.78	221.58	234.56	0.90	1.04	0.95	0.22	7.12	34.75

TC-27	101.55	7.35	73.54	9.12	167.55	169.45	0.64	0.98	0.65	0.13	5.76	32.49
TC-28	83.97	10.85	50.85	10.76	156.48	174.76	0.53	0.89	0.62	0.16	4.47	29.47
TC-29	99.12	5.78	79.56	8.56	181.84	171.98	0.78	0.64	0.64	0.19	4.86	30.56
TC-30	107.53	8.22	78.63	16.71	234.56	260.57	0.75	0.88	0.59	0.18	6.94	31.86
TC-31	95.97	7.05	67.71	13.41	160.38	198.03	0.69	0.56	0.96	0.09	5.81	28.56
TC-32	113.46	11.43	75.76	16.08	219.76	255.61	0.86	1.12	1.44	0.27	6.41	33.60
TC-33	85.67	5.82	62.51	10.38	168.35	185.76	0.69	0.86	0.72	0.17	5.28	25.54
TC-34	92.86	7.92	74.48	9.78	196.67	177.56	0.63	0.93	0.74	0.16	5.62	23.89
TC-35	76.84	9.46	63.86	12.01	142.74	204.67	0.77	0.86	1.24	0.19	3.85	21.41
TC-36	82.78	6.76	54.48	11.09	176.48	186.76	0.84	0.75	0.65	0.15	5.42	30.86
TC-37	85.67	8.56	64.88	8.32	154.75	194.86	0.71	0.94	0.82	0.19	6.09	26.86
TC-38	75.67	7.12	53.85	10.05	177.71	197.67	0.79	0.85	0.86	0.17	4.28	30.56
TC-39	78.09	6.55	66.44	13.55	185.56	201.34	0.65	0.67	0.83	0.18	5.08	31.41
TC-40	105.67	9.98	84.85	16.64	220.94	235.67	0.91	1.13	1.39	0.25	6.05	33.84
TC-41	94.56	6.85	69.42	10.98	156.75	183.82	0.74	0.95	0.69	0.15	3.63	29.13
TC-42	77.98	10.42	66.67	10.74	192.84	182.86	0.67	0.86	1.01	0.17	4.92	25.80
TC-43	98.57	8.82	59.84	8.47	214.47	212.18	0.82	0.76	0.76	0.20	5.76	31.95
TC-44	64.89	6.07	56.67	11.54	163.47	176.12	0.68	0.67	1.22	0.18	5.43	26.49
TC-45	91.81	8.05	52.01	8.85	174.71	183.56	0.56	0.78	0.73	0.23	5.21	29.47
TC-46	79.56	9.28	71.08	9.18	185.67	226.46	0.73	0.87	0.87	0.18	4.26	28.62
TC-47	76.78	7.98	52.67	12.61	148.56	185.67	0.49	0.92	1.12	0.19	5.19	31.96
TC-48	99.10	9.58	69.91	8.79	169.08	163.56	0.68	0.83	0.75	0.19	6.19	24.47
C.D.	3.841	0.352	2.637	0.469	8.546	8.049	0.029	0.035	0.039	0.008	0.24	1.284
SE(m)	1.366	0.125	0.938	0.167	3.039	2.862	0.010	0.013	0.014	0.003	0.08	0.456

Phe.- Phenol (mg/g), Pro.- Protein content (%), Car.- Carbohydrate content (mg/g), Fla.- Flavonoids content (mg/g), Tan.- Tannin content (mg/g), AO- Anti-oxidant content (%), N-Nitrogen (%), P-Phosphorus (%), K-Potassium (%), Ca-Calcium (%), Mg-Magnesium (%), Fe-Iron (ppm)

Table 4. Biochemical characterization of *Tinospora cordifolia* from diverse accessions

Accession	Zn (ppm)	Mn (ppm)	Cu (ppm)	Fat (%)	Sap. (%)	Ca. Gl. (%)	Re. Su. (%)	Fib. (%)	Ene. (Kcal/100g)	Vit. C (mg/100g)	Vit. A (mg/100g)
TC-01	9.48	10.57	5.11	4.98	2.34	1.42	0.13	11.32	88.60	56.03	0.30
TC-02	8.72	9.32	3.84	6.73	2.59	0.58	0.15	10.91	86.44	61.25	0.23
TC-03	10.54	8.52	4.99	5.49	2.10	0.26	0.19	11.18	81.39	49.69	0.28
TC-04	6.26	12.99	5.52	6.56	2.63	1.86	0.20	11.86	82.68	52.89	0.36
TC-05	8.13	14.82	5.87	7.17	2.32	1.57	0.10	10.36	85.27	65.64	0.41
TC-06	7.67	12.59	4.51	5.72	2.28	0.30	0.09	12.11	79.66	57.57	0.22
TC-07	12.22	9.62	3.99	7.51	2.05	0.87	0.12	10.76	94.34	59.56	0.25
TC-08	9.88	9.66	4.62	4.49	2.95	1.02	0.17	11.59	84.58	50.76	0.43
TC-09	7.56	12.92	4.86	7.79	2.46	1.98	0.11	12.02	95.65	66.18	0.32
TC-10	7.06	9.82	3.76	2.62	2.60	2.05	0.18	11.00	93.47	55.89	0.19
TC-11	8.92	9.88	4.62	4.49	2.30	0.85	0.15	12.93	96.55	53.60	0.36
TC-12	9.56	13.56	4.26	4.83	2.20	0.23	0.19	10.21	90.73	64.58	0.30
TC-13	7.98	14.86	3.28	3.97	2.00	1.34	0.22	11.25	89.60	55.32	0.31
TC-14	8.57	10.56	3.82	6.47	3.19	2.00	0.26	10.83	87.22	61.16	0.23
TC-15	11.54	9.97	4.39	6.49	1.99	1.44	0.21	11.25	80.45	48.75	0.28
TC-16	7.42	14.91	3.82	6.36	3.65	1.35	0.17	11.94	83.18	54.32	0.37
TC-17	12.85	15.67	4.86	4.42	2.45	1.76	0.14	10.29	84.67	64.66	0.41
TC-18	8.88	10.56	3.69	8.36	2.90	1.19	0.20	12.10	79.70	56.32	0.22
TC-19	10.56	12.48	3.22	8.40	3.05	0.34	0.13	10.52	92.64	59.26	0.25
TC-20	15.67	16.49	5.76	3.46	3.88	1.30	0.15	11.54	83.73	53.09	0.41
TC-21	10.85	15.52	5.63	7.44	3.13	0.98	0.19	11.80	92.13	65.49	0.32
TC-22	13.67	17.76	4.96	4.54	2.22	2.27	0.20	12.92	94.87	56.31	0.19
TC-23	9.32	15.50	5.17	7.68	0.98	1.41	0.10	12.54	97.59	54.48	0.37
TC-24	11.84	16.06	3.49	5.96	2.27	0.57	0.09	10.32	89.37	64.21	0.29
TC-25	11.56	14.31	4.09	1.92	2.64	0.25	0.12	11.11	88.93	55.79	0.29

TC-26	13.38	16.91	4.42	3.97	2.08	1.85	0.18	10.66	86.25	61.62	0.22
TC-27	8.42	14.82	5.03	7.74	2.66	1.53	0.11	11.40	85.13	50.40	0.28
TC-28	9.86	11.56	4.63	6.56	2.31	1.43	0.19	11.89	84.17	52.53	0.34
TC-29	8.74	13.34	3.82	8.17	2.28	0.59	0.15	10.45	85.42	66.95	0.43
TC-30	13.30	16.59	6.19	8.81	2.15	0.26	0.19	12.09	79.50	57.15	0.22
TC-31	12.02	9.49	5.62	7.47	2.92	1.90	0.13	10.94	93.77	60.75	0.25
TC-32	14.47	8.57	4.96	7.21	2.47	1.58	0.14	11.80	85.21	49.01	0.43
TC-33	11.57	13.52	3.98	6.36	2.62	1.42	0.19	12.31	92.34	68.96	0.44
TC-34	10.42	9.99	5.62	6.28	2.25	0.58	0.20	10.90	91.60	54.21	0.19
TC-35	6.75	8.98	5.06	8.44	2.19	0.27	0.10	13.22	99.66	53.89	0.34
TC-36	8.63	10.46	5.36	7.80	2.39	1.83	0.09	10.55	90.28	66.07	0.30
TC-37	6.92	14.96	3.86	2.18	2.61	1.60	0.12	11.60	87.27	56.98	0.30
TC-38	8.56	11.84	5.09	3.39	2.11	1.41	0.17	11.24	85.86	60.97	0.24
TC-39	8.56	13.56	3.43	8.56	2.64	2.00	0.11	10.89	78.58	49.93	0.28
TC-40	13.37	16.36	5.43	5.64	2.36	1.49	0.17	11.75	80.70	51.82	0.36
TC-41	8.56	11.41	4.56	6.32	2.26	1.37	0.15	10.34	85.72	65.31	0.40
TC-42	11.86	13.86	3.85	4.49	1.98	1.75	0.19	12.13	79.78	59.24	0.22
TC-43	9.86	14.82	4.42	6.47	2.97	1.18	0.13	10.82	96.60	58.67	0.25
TC-44	8.48	13.86	3.62	7.37	2.45	1.99	0.15	11.43	84.79	50.18	0.32
TC-45	6.83	14.71	5.63	7.92	2.66	1.37	0.19	11.95	94.48	64.10	0.32
TC-46	7.67	12.56	4.82	6.37	2.31	1.33	0.20	11.07	93.94	57.15	0.18
TC-47	12.67	12.58	3.93	2.96	2.18	1.77	0.10	13.03	92.40	52.42	0.36
TC-48	9.74	11.47	5.42	8.56	2.36	1.19	0.09	9.75	92.54	63.47	0.31
C.D.	0.414	0.592	0.183	0.245	0.094	0.066	0.008	0.509	3.628	2.639	0.013
SE(m)	0.147	0.210	0.065	0.087	0.034	0.024	0.003	0.181	1.290	0.938	0.004

Zn- Zinc (ppm), Mn- Manganese (ppm), Cu- Copper (ppm), Fat content (%), Sap- Saponin (%), Ca. Gl.- Cardiac Glycosides (%), Re. Su.- Reducing sugar (%), Fib.- Fibre (%), Energy (Kcal/100g), Vit. C- Vitamin C (mg/100g), Vit. A- Vitamin A (mg/100g).

Conclusion

The current investigation reveals high degree of variability in biochemical constituents of *Tinospora* which varied among locations. Significant diversity was seen in terms of the qualitative indices. Collections with superior characters were identified viz. TC-22 (Nemili, Ranipet district) recorded the highest phenol (122.73 mg/g), flavonoid (18.84 mg/g), nitrogen (1.06%), phosphorus (1.17%), magnesium (7.55%) manganese (17.76 ppm) and cardiac glycosides (2.27%) contents. TC-20 (Thovalai, Kanyakumari district) showed significantly higher protein (13.85%), carbohydrate (86.92mg/g), antioxidant activity (261.74 %), potassium (1.45%), calcium (0.35%) and zinc (15.67 µg/g). TC-30 (Acharavakkam, Chengalpattu) recorded maximum tannin (234.56 mg/g), copper (6.19µg/g) and fat (8.81%) contents. TC-35 (Courtallam, Tenkasi district) was identified as higher fibre (13.22%) and energy rich (99.66 Kcal/100g). Maximum vit. C (68.96 mg/100g) and vit. A (0.44 mg/100g) were observed in TC 33 (Tindivanam, Villupuram). The findings indicates that ecological condition under which *Tinospora* is grown determines its biochemical and nutritional quality.

References

- Ahmed S M, Manhas L R, Verma V & Khajuria R K 2006 Quantitative determination of four constituents of *Tinospora* sps. by a reversed-phase HPLC-UV-DAD method. Broad-based studies revealing variation in content of four secondary metabolites in the plant from different eco-geographical regions of India. J. Chromatogr. Sci.. 44(8): 504-509.
- AOAC 1990 Official Methods of Analysis, 15th edn. Association of Official Analytical Chemists, Arlington, VA
- Aranha I, Clement F & Venkatesh YP 2012 Immunostimulatory properties of the major protein from the stem of the Ayurvedic medicinal herb, guduchi (*Tinospora cordifolia*). J. Ethnopharmacol . 139(2): 366-372.
- Armstrong M J, & Kirkby E A 1979 Estimation of potassium recirculation in tomato plants by comparison of the rates of potassium and calcium accumulation in the tops with their fluxes in the xylem stream. Plant Physiol. 63 (6): 1143- 1148.
- Basai 2023 <https://basai.org/wp-content/uploads/2023/08/Cultivation-and-Export-of-Medicinal-Plants.pdf> . Accessed 03 December 2024
- Bhandal I S & Malik C P 1988 Potassium estimation, uptake, and its role in the physiology and metabolism of flowering plants. In International review of cytology (Vol. 110, pp. 205-254). Academic Press.
- Bradford M M 1976 A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. Anal. Biochem. 72(1-2): 248-254.
- Chang C C, Yang M H, Wen H M & Chern J C 2002 Estimation of total flavonoid

- content in propolis by two complementary colorimetric methods. *J. Food Drug Anal.* 10(3): 178-182
- Choudhry N, Singh S, Siddiqui M B & Khatoon S 2014 Impact of seasons and dioecy on therapeutic phytoconstituents of *Tinospora cordifolia*, a Rasayana drug. *Biomed Res. Int.*, 2014:1-11.
- David D J 1958 Determination of zinc and other elements in plants by atomic-absorption spectroscopy. *Analyst.* 83(9 93): 655-661.
- Energy F F 2003 Methods of analysis and conversion factors food and paper nutrition paper. Food and Agriculture Organization: Rome, Italy.
- Gahlaut A, Gothwal A & Dabur R 2012 TLC based analysis of allelopathic effects on tinosporoside contents in *Tinospora cordifolia*. *J. Chem. Pharm. Res.* 4: 3082-3088.
- Garg A N, Singh R, Maharia R S, Dutta R K & Datta A 2022 Quantification of minor, trace and toxic elements in stems of *Santalum album* (L.), *Mangifera indica* (L.) and *Tinospora cordifolia* by instrumental neutron activation analysis. *J. Plant Sci. Phytopathol.* 6: 8-14.
- Gupta R & Sharma V 2011 Ameliorative effects of *Tinospora cordifolia* root extract on histopathological and biochemical changes induced by aflatoxin-B1 in mice kidney. *Toxicol. Int.* 18(2): 94-98.
- Hassan A S & Hassan H S 2008 Quantitative estimation of vitamin C in some local fruits. *Sci. World J.*, 3(2): DOI:10.4314/swj.v3i2.51805
- Ingberman A K 1958 Functionality of Phenols by Bromination. *Anal. Chem.*, 30(5): 1003-1004.
- Jones Jr J B 1991 Kjeldahl method for nitrogen determination. Micro-Macro Publishing, Inc.
- Khan I M, Sri Harsha P S C , Giridhar P & Ravishankar G A 2011 Pigment identification, antioxidant activity, and nutrient composition of *Tinospora cordifolia* (Willd.) Miers ex Hook. f & Thoms fruit. *Int. J. Food Sci. Nutr.* 62(3): 239-249.
- Khan T A, Ipshita A H, Mazumdar R M, Abdullah A T M, Islam G M R & Rahman M M 2020 Bioactive polyphenol profiling and *invitro* antioxidant activity of *Tinospora cordifolia* Miers ex Hook F and Thoms: a potential ingredient for functional food development. *Bangladesh J. Sci. Ind. Res.* 55(1): 23-34.
- Kirk P L 1950 Kjeldahl method for total nitrogen. *Anal. chem.* 22(2): 354-358.
- Koleva I I, Van Beek T A, Linssen J P, Groot A D & Evstatieva L N 2002 Screening of plant extracts for antioxidant activity: a comparative study on three testing methods. *Phytochem Anal.* 13(1): 8-17.

- Kumar A, Kumar M, Dandapat S & Sinha M P 2013 Antioxidant activity and pharmacological screening of *Tinospora cordifolia* (Thunb.). The Bioscan. 8(2): 689-693
- Lopez-Teros V, Ford J L, Green M H, Tang G, Grusak M A, Quihui-Cota L, & Astiazaran-Garcia H 2017 Use of a “super-child” approach to assess the vitamin A equivalence of *Moringa oleifera* leaves, develop a compartmental model for vitamin A kinetics, and estimate vitamin A total body stores in young Mexican children. J. Nutr. 147(12): 2356-2363.
- Majed N, Li Y & Gu A Z 2012 Advances in techniques for phosphorus analysis in biological sources. Curr. Opin. Biotechnol. 23(6): 852-859.
- McDonald S, Prenzler P D, Antolovich M & Robards K 2001 Phenolic content and antioxidant activity of olive extracts. Food Chem. 73(1): 73-84.
- Modi B, Kumari Shah K, Shrestha J, Shrestha P, Basnet A, Tiwari I & Prasad Aryal S 2020 Morphology, Biological Activity, Chemical Composition, and Medicinal Value of *Tinospora cordifolia* (Willd.) Miers. Adv. J. Chem. B 3(1): 36-53.
- NMPB <https://www.nmpb.nic.in>. Accessed 03 December 2024.
- Obadoni B O & Ochuko P O 2002 Phytochemical studies and comparative efficacy of the crude extracts of some haemostatic plants in Edo and Delta States of Nigeria. Global J. Pure Appl. Sci. (2): 203-208.
- Premanath R & Lakshmidevi N 2010 Studies on anti-oxidant activity of *Tinospora cordifolia* (Miers.) leaves using in vitro models. Am. J. Sci. 6(10): 736-743.
- Puratchimani V & Jha S 2007 HPTLC standardization of *Tinospora cordifolia* using tinosporaside. Indian J. Pharm. Sci. 69(4): 578-581.
- Sahrawat K L 1987 Determination of calcium, magnesium, zinc and manganese in plant tissue using a dilute HCl extraction method 1. Commun. Soil Sci. Plant Anal. 18(9): 947-962.
- Shervani Z A & Mishra P K 2017 Phytochemical study of *Tinospora cordifolia* grown on three different soil conditions. Res. J. Life Sci. Bioinform. Pharm. Chem. Sci. 5: 810-815.
- Shivraj H & Khobragade C N N 2009 Determination of nutritive value and mineral elements of some important medicinal plants from western part of India. J. Med. Plants. 8(29): 79-88.
- Shukla S & Gardner J 2006 Local knowledge in community-based approaches to medicinal plant conservation: lessons from India. J. Ethnobiol. Ethnomed. 2: 1-9.
- Solich P, Sedliakova V & Karlicek R 1992 Spectrophotometric determination of cardiac glycosides by flow-injection analysis. Analytica. Chimica. Acta. 269(2): 199-203.
- Srinivasan G V, Unnikrishnan K P, Shree A R & Balachandran I 2008 HPLC

- estimation of berberine in *Tinospora cordifolia* and *Tinospora sinensis*. Indian J. Pharm. Sci. 70(1): 96-99.
- Stien L H, Kiessling A & Manne F 2007 Rapid estimation of fat content in salmon fillets by colour image analysis. J. Food Compos. Anal. 20(2): 73-79.
- Talbot G H, Bradley J, Edwards J E, Gilbert D, Scheld M & Bartlett J G 2006 Erratum: Bad bugs need drugs: An update on the development pipeline from the Antimicrobial Availability Task Force of the Infectious Diseases Society of America (Clinical Infectious Diseases (March 1, 2006) 42 (657-668)). Clin. Infect. Dis. 42(7): 1065.
- Tanwar S I, Jain J, Verma S & Solanki D 2012 Standardization and phytochemical evaluation of *Tinospora cordifolia* (Willd.) Miers.(Menispermaceae). Int. J. Pharm. Pharm. Sci. 4: 219-223.
- Thomas W & Dutcher R A 1924 The colorimetric determination of carbohydrates in plants by the picric acid reduction method 1. The estimation of reducing sugars and sucrose 1. J. Am. Chem. Soc. 46(7): 1662-1669.
- Tinggi U, Reilly C & Patterson C 1997 Determination of manganese and chromium in foods by atomic absorption spectrometry after wet digestion. Food Chem. 60(1): 123-128.
- Tyagi P & Chauhan A K 2020 Optimization and characterization of functional cookies with addition of *Tinospora cordifolia* as a source of bioactive phenolic antioxidants. LWT, 130: 109639.
- Uadia N R, Ugwu I, Eramah T & Osunde E 2017 Estimation of tannins, alkaloids, saponins and proximate composition of *Vernonia amygdalina* (Del) root. Int. J. Herb. Med. 5(3): 88-92.
- Upadhyay N, Ganie S A, Agnihotri R K & Sharma R 2014 Free radical scavenging activity of *Tinospora cordifolia* (Willd.) Miers. J. Pharmacogn. Phytochem. 3(2): 63-69.
- Van Soest P J & McQueen R W 1973 The chemistry and estimation of fibre. Proc. Nutr. Soc. 32(3): 123-130.
- Yadav R N S & Agarwala M 2011 Phytochemical analysis of some medicinal plants. J. Phytol. 3(12): 10-14
- Yemm E W & Willis A 1954 The estimation of carbohydrates in plant extracts by anthrone. Biochem. J. 57(3): 508-514.