



ISSN: 2455-0485

Antiviral medicinal plants of India as a potential tool against COVID-19: A review with ethno scientific evidence

D. Mishra¹, A. Kumar², A. Tiwari¹, Deepika¹, P. Chaturvedi^{1,*}

¹Department of Biological Sciences, College of Basic Sciences and Humanities, G.B. Pant University of Agriculture and Technology, Pantnagar-26314, Uttarakhand, India, ²Department of Chemistry, College of Basic Sciences and Humanities, G.B. Pant University of Agriculture and Technology, Pantnagar-26314, Uttarakhand, India

ABSTRACT

Indian traditional medicinal systems are one of the oldest therapeutic systems in the world. Medicinal and aromatic plants play a dominant role in Indian traditional medicinal systems. Traditionally, many medicinal plants are used in India for their therapeutic relevance so much so that they have acquired a significant role in Indian religion as well. Many of these plants have proven antiviral effects. This review documents up-to-date information about many such medicinal herbs used in India which have got pharmacological significance in fighting viral infections. These plants surely have the potential to provide protection against Covid-19. The review presents a list of such plants along with their chemical ingredients and possible modes of action against the respective viral diseases. All information has been obtained by consulting the databases of Scopus, PubMed, Science Direct, Elsevier, Springer and relevant research papers and reports on COVID-19. The cited medicinal plants are used extensively in India as herbal remedies. The use of these plants is validated in light of research papers citing their ethnobotanical uses, important active principles and modes of action of the of medicinally important natural products. The plants listed have great potential to fight COVID-19 and other viral infections. Many of these are immunity boosters providing strength to the body to control the onset of diseases.

Received: October 20, 2022
Revised: December 05, 2022
Accepted: December 08, 2022
Published: March 23, 2023

*Corresponding author:
P. Chaturvedi
E-mail: an_priti@yahoo.com

KEYWORDS: COVID-19, Medicinal herbs, Viral diseases, Herbal remedy, Immunity boosters, Active principle

INTRODUCTION

While medicinal plants have been used to treat viral infections in the past, the effort of the Boots drug company (Nottingham, England) to screen 288 plants for anti-influenza activity was the first recognized step in the production of antiviral agents (Chantrill *et al.*, 1952). Since then, more than 90 drugs from various functional groups have been approved for the treatment of viral infections, including Human Immunodeficiency virus (HIV), Hepatitis B virus (HBV), Hepatitis C virus (HCV) and influenza virus (DeClercq & Li, 2016). The production of safe and novel antiviral drugs has been necessitated by the high prices and adverse side effects of synthetic drugs. Herbal extracts have emerged as a novel option for the development of antiviral drugs that target various stages of the virus replication cycle. Many of the currently available medicines for microbial infections are derived from plants. One-fourth of the widely used medicines contain at least one active compound derived from plants (Farnsworth & Soejarto, 1991). Ethnobotanical sources also corroborate the fact that the various traditional medicines used in India are mainly obtained from medicinal plants (Ahvazi *et al.*, 2007).

Introduction to Coronavirus

Coronavirus has been described as a pathogen that causes disease and death as a result of massive alveolar damage and progressive respiratory failure. According to the WHO report, Coronavirus pandemic has globally affected more than 100 million people and killed more than 2904686 people as of 8th April 2021. Coronavirus disease, 2019 has been abbreviated and named as COVID-19 by WHO on 11th February 2020. A simple Google search suggests there is an upsurge in the number of Coronavirus related papers (almost 30.44 times) published in scientific journals in 2020 (Figure 1). This pandemic has caused havoc worldwide, affecting almost more than half of the entire human world population due to its contagious nature and high transmission rate.

Coronavirus belongs to the Coronaviridae family of Nidovirales order. Coronavirus (CoV) are microscopic virus particles with an encapsulated positive single-stranded RNA of 26-32 kb (largest RNA genome), infecting humans as well as animals. It is divided into alpha, beta, gamma and delta subgroups and

Copyright: © The authors. This article is open access and licensed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0/>) which permits unrestricted, use, distribution and reproduction in any medium, or format for any purpose, even commercially provided the work is properly cited. Attribution — You must give appropriate credit, provide a link to the license, and indicate if changes were made.

almost all the known human targeting CoVs belong to either the alpha subgroup i.e., Human CoV-NL63 and CoV-229E or the beta subgroup i.e., Human CoV-OC43, HKU1 (Human Coronavirus), SARS-CoV (Severe acute respiratory syndrome-coronavirus), MERS-CoV (Middle East respiratory syndrome-coronavirus) and the recently emerging SARS-CoV-2 (Lu et al., 2015). In 2002-2003, the first ‘Severe Acute Respiratory Syndrome’ (SARS-CoV) epidemic was caused by Coronavirus in Guangdong, China and approximately 10 years later ‘Middle East Respiratory Syndrome’ (MERS-CoV) outbreak occurred in the Middle East in 2012. Further, by the end of 2019, Wuhan in China witnessed the flare-up of novel Coronavirus (nCov-19) which has been currently keeping the world on its toes with its global outbreak in 2020. The International Committee on Taxonomy of Viruses (ICTV) named it as SARS-CoV-2 (Zhang et al., 2020).

Origin and Transmission

The initial outbreak of coronavirus reported from the Hunan market of Wuhan in China leads to the presumption of an intermediate animal source for respective viral attacks in humans (Andersen et al., 2020). Bats are the reservoir of many viruses including SARS-CoV and MERS-CoV (Hu et al., 2015). It was observed that the genetic constitution of SARS-CoV-2 is similar to CoV RATG of bats by 96.2% and 79.5% identical to SARS-CoV (Guo et al., 2019). Although RATG₁₃ is approximately 96% similar to SARS-CoV-2, divergence in the receptor binding domain of the spike leads to inefficacy in binding to human Angiotensin-converting enzyme 2 (ACE2) receptors present in the cells of the respiratory tract (Andersen et al., 2020).

Clinical Symptoms

Symptoms of Coronavirus differ from one person to another. Probably, depending on the immunity and medical history, a person may show mild to severe symptoms proving to be fatal or asymptomatic (Guan et al., 2020). The common symptoms of COVID-19 include fever, sore throat, dry cough, muscle pain, and shortness of breath; sometimes, symptoms like nausea and diarrhea are also observed. Severe symptoms lead to lung and kidney damage and can prove to be fatal (Chen et al., 2020). The incubation period of the virus is 2-14 days after being exposed to the virus. Older people along with people suffering from diabetes, cardiovascular disease, respiratory problems and other medical problems tend to develop more severe symptoms (By CDCP 2019).

Structure of Coronavirus

Coronaviruses are encapsulated particles with a central core of nucleocapsid proteins enclosing a single-stranded positive RNA and the surface envelope embedded with ‘spikes’ i.e., glycoprotein projections. The spikes play an important role in attachment to the host cell receptors (Tyrrell & Myint, 1996). In addition to receptor binding, spikes play an important role in virus-host membrane integration, antibody neutralization

and mediating entrance of viruses into the host. The spike glycoprotein is trimeric in nature with a single unit of S protein (Ou et al., 2020). The receptor-binding domain (RBD) of S protein binds to the human angiotensin-converting enzyme (hACE2) (Wrapp et al., 2020) present in epithelial cells of lung alveoli and noticed in absorptive cells lining the small intestine as well (Hamming et al., 2004).

Comparison of SARS-CoV and SARS-CoV-2 also implies that the latter is evolved to bind more strongly to the ACE2 receptor. The receptor binding domain has conserved regions of antigen for antibodies to interact and thus serves as a potential target for cross-reactive antibody therapy against coronavirus (Lan et al., 2020). Protease, papain-like protease (PLpro) from the coronavirus causes severe acute respiratory syndrome (SARS-CoV). This is a unique protease because it is not only responsible for processing the viral poly protein into its functional units but it is also capable of cleaving ubiquitin and ISG15 conjugates and plays a significant role in helping SARS-CoV evade the human immune system (Ratia et al., 2008). SARS-CoV-2 shows 77.5% and 50% sequence homology with SARS-CoV and MERS-CoV respectively (Kim et al., 2020). Due to these similarities, medicinal plants and their derivative compounds which have been used to treat SARS-CoV and MERS-CoV can also be used as potent inhibitors of SARS-CoV-2. Another important aspect of the Coronavirus which may serve as the most probable drug target is the Main Protease, also called 3CLpro. Upon transcription, the coronavirus genome produces a polypeptide that undergoes enzymatic cleavage to result in a non-structural protein constituting the viral replication machinery. 3-chymotrypsin like protease (3CLpro) produces its cleavage activity at 11 different sites of the polypeptide (Anand et al., 2003).

With the main protease being the major drug target so far and along with the current development and clinical trials of a drug or vaccine, the best way to cope with the nCoV-19 without any major side effects is to rely on the medicinal plants which constitute a holistic effect of various compounds from boosting immunity to treating infections. Thus, medicinal plants can serve as a potential source to deal with this novel coronavirus due to active compounds like flavonoids such as quercetin which can degrade dengue virus envelope protein (Mir et al., 2016), protease inhibiting compounds (Majumdar, 2013) which may inhibit 3CLpro, terpenoids etc. and show effective antiviral activity (UlQamar et al., 2016). Also, the compounds obtained from medicinal plants against nCoV-19 can be used as a potential source to design a virus-targeting drug. In this review, we focus on the phytochemicals derived from various medicinal plants which are used in India as antiviral agents that can also serve as the potential therapeutic source against novel coronavirus.

Natural Medicine to Control Virus

In the development of human civilization, medicinal plants always played a critical role. Plants have been used for thousands of years to prevent epidemics, treat health disorders and control diseases besides providing a variety of nutritional foods. The

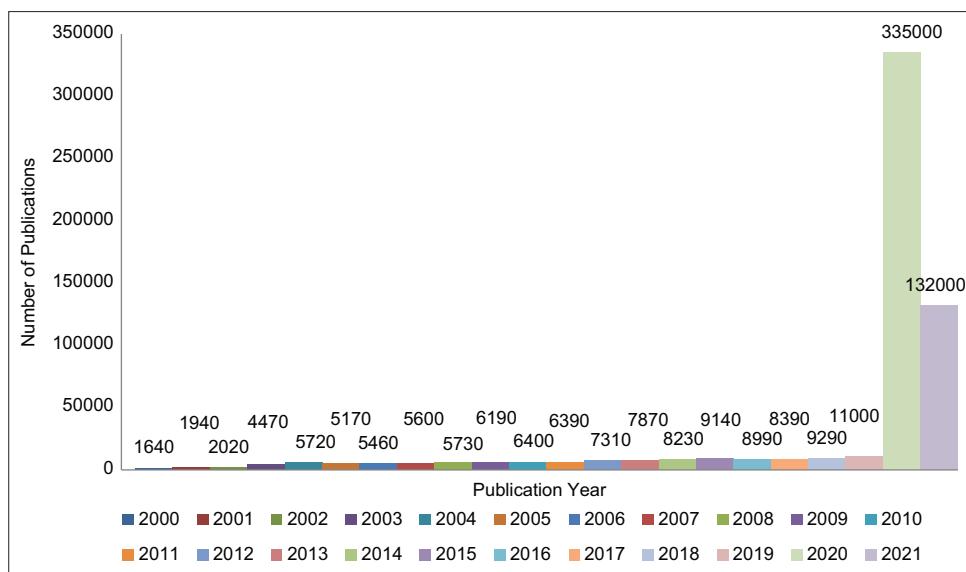


Figure 1: Year-wise publications in scientific journals once “Coronavirus” was entered as a keyword (In total, 187810 articles were found collectively)

two important traditional systems of medicine, Traditional Indian Medicine (TIM) i.e., Ayurveda, and Traditional Chinese Medicine (TCM) system are used extensively for the treatment of diseases (Patwardhan *et al.*, 2005). All 6 systems of medicine that are recognized in India, *viz.*, Ayurveda, Yoga, Siddha, Unani, Naturopathy and Homeopathy, depend a lot on the use of medicinal plants and their chemical components (Ravishankar & Shukla, 2007). Among all these systems, medicinal plants are traditionally being used as primary health care against many diseases. Hence, medicinal plants are very rightly called as “backbone of traditional medicine.” About 3.3 million people utilize medicinal plants in less developed countries on a regular basis (Davidson–Hunt, 2000). Plants-based therapies rely on different kinds of secondary metabolites produced by the plants which are responsible for their biological activities. These metabolites possess antiviral, antibacterial, antioxidant, anti-inflammatory and antispasmodic effects. Table 1 enlists several vegetables, fruits and medicinal plants commonly used in India for antiviral effects with Solanaceae and Zingiberaceae contributing a maximum number of antiviral plants followed by Euphorbiaceae, Acanthaceae, Rutaceae, Fabaceae and Lamiaceae (Figure 2). Many of these plants are also used as ethnomedicine in different parts of the world (Table 2).

The compound called withanone isolated from *Withania somnifera* (L.) Dunal may inhibit novel Coronavirus-19 by disrupting the interaction between host ACE 2 and viral spike-protein Receptor Binding Domain (RBD) (Varshney *et al.*, 2020). Another compound Withaferin A (WA), an active compound isolated from *W. somnifera* act against H1N1 influenza (Cai *et al.*, 2015). This compound also inhibits herpes simplex virus (HSV) (Grover *et al.*, 2011). Roots of *W. somnifera* are also used against β -amyloid (1–42)-induced cytotoxicity in HIV-1Ba-L (clade B) infection (Kurapati *et al.*, 2016).

Similarly, *Ocimum sanctum* L. and *Azadirachta indica* A. Juss., the two high-value important medicinal plants have exceptionally

significant antimicrobial activity. Due to their excessive use in Indian systems of medicine, these plants have acquired very high religious significance in Hinduism, the age-old religion in India. Natural compounds from *O. sanctum* (Tulsi) and *A. indica* (Neem) have high binding efficacy against SARS-CoV-2 targets (Kumar, 2020). Oleanolic acid, methyl eugenol and ursolic acid show high binding potential with surface spike glycoprotein and RNA polymerase in SARS-CoV-19. Epoxy-azadiradione, gedunin show efficacy against m-protease of SARS-CoV-19. Active compounds from *O. sanctum* also inhibit replication and protein synthesis in the Swine flu H1N1 virus (Joshi, 2014). Tulsi strengthens the immune response by enhancing both cellular and humoral immunity by boosting the cell-mediated immune responsiveness and gamma-aminobutyric acid (GABA) (Mukherjee *et al.*, 2005). Eugenol and other essential oils from *O. sanctum* considerably improve immune response due to immuno-stimulating effects (Sen, 1993). *A. indica* bark is known to block the entry of Herpes Simplex Virus (HSV) (Tiwari *et al.*, 2010). Two compounds from neem namely, rutin and tiplasinin have been used against influenza strains (Ahmad *et al.*, 2015).

Tinocordiside, berberine, magnoflorino and isocolumbinare the naturally occurring compounds isolated from *Tinospora cordifolia* (Willd.) Miers (Giloy) have the potential to inhibit the 4 key targets (surface glycoprotein, receptor binding domain, RNA-dependent RNA polymerase and main protease) in SARS-CoV-2 (Sagar & Kumar, 2020). Active compounds isolated from Giloy can also be used against Herpes Simplex Virus-1 (HSV-1) (Pruthvish & Gopinatha, 2018). Giloy was also used as a potent inhibitor against H1N1 flu due to its immunostimulant property (Yasmin *et al.*, 2020). Besides the above-mentioned compounds, agglutins from *Nicotiana tabacum* L. (tobacco), *Urtica dioica* L. (stinging needle) and *Morus nigra* L. (Black mulberry) are also reported to inhibit viral attachment in case of SARS-CoV (Keyaerts *et al.*, 2007).

Table 1: Plants with antiviral potential

S. No.	Vernacular Name	Scientific Name	Family	Key compound	Viral disease	Mode of action	References
1	Bhui Amla	<i>Phyllanthus amarus</i> Schumach. & Thonn.	<i>Phyllanthaceae</i>	Quercetin, Ellagicacid, Rutin, luteolin, kaempferol, quinic acid, andursolic acid	AIDS Hepatitis B Hepatitis C	Inhibit HIV replication (blocking the interaction of HIV-1 gp120 with its primary cellular receptor CD4) Inhibition of mRNA transcription	Notka et al., 2004 Lee et al., 1996
2	Chankapiedra	<i>Phyllanthus niruri</i> L.	Euphorbiaceae	Nirurin, ent-norsecurin, Quercetin, rutin, astragalin, isoquercitrin, kaempferol-4-rhamnopyranoside	Hepatitis B	Inhibits DNA polymerase, binds to the surface antigen of hepatitis B virus	Ravikumar et al., 2011 Venkateswaran et al., 1987
3	Amla	<i>Embllica officinalis</i> Gaertn.	Euphorbiaceae	EGCG, theaflavin digallate, resveratrol (RV), hydroxytyrosol	Influenza A	Reduction of plasma membrane, accumulation of nucleoprotein (NP) at the late stage of the replication cycle	Liu et al., 2011
4	Kalmegh	<i>Andrographis paniculata</i> (Burm. f.) Nees	Acanthaceae	Andrographolide, neoandrographolide	Dengue fever Herpes simplex virus 1 Influenza A	Inhibit replication and pyrimidine biosynthesis Not determined (ND)	Tang et al., 2012 Wiant et al., 2005 Chen et al., 2009
5	Haldi	<i>Curcuma longa</i> L.	Zingiberaceae	Curcumin	HIV Influenza Herpes simplex virus (1&2) Coxsackievirus Hepatitis B Human papilloma virus (HPV) Japanese encephalitis virus (JEV) Severe acute respiratory syndrome (SARS)	Inhibition of HIV-1 and HIV-2 proteases Inhibition of haemagglutination Reduction of replication Replication inhibition through UPS dysregulation Suppression of HBV replication by increasing the p53 level Inhibition expression of viral oncoproteins of E6 and E7 Reduction in production of infective viral particles Inhibition expression of Mpro protease	Sui et al., 1993 Chen et al., 2010 Kutluay et al., 2008; Zandi et al., 2010; Si et al., 2007 Kim et al., 2009 Divya & Pillai, 2006 Dutta et al., 2009 Kim et al., 2008
6	Indian saffron	<i>Curcuma zanthorrhiza</i> Roxb.	Zingiberaceae	Demethoxycurcumine,	Hepatitis B	Reduction of replication	Devaraj et al., 2010
7	White Himalayan Rue	<i>Boenninghausenia albiflora</i> (Hook.) Rchb. ex Meisn.	Rutaceae	Leptodactylone, rutarenzin, rutamarin	SARS-CoV Kaposi's sarcoma-associated herpesvirus (KSHV)	Not determined Replication inhibition	Yang et al., 2007 Xu et al., 2014
8	Nargis	<i>Narcissus tazetta</i> L.	Amaryllidaceae	<i>Narcissus tazetta</i> lectin (NTL) protein	Influenza A and B	Replication inhibition	Ooi et al., 2010
9	Orchid	<i>Listera ovata</i> (L.) R.Br.	Orchidaceae	Lectins-mannose-specific protines	HIV SARS-CoV	Inhibit virus cell binding ND	van Damme et al., 1994 Keyaerts et al., 2007
10	Adrak	<i>Zingiber officinale</i> Rosc.	Zingiberaceae	Gingerol, β -sitosterol shogaol, dehydrogingerdione	Chikungunya HIV COVID-19	Replication inhibition Inhibiton of Reverse Transcriptase (RT) enzyme Inhibition expression of Mpro protease	Kaushik et al., 2020 Kharisma et al., 2018 Rathinavel et al., 2020

(Contd...)

Table 1: (Continued)

S. No.	Vernacular Name	Scientific Name	Family	Key compound	Viral disease	Mode of action	References
11	Giloy	<i>Tinospora sinensis</i> (Lour.) Merr.	Menispermaceae	Berberine, Isocolumbin, Magnoflorine Tinocordiside	SARS-CoV-2	Inhibition expression of viral protease (6Y84) and RNA dependent RNA polymerase (6M71)	Sagar et al., 2020
					Influenza A	Inhibition expression of viral protease	Saikia et al., 2019
					HIV	ND	Gupta et al., 2010
12	Mulathee	<i>Glycyrrhiza glabra</i> L.	Fabaceae	Glycyrrhizin, Licorice, Glycyrrhizic acid	JEV	Inhibition of viral replication	Badam, 1997
					HIV	Inhibition of viral replication	De Clercq, 2000
13	Ashwagandha	<i>Withania somnifera</i> L.	Solanaceae	Withaferin-A, Withanone	Herpes Simplex type I	Inhibition of viral replication	Pompeii et al., 1979
					Herpes Simplex type I	Inhibit virus cell binding	Kambizi et al., 2007
					HIV 1 and 2	Inhibition expression of viral protein	Asres et al., 2001
14	Gheekumari	<i>Aloe vera</i> (L.) Burm.f.	Asphodelaceae	aloin A , aloin B, elgonica dimer A, feralolide, isoaloeresin D, aloeresin E	SARS-CoV-2	Inhibition expression of viral protease	Kumar et al., 2020
						Inhibition of viral replication	Zandi et al., 2007
15	Neem	<i>Azadirachta indica</i> A. Juss.	Meliaceae	kaempferol-3-Orutinoside, rutin, Dengue virus hyperoside, epicatechin, quercetin	Inhibition expression of viral protease (NS2B-NS3pro)	Dwivedi et al., 2021	
					ND		Badam et al., 1999
					Coxsackie virus B		Faccin-Galhardi et al., 2012
16	Jamun	<i>Syzygium cumini</i> (L.) Skeels	Myrtaceae	Caffeic acids, Citronellol, Eugenol, Ferulic acids, Kaempferol cichoric acid	Inhibition of replication of PV-1	Ashfaq et al., 2016	
					HCV	Inhibition expression of viral protease (NS3/4A)	
					Influenza (H5N1)	ND	Ramya et al., 2012; Sood et al., 2012
17	Purple coneflower	<i>Echinacea purpurea</i> (L.) Moench	Asteraceae	HSV-1&2 HIV	Inhibition of replication	Thompson, 1998	
					Cytochrome P450 (CYP) 3A and P-glycoprotein activity	Robinson, 1998; Penzak et al., 2010	
					Influenza A	Influence activity of Cytochrome P450 and P- Glycoprotine in human	Pleschka et al., 2009
18	Elderberry	<i>Sambucus nigra</i> L.	Adoxaceae	Kaempfero, Quercetin, Catechin, Epicatechin, Cyanidin-3-rutinoside	HIV	Inhibition expression of viral protease	Uncini et al., 2005
					Influenza A & B	Inhibit virus-cell binding	Serkedjieva et al., 1990; Roscheket al., 2009
					SARS-CoV	Inhibition of replication	Keyaerts et al., 2007
19	Tulsi	<i>Ocimum sanctum</i> L.	Lamiaceae	Eugenol, methyl eugenol, caryophyllene, cirsilineol, cirsimarin, rosmarinic, Quercetin	HSV-1 & 2	ND	Jadhav et al., 2012
					New Castle Disease virus (NDV)	ND	Jayati et al., 2013
					HSV 2	Inhibition of replication	Chiang et al., 2005
20	Chakrphool	<i>Illicium angustisepalum</i> A.C. Sm.	Schisandraceae	cis- and trans-anethole, α -pinene, farnesol, β -phellandrene, α -terpineol, Illicinone A 23-dioic acid	Newcastle disease virus (NDV)	ND	Alhajj et al., 2020
					HIV	ND	Song et al., 2007
					HSV	Inhibition of replication	Astani et al., 2011
					SARS-CoV-2	Inhibition expression of viral protease (Mpro)	Silva et al., 2020

(Contd...)

Table 1: (Continued)

S. No.	Vernacular Name	Scientific Name	Family	Key compound	Viral disease	Mode of action	References
21	Shatavari	<i>Asparagus racemosus</i> Willd.	Asparagaceae	Asparanin A, Muzanzagenin, Asparagamine A, Rutin, Shatavarin, Tinocordiside, 20-βHydroxy Ecdysone	HIV-1 SARS-CoV-2	Inhibit virus cell binding (glycoprotein gp120/41 and the cellular receptor protein CD4) Inhibition expression of viral protease	Panvilai et al., 2020 Borse et al., 2020
22	Saag	<i>Spinacia oleracea</i> L.	Chenopodiaceae	Kaempferol	SARS-CoV-2	Inhibition expression of viral protease (Mpro)	Tallei et al., 2020
23	Ban tulsi	<i>Origanum vulgare</i> L.	Lamiaceae	caffeoylequinic acids, C-glycosides, 2,5-dihydroxybenzoic acid, rosmarinic acid, luteolin	Animal herpesviruses Influenza (H1N1) Coxsackie virus B3	ND ND	Fernandes et al., 2012 Vimalanathan & Hudson, 2012 Zhang et al., 2014
24	Pyaz	<i>Allium cepa</i> L.	Liliaceae	Quercetin Hexadecanoic acid Tricosane Isopropylidithioisopropane	HSV-1 Adenoviruses (ADV)	Inhibition expression of viral protease ND	Romeilah et al., 2010 Chen et al., 2011
25	Shimla mirch	<i>Capsicum annuum</i> L.	Solanaceae	β-ionone camphor thymol benzoic acid	Newcastle disease virus (NDV)	ND	Eggink et al., 2012; Hanif et al., 2016;
26	Sweet orange	<i>Citrus sinensis</i> (L.) Osbeck	Rutaceae	Myricetin, Naringenin, Naringin, coronavirus (CoV) Neryl-acetate, Pcmene, Quercetin, Rutin, Scutellarein, Stigmastanol, Subaphyllin	Inhibition of replication and expression of TRP genes	Sathy & Gopalakrishnan, 2013; Ulasli et al., 2014	
27	Kushmul	<i>Berberis lycium</i> Royle	Berberidaceae	Baberine Chenabine, Diphenolic, Palmatine, Gilgitine, Jhelumine, Kara-koramine	Hepatitis C	ND	Ikram, 1975; Yousaf et al., 2018
28	Chaaty	<i>Camellia sinensis</i> (L.) Kuntze	Theaceae	Epigallocatechin-3-Gallate (EGCG), Epicatechin Gallate (ECG), Epigallocatechin (EGC)	Infectious Laryngotrachietis Virus (ILTV) Feline calicivirus (FCV) Influenza A (H1N1) HIV-1 Enterovirus 71 (EV71) HSV-1	ND Suppression the level of ROS Inhibition of replication ND Inhibition of replication ND	Zaher et al., 2008 Oh et al., 2013 Ling et al., 2012 Li et al., 2011 Ho et al., 2009 De Oliveira et al., 2013
29	Lahsun	<i>Allium sativum</i> L.	Alliaceae	Alliin, Allicin, E-Ajoene	Human cytomegalovirus (HCMV) Influenza B	Inhibition of replication ND	Zhen et al., 2006
30	Calamus/ Ghorbach	<i>Acorus calamus</i> L.	Araceae	Tatanan, b-asarone, a-asarone	Dengue virus (DENV) SARS CoV2/COVID19	Inhibition of RNA replication Inhibition expression of viral protease (Mpro)	Tsai et al., 1985 Rajput et al., 2014 Krishnasamy et al., 2020
31	Papita	<i>Carica papaya</i> L.	Caricaceae	Chlorogenic acid, 5, 7-dimethoxycoumarin kaempferol quercetin	Dengue virus SARS CoV2/COVID19	Inhibition expression of viral protease (NS2B-NS3 serine protease) Inhibition expression of viral protease (Mpro)	Senthivel et al., 2013; Sharma et al., 2019
32	Aam	<i>Mangifera indica</i> L.	Anacardiaceae	Mangiferin	HSV-2 HIV	Inhibition of replication ND	Yang et al., 2020 Zhu et al., 1993 Guha et al., 1996
33	Mungna/ saijna	<i>Moringa oleifera</i> Lam.	Moringaceae	niaziminic, niazirin, beta-sitosterol, kaempferide	NDV HIV	ND Inhibition of replication	Chollom et al., 2012 Nworu et al., 2013
34	Mokoi	<i>Solanum nigrum</i> L.	Solanaceae	Rutin, Gallic acid, Protocatechuic acid	Hepatitis C HIV-1	Inhibition expression of viral protease (NS3) Inhibition of replication	Javed et al., 2011 Yu, 2004

(Contd...)

Table 1: (Continued)

S. No.	Vernacular Name	Scientific Name	Family	Key compound	Viral disease	Mode of action	References
35	Babul	<i>Acacia nilotica</i> (L.) Delile	Fabaceae		Hepatitis C	Inhibition expression of viral protease (NS3-4A protease)	Rehman et al., 2011
36	Sankuppi	<i>Clerodendrum inerme</i> (L.) Gaertn.	Verbenaceae	cirisimarinin, pectolinarigenin, apigenin	SARS CoV	Inhibition expression of Ribosome	Olivieri et al., 1996
37	Arus	<i>Justicia adhatoda</i> L.	Acanthaceae	alpha-Sitosterol, Amrinone, Pentadecanoic Acid, Stigmasterol	Influenza	Inhibition of viral attachment and/or viral replication	Chavan & Chowdhary, 2014
38	Isabgol	<i>Plantago major</i> L.	Plantaginaceae	caffeic acid, chlorogenic acid, Luteolin, p-coumaric acid	Adenoviruses (ADV)	Inhibition of viral attachment and/or viral replication	Chiang et al., 2002
39	Harad	<i>Terminalia chebula</i> Retz.	Combretaceae	chebumeinin A, chebumeinin B, Ellagic acid, Gallic acid, Chebulic acid	HSV-1 & 2 Hepatitis B Hepatitis C	Enhancing secretion of interferon- γ Inhibition of replication Inhibition expression of viral protease (NS3-4A protease)	Chiang et al., 2003 Chung et al., 1997 Ajala et al., 2014

Table 2: Ethnobotanical uses of plants

S. No.	Scientific Name	Ethnobotanical uses	References
1	<i>Phyllanthus amarus</i> Schumach. & Thonn.	In jaundice paste or fresh roots are given orally. Whole plant is taken as an antimalarial.	Balakrishnan et al., 2009 Upadhyay et al., 2010
2	<i>P. niruri</i> L.	The fresh roots powder for jaundice. Decoction of dried leaves is taken orally for indigestion and used in bath for fever.	Singh, 1986 Weniger et al., 1986
3	<i>Emblia officinalis</i> Gaertn.	Leaf juice are used in dysentery and gonorrhea.	Narayan & Singh, 2017
4	<i>Andrographis paniculata</i> (Burm. f.) Nees	Paste of leaves is applied externally on bitten site of scorpion sting and snakebites.	Samy et al., 2008
5	<i>Curcuma longa</i> L.	Leaf extract is applied for various skin allergies. Paste of leaves used for asthma.	Thomas et al., 2013 Velayudhan et al., 2012
6	<i>C. zanthorrhiza</i> Roxb.	Paste of leaves used in cold and influenza. One glass of milk with two teaspoons turmeric powder used in cold and influenza.	Bouzabata & Boukhari, 2014
7	<i>Boenninghausenia albiflora</i> (Hook.) Rchb. ex Meisn.	Tuberous root sap used for heartburn. Rhizome used for stomachache.	Sujarwo & Caneva, 2015 Rahayu & Andini, 2019
8	<i>Narcissus tazetta</i> L.	Leaves are used to kill the bed bug. Leaves are used for cuts.	Singh, 2008 Singh et al., 2014
9	<i>Listera ovata</i> (L.) R.Br.	Flower and bulb juice taken with milk for skin disease and wounds.	Gürdal & Kültür, 2013
10	<i>Zingiber officinale</i> Rosc.	Whole plant used for blood purification. Tincture of tubers used for externally skin tone.	Khan et al., 2015 Tamm, 1972
11	<i>Tinospora sinensis</i> (Lour.) Merr.	Whole herb is boiled and used for indigestion.	Shapoori et al., 2013
12	<i>Glycrrhiza glabra</i> L.	Rhizome powder paste used for cough and vomiting. Rhizome powder used for muscular pain.	Ahmad et al., 2018 Kumar & Pandey, 2015
13	<i>Withania somnifera</i> L.Dunal	Stem part used in gonorrhea and chlamydia. Extract of its stem along with bark of <i>Alstonia scholaris</i> (L.) R. Br. is taken twice a day for Dysentery.	Acharya & Pokhrel, 2006 Saha et al., 2014
14	<i>Aloe vera</i> (L.) Burm.f.	Root decoction used for Pectoralgia.	Nasab & Khosravi, 2014
15	<i>Azadirachta indica</i> A. Juss.	Whole plant used for bronchitis and ulcer. Leaves are applied on tumour glands.	Dolatkhahi et al., 2014 Jabeen et al., 2010 Rashid & Marwat, 2006
16	<i>Syzygium cumini</i> (L.) Skeels	Leaf pulp is used to promote menstrual flow and mixed leaf pulp in turmeric powder and applied on eyes to relieve eyes pain.	Shende et al., 2014
17	<i>Echinacea purpurea</i> (L.) Moench	Fresh leafy twigs are used for spraying cow's urine inside the hut to keep away insect and mites. Juice made from young leaves mixed with excess water of boil rice used in worm.	Ayyanar et al., 2010 Anisuzzaman et al., 2007
		Leaves paste used for indigestion.	Ayyanar & Ignacimuthu, 2011
		Its bark is used as digestive.	Nadkarni & Nadkarni, 1976
		Leaves and flower powder used for influenza.	Korkmaza et al., 2016
		The roots used to treat blood poisoning, snake poisoning, skin disease.	Kumar & Ramaiah, 2011

(Contd...)

Table 2: (Continued)

S. No.	Scientific Name	Ethnobotanical uses	References
18	<i>Sambucus nigra</i> L.	Flowers are used for inflammation, lowers fever and colds. Leaves and inner bark have also been used for their purgative, emetic, diuretic, topical emollient, expectorant, and diaphoretic action.	Jarić et al., 2007 Kultur, 2007
19	<i>Ocimum sanctum</i> L.	Leaf juice used for glaucoma, cataract, chronic conjunctivitis & other painful eye diseases. Leaves used for skin disease.	Surkar et al., 1994
20	<i>Spinacia oleracea</i> L.	Whole plant used for asthma and leprosy.	Jain et al., 2010 Kumar & Singh, 2019
21	<i>Origanum vulgare</i> L.	Seeds used for fevers, inflammation of the liver and jaundice. Whole plant Used in bronchitis, colic and diarrhea.	Kumar & Singh, 2019 Tiwari et al., 2016
22	<i>Allium cepa</i> L.	Leaves Flowering branches used for Cold and flu.	Polat & Satil, 2012 Ugulu, 2011
23	<i>Capsicum annuum</i> L.	Bulbus used for sprain and edema. Bulbus used as Tonic and for colds, cough.	Singh & Singh, 2009 Ishtiaq et al., 2007
24	<i>Citrus sinensis</i> (L.) Osbeck	Fruits used for snakebite, scarlatina. Fruits are eaten during loss of appetite, indigestion and to 'purify blood'.	Kichu et al., 2015
25	<i>Berberis lycium</i> Royle	Peels and fruits used as mosquito repellent. Leaf decoction used for cold.	Obata & Aigbokhan, 2012 Camejo-Rodrigues et al., 2003
26	<i>Camellia sinensis</i> (L.) Kuntze	Fresh leaves decoction is taken orally for Jaundice. Roots decoction used for diabetes and body pain.	Abbas et al., 2013 Ahmad et al., 2018
27	<i>Allium sativum</i> L.	Root, flower decoction used for hypertension and coughing. Leaves used for Hepatitis.	Au et al., 2008 Amiri & Joharchi, 2013
28	<i>Acorus calamus</i> L.	Bulbils extract used for Bee bite and high blood pressure. Bulbils extract used for treating whooping cough in children.	Bulut & Tuzlaci, 2015 Jarić et al., 2015
29	<i>Carica papaya</i> L.	Roots used for pneumonia and cough. Roots used for snake bite, liver disorder and cold.	Hossan et al., 2009 Balakumbahan et al., 2010
30	<i>Mangifera indica</i> L.	Roots and leaves used for typhoid, stomach upset, malaria and diarrhea. Fruits and decoction of leaves used for constipation.	Saotoing et al., 2011 Abe & Ohtani, 2013
31	<i>Moringa oleifera</i> Lam	Kernels of mango used for diarrhoea. Leaves are inhaled to get relief from throat infections.	Sairam et al., 2003 Narayanan et al., 2011
32	<i>Solanum nigrum</i> L.	Leaves and seeds are used for diabetes, anemia and high blood pressure.	Abe & Ohtani, 2013
33	<i>Acacia nilotica</i> (L.) Delile	Leaves extract used for washing painful eyes. Leaves and fruits are used as liver tonic.	Abbasi et al., 2013 Singh et al., 2014
34	<i>Clerodendrum inerme</i> (L.) Gaertn.	Dysentery, asthma and stomachache. Gum used in cough, bark used for diarrhea.	Rashid et al., 2015 Hussain et al., 2008
35	<i>Justicia adhatoda</i> L.	Leaf paste is used in fever. Bathing with the leaf decoction is claimed to be effective against fever.	Partha, 2014 Bhandary et al., 1995
36	<i>Plantago major</i> L.	Whole plant used for jaundice and hepatitis. Crushed leaves applied on chest for pneumonia treatment.	Abbasi et al., 2009 Vijayan, 2007
37	<i>Terminalia chebula</i> Retz	Leaves and roots are used for wound healing. Leaf used for coughs, stomach ulcers and insect bites.	Azizi & Keshavarzi, 2015 Jarić et al., 2015
		Seeds and fruits for indigestion. Fruits used for intestinal ulcer.	Sahu et al., 2013 Ayyanar & Ignacimuthu, 2011

Similarly, andrographolide from *Andrographis paniculata* (Burm. f.) Nees (green chireta) can be used as a potent inhibitor of the main protease of SARS-CoV-2 (Mpro) (Enmozhi et al., 2020). Bis-andrographolide ether and 6 other compounds isolated from the aerial parts of *A. paniculata* showed anti-HIV and cytotoxic activity as well (Niranjan et al., 2005). Andrographolide inhibits the production of mature viral particles (Pongtuluran et al., 2015). Compounds viz., α - Hederin and Nigellidine isolated from another medicinal plant cum Indian spice, *Nigella sativa* L. (Kalonji), might inhibit the activity of COVID-19 by acting on the main protease (Mpro) (Bouchentouf & Missoum, 2020). Besides, the *N. sativa* mixture is also found to have virucidal activity as it reduces viral load significantly along with the symptoms of HIV infection (Onifade et al., 2013). *N. sativa* modulates the pro

and anti-inflammatory cytokines and acts as a balancing factor between Th1/Th2 lymphocytes (Gholamnezhad et al., 2014).

Glabrin B, shimppterocarpin, glabridin, glycyrrhetic acid, hispaglabridin A, licochalcone A, prenyllicolavone, apioside A, liquiritigenin, isoliquiritin, liquiritinapioside, shinflavanone and isoliquiritigenin like phytochemicals isolated from *Glycyrrhiza glabra* L. (Yashtimadhu) act against SARS-CoV-2 main protease (Mpro). Glycyrrhetic acid, apioside, glabridin, liquiritinapioside, shimppterocarpin and shinflavanone are the key phytochemicals isolated from *G. glabra* that act against human ACE2 receptor and their different protein residues which are involved in the interaction (Maurya, 2020). Glycyrrhizin, an active compound of liquorice roots is found to inhibit the

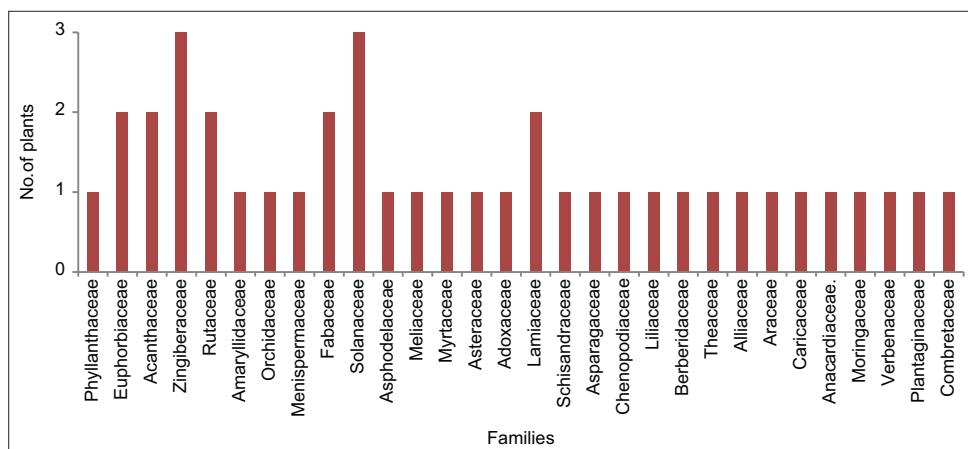


Figure 2: Percent (%) contribution of reported plant families as source of antiviral compounds

replication of the SARS-associated virus (Cinatl *et al.*, 2003). Glycyrrhizin (GL) is also used in infection against the influenza A virus (IAV) (Wolkerstorfer *et al.*, 2009). Likewise, essential oils *viz.*, allyl disulfide and allyl trisulfide account for 51.3% of total essential oils isolated from *Allium sativum* L. (Garlic), have strong interactions with the amino acids of the ACE2 protein and the main protease PDB6LU7 of SARS-CoV-2 (Thuy *et al.*, 2020). All these compounds have huge potential to be used as potent inhibitors against SARS-CoV-2. *A. sativum* and *A. cepa* L. (onion) contain organosulphur compounds like quercetin and allicin which are also associated with the inhibition of viral infection. Quercetin shows effect on entry and attachment of Enterovirus and Influenza virus. This compound also inhibits RNA polymerase which is necessary for viral infection. Organosulphur compounds allicin, diallyl trisulphide and ajoene are the main chemical compounds that are responsible for the antiviral activity of garlic (Sharma, 2019). Chemical compounds present in garlic are also used in the treatment of cancer, diabetes, atherosclerosis and hyperlipidemia (Gebreyohannes & Gebreyohannes, 2013).

Astragalin, lupeol and nictoflorin isolated from *Nyctanthes arbor-tristis* L. (Harsingar) and aloenin and aloesin from *Aloe barbadensis* Miller can also be used as inhibitors of COVID-19 protease (Srivastava *et al.*, 2020). *Aloe* polysaccharides (APS) extracted from *A. vera* leaves inhibit the replication of the H1N1 subtype influenza virus (Sun *et al.*, 2018). Punicafolin from *Phyllanthus emblica* L. (Indian gooseberry), amaroswerin from *Swertia chirata* Buch.-Ham. ex Wall (Chirayta), heptacosanol from *Eclipta prostrata* L. (Bhringraj) have binding affinity for inhibition of spike protein. Amarogentin from *S. chirata*, azadirachtin and rutin from *A. indica*, phyllanemblinin A from *P. emblica* L. also inhibits M-Pro protein in case of COVID-19 (Ranjan, 2020).

One of the major phytoconstituents isolated from *Rheum emodi* Wall. (Indian Rhubarb) *i.e.*, emodin is also reported to have an inhibitory effect on the interaction of SARS-CoV. It has been also found that emodin blocks both the binding of the SARS-CoV S protein to ACE2 and the infectivity of the S-protein (Bilal *et al.*, 2013). Theaflavin, an active compound isolated from *Camellia sinensis* (L.) Kuntze (tea plant) could also be

used against SARS-CoV-2 as it binds with RNA-dependent RNA polymerase of COVID-19 (Lung *et al.*, 2020). Catechin from *C. sinesis* (green tea) and curcumin from *Curcuma longa* (turmeric) also have a dual binding affinity. Both these molecules bind to ACE2 and viral S-protein. Curcumin binds directly to the receptor binding Domain (RBD) of viral S-protein. Catechin binds with ACE2 and S-protein (Jena *et al.*, 2020). Curcumin also inhibits Chikungunya and Zika virus replication. Curcumin mainly works by interfering with virus cell binding (Mounce *et al.*, 2017) and shows inhibition against Influenza virus infection and exhibit haemagglutination activity (Chen *et al.*, 2010).

Dandelion, *Taraxacum officinale* (L.) Weber ex F.H.Wigg, inhibits the influenza virus infection by inhibiting the viral polymerase activity and the reduction of the virus nucleoprotein (NP) RNA (He *et al.*, 2011). Likewise, 6-gingerol extracted from the rhizome of *Zingiber officinale* Roscoe have a binding affinity with various COVID-19 viral proteins and thus could be used as a potential choice to treat COVID-19 (Rathinavel *et al.*, 2020). Fresh ginger by stimulating mucosal cells to secrete Interferon-beta (IFN- β), also contributes to counteracting viral infection (San *et al.*, 2013). Leaves of *Olea europaea* L. (olive) also show inhibitory activity against the influenza virus (Nekooeian, 2006). Olive leaves inhibit the replication of HIV, cell to cell transmission and acute infection (Lee-Huang *et al.*, 2003).

Medicinal Plants used to Enhance Immunity

Besides, fighting the infection, another strategy to overcome the deadly virus is to strengthen the body's immune system. There are several plants in Indian traditional systems of medicine that are used as excellent immunity boosters. Different formulations are made using these plants or these are used in raw form individually or in combinations to augment immunity.

Infact, viral and bacterial infections attack the immunity system of the host. Many of the vitamins and proteins play an important role to fight these infectious agents. Among the vitamins, vitamin C (ascorbic acid) plays an important role in enhancing immunity. Ascorbic acid is essential for the stimulation of immune response by acting as an animmunostimulant, anti-inflammatory, antiviral

and antibacterial nutrient supplement (Sorice *et al.*, 2014). Citrus fruits (lemon and oranges), tomato, papaya, strawberry, green chillies, pepper, mango etc. are some of the vitamin C rich fruits. Similarly, *Tinospora cordifolia* (Willd.) Miers (Guduchi) is a rich source of vitamin C and effective in inhibiting the growth of bacteria and has a tremendous ability to boost immunity (Saha & Ghosh, 2012). *Emblica officinalis* Gaertn. (Amla) also contains a high amount of natural vitamin C and, therefore, is used extensively in raw as well as processed forms as an immunostimulant in India. *Picrorhiza kurroa* Royle ex Benth. (Kutki) also possesses the ability to stimulate a humoral response by acting on various levels of immune mechanisms such as antibody production (Hussain *et al.*, 2013). Pectins from *Centella asiatica* L. (Brahmi) also showed immunostimulating activities (Wang *et al.*, 2003).

Plant Based Immunity Booster Drugs Developed in India

Ministry of AYUSH, India, made a herbal formulation named 'AyushKwath' or 'AyushKudineer' or 'AyushJoshanda'. This mixture consists of *Piper nigrum* L. (Krishna Marich), *Cinnamomum verum* J. Presl bark (Dalchini), *O. sanctum* leaves (Tulsi), dry *Z. officinale* Rosc. (Sunthi) and is used as an immunity booster. Trikatu, as per Ayurvedic's Bhaisajya Ratnawali is a mixture of three bitter herbs, consisting of dried fruits of *Piper nigrum* L. (Maricha), *P. longum* L. (Peepli) and *Z. officinale* Roscoe (Sunthi) (Kaushik & Trikatu, 2018). Triphala composed of *Terminalia chebula* Retz., *T. belerica* Wall and *Phyllanthus emblica* L. is used as an immunomodulator, antibacterial, radioprotective, antioxidant, anti-stress, antimutagenic, chemoprotective, anticancerous and anti-inflammatory herbal medicine (Baliga *et al.*, 2012). Tablet Divya Swasari Ras (DSR) is a unique combination of various plant parts, namely, galls of *Pistacia integerrima* J.L. Stewart ex Brandis (Zebrawood/Karkatashringi), fruits of *Glycyrrhiza glabra* L. (Licorice/Mulethi), the bark of *Cinnamomum zeylanicum* Blume (Cinnamon/Dalchini), fruits of *Cressa cretica* L. (Salt Cresse/Rudanti), rhizomes of *Z. officinale*, fruits of *P. nigrum* L. (Maricha/Black pepper), fruits of *P. longum* L. (Long pepper/Pippali), roots of *Anacyclus pyrethrum* L. and buds of *Syzygium aromaticum* (L.) Merr. & L.M.Perry (Cloves/Lavanga). It is used to protect against allergic airway inflammation and possesses a potential therapeutic option for viral and allergic asthma management (Balkrishna *et al.*, 2020).

Chyawanprash, an ayurvedic mixture of many herbs is a powerful immunity booster formulation that is used in India since time immemorial for fighting general debility and body rejuvenation for delaying the ageing process (Gupta *et al.*, 2017). Chyawanprash consists of several ingredients like *Sida cordifolia* Linn. (bala), *Tribulus terrestris* Linn. (Gokshur), *Aegle marmelos* Correa (Bael), *W. somnifera* (Aswagandha), *P. longum* Linn. (Pippali), *Emblica officinalis* (Amla), *Asparagus racemosus* (Shatavari), *C. asiatica* (Brahmi) and *Eugenia caryophyllus* Linn (Sharma *et al.*, 2019).

From ancient times, plants are used for therapeutic purposes in many civilizations across the globe. Even today, plants serve as

the foundation for many pharmaceuticals. As a relief to a variety of illnesses, the use of medicinal plants is a traditional practice among different ethnic groups. However, this information is rarely documented and often passed down orally from generation to generation. When a traditional healer dies without passing on his experience to the next generation, the world loses hundreds of years of invaluable therapeutic knowledge. This ethnobotanical knowledge is highly significant and must be prioritised for preservation. Undoubtedly, the ethnobotanical know how of traditional systems of medicines helped a lot in boosting the immunity of people in the prevailing COVID era. However, the scientific base of this must be clinically evaluated for therapeutic effectiveness. Table 2 lists the ethnobotanical applications of plants used as antiviral agents in various scientific studies.

CONCLUSION

Traditional Indian medicinal practices relied solely on medicinal plants and even today, the potential of phytochemicals obtained from medicinal plants in curing diseases cannot be denied. Research is advancing, with the worldwide increase in the number of people affected by coronavirus, in the field of isolating and analyzing the phytochemicals that possess antiviral activity and to elucidate their mode of action. A considerable number of medicinal plants and their phytochemicals have been investigated for antiviral activity. Also, the immune-boosting effect of bioactive compounds obtained from these medicinal plants provides a holistic approach to cure various infections. With the ongoing evolution of various viruses resulting in increased resistance against the current drug therapy, the phytochemicals obtained from plants can be a significant potential source of therapy to treat various viral infections including the infection caused by changing strains of coronavirus. Many promising herbal treatments exist for viral diseases with proof of their efficacy and safety in advanced clinical trials. Through this article, we try to summarize and consolidate most of the commonly available potential Indian medicinal plant sources that can be used against viral infections especially the challenging strains of COVID-19. However, a lot of work still remains to be done to determine optimal treatments, doses, and formulae for those herbal preparations. Researches need to proceed in the direction of drug formation from phytochemicals against COVID-19. Besides, the pharmacodynamic and pharmacokinetic mechanisms of action of bioactive compounds derived from these medicinal plants on the structure and function of novel coronavirus needs to be studied as well.

REFERENCES

- Abbasi, A. M., Khan, M. A., Ahmad, M., Zafar, M., Khan, H., Muhammad, N., & Sultana, S. (2009). Medicinal plants used for the treatment of jaundice and hepatitis based on socio-economic documentation. *African Journal of Biotechnology*, 8(8), 1643-1650.
- Abbasi, A. M., Khan, M. A., Shah, M. H., Shah, M. M., Pervez, A., & Ahmad, M. (2013). Ethnobotanical appraisal and cultural values of medicinally important wild edible vegetables of Lesser Himalayas-Pakistan. *Journal of Ethnobiology and Ethnomedicine*, 9(66), 1-13. DOI <https://doi.org/10.1186/1746-4269-9-66>
- Abe, R., & Ohtani, K. (2013). An ethnobotanical study of medicinal plants and traditional therapies on Batan Island, the Philippines. *Journal*

- of *Ethnopharmacology*, 145(2), 554-565. <https://doi.org/10.1016/j.jep.2012.11.029>
- Acharya, E., & Pokhrel, B. (2006). Ethno-medicinal plants used by Bantar of Baudaha, Morang, Nepal. *Our nature*, 4(1), 96-103. <https://doi.org/10.3126/on.v4i1.508>.
- Ahmad, A., Ahad, A., Rao, A. Q., & Husnain, T. (2015). Molecular docking based screening of neem-derived compounds with the NS1 protein of Influenza virus. *Bioinformation*, 11(7), 359-365. <https://doi.org/10.6026/97320630011359>.
- Ahmad, M., Zafar, M., Shahzadi, N., Yaseen, G., Murphrey, T. M., & Sultana, S. (2018). Ethnobotanical importance of medicinal plants traded in Herbal markets of Rawalpindi-Pakistan. *Journal of Herbal Medicine*, 11, 78-89. <https://doi.org/10.1016/j.hermed.2017.10.001>
- Ahmad, M., Zafar, M., Shahzadi, N., Yaseen, G., Murphrey, T. M., & Sultana, S. (2018). Ethnobotanical importance of medicinal plants traded in Herbal markets of Rawalpindi-Pakistan. *Journal of Herbal Medicine*, 11, 78-89. <https://doi.org/10.1016/j.hermed.2017.10.001>
- Ahvazi, M., Mozaffarian, V., Nejadsatari, T., Mojtaba, F., Charkhchiyan, M. M., Khalighi-Sigaroodi, F., & Ajani, Y. (2007). Medicinal application of native plants (Lamiaceae and Rosaceae family) in Alamut region in Gazvin province. *Journal of Medicinal Plants*, 6(24), 74-84.
- Ajala, O. S., Jukov, A., & Ma, C. M. (2014). Hepatitis C virus inhibitory hydrolysable tannins from the fruits of *Terminalia chebula*. *Fitoterapia*, 99, 117-123. <https://doi.org/10.1016/j.fitote.2014.09.014>
- Alhajj, M. S., Qasem, M. A., & Al-Mufarrej, S. I. (2020). Inhibitory activity of *Ilicium verum* extracts against avian viruses. *Advances in Virology*, 2020, 4594635. <https://doi.org/10.1155/2020/4594635>
- Amiri, M. S., & Joharchi, M. R. (2013). Ethnobotanical investigation of traditional medicinal plants commercialized in the markets of Mashhad, Iran. *Avicenna Journal of Phytomedicine*, 3(3), 254-271.
- Anand, K., Ziebuhr, J., Wadhwani, P., Mesters, J. R., & Hilgenfeld, R. (2003). Coronavirus main proteinase (3CLpro) structure: basis for design of anti-SARS drugs. *Science*, 300(5626), 1763-1767. <https://doi.org/10.1126/science.1085658>
- Andersen, K. G., Rambaut, A., Lipkin, W. I., Holmes, E. C., & Garry, R. F. (2020). The proximal origin of SARS-CoV-2. *Nature Medicine*, 26(4), 450-452. <https://doi.org/10.1038/s41591-020-0820-9>
- Anisuzzaman, M., Rahman, A. H. M. M., Harun-Or-Rashid, M., Naderuzzaman, A. T. M., & Islam, A. K. M. R. (2007). An ethnobotanical study of Madhupur, Tangail. *Journal of Applied Sciences Research*, 3(7), 519-530.
- Ashfaq, U. A., Jalil, A., & ul Qamar, M. T. (2016). Antiviral phytochemicals identification from *Azadirachtaindica* leaves against HCV NS3 protease: an *in silico* approach. *Natural Product Research*, 30(16), 1866-1869. <https://doi.org/10.1080/14786419.2015.1075527>
- Asres, K., Bucar, F., Kartnig, T., Witvrouw, M., Pannecouque, C., & De Clercq, E. (2001). Antiviral activity against human immunodeficiency virus type 1 (HIV-1) and type 2 (HIV-2) of ethnobotanically selected Ethiopian medicinal plants. *Phytotherapy Research*, 15(1), 62-69. [https://doi.org/10.1002/1099-1573\(200102\)15:1<62::AID-PTR956>3.0.CO;2-X](https://doi.org/10.1002/1099-1573(200102)15:1<62::AID-PTR956>3.0.CO;2-X)
- Astani, A., Reichling, J., & Schnitzler, P. (2011). Screening for antiviral activities of isolated compounds from essential oils. *Evidence-based Complementary and Alternative Medicine*, 2011, 253643. <https://doi.org/10.1093/ecam/nep187>
- Au, D. T., Wu, J., Jiang, Z., Chen, H., Lu, G., & Zhao, Z. (2008). Ethnobotanical study of medicinal plants used by Hakka in Guangdong, China. *Journal of Ethnopharmacology*, 117(1), 41-50. <https://doi.org/10.1016/j.jep.2008.01.016>
- Ayyanar, M., & Ignacimuthu, S. (2011). Ethnobotanical survey of medicinal plants commonly used by Kanitribals in Tirunelveli hills of Western Ghats, India. *Journal of Ethnopharmacology*, 134(3), 851-864. <https://doi.org/10.1016/j.jep.2011.01.029>
- Ayyanar, M., Sankarasivaraman, K., Ignacimuthu, S., & Sekar, T. (2010). Plant species with ethnobotanical importance other than medicinal in Theni district of Tamil Nadu, Southern India. *Asian Journal of Experimental Biological Sciences*, 1(4), 765-771.
- Azizi, H., & Keshavarzi, M. (2015). Ethnobotanical study of medicinal plants of Sardasht, Western Azerbaijan, Iran. *Journal of Herbal Drugs*, 6(2), 113-119.
- Badam, L. (1997). *In vitro* antiviral activity of indigenous glycyrrhizin, licorice and glycyrrhetic acid (Sigma) on Japanese encephalitis virus. *The Journal of Communicable Diseases*, 29(2), 91-99.
- Badam, L., Joshi, S. P., & Bedekar, S. S. (1999). *In vitro*antiviral activity of neem (*Azadirachtaindica* A. Juss) leaf extract against group B coxsackieviruses. *The Journal of Communicable Diseases*, 31(2), 79-90.
- Balakrishnan, V., Prema, P., Ravindran, K. C., & Robinson, J. P. (2009). Ethnobotanical studies among villagers from Dharapuram taluk, Tamil Nadu, India. *Global Journal of Pharmacology*, 3(1), 08-14.
- Balakumbahan, R., Rajamani, K., & Kumanan, K. (2010) *Acorus calamus*: an overview. *Journal of Medicinal Plants Research*, 4(25), 2740-2745.
- Baliga, M. S., Meera, S., Mathai, B., Rai, M. P., Pawar, V., & Palatty, P. L. (2012). Scientific validation of the ethnomedicinal properties of the Ayurvedic drug Triphala: a review. *Chinese Journal of Integrative Medicine*, 18(12), 946-954. <https://doi.org/10.1007/s11655-012-1299-x>
- Balkrishna, A., Solleti, S. K., Singh, H., Tomer, M., Sharma, N., & Varshney, A. (2020). Calcio-herbal formulation, Divya-Swasari-Ras, alleviates chronic inflammation and suppresses airway remodelling in mouse model of allergic asthma by modulating pro-inflammatory cytokine response. *Biomedicine & Pharmacotherapy*, 12(110063), 1-15. <https://doi.org/10.1016/j.bioph.2020.110063>
- Bhandary, M. J., Chandrashekhar, K. R., & Kaveriappa, K. M. (1995). Medical ethnobotany of the siddis of Uttara Kannada district, Karnataka, India. *Journal of Ethnopharmacology*, 47(3), 149-158. [https://doi.org/10.1016/0378-8741\(95\)01274-H](https://doi.org/10.1016/0378-8741(95)01274-H)
- Bilal, S., Mir, M. R., Parrah, J. D., Tiwari, B. K., Tripathi, V., Singh, P., & Abidi, A. B. (2013). Rhubarb: the wondrous drug. A review. *International Journal of Pharmacy and Biological Sciences*, 3(3), 228-233.
- Borse, S., Joshi, M., Saggam, A., Bhat, V., Walia, S., Marathe, A., & Tillu, G. (2021). Ayurveda botanicals in COVID-19 management: an *in silico* multi-target approach. *Plos One*, 16(6), 1-29. <https://doi.org/10.1371/journal.pone.025361/v1>
- Bouchentouf, S., & Missoum, N. (2020). Identification of Compounds from *Nigella Sativa* as New Potential Inhibitors of 2019 Novel Coronavirus (Covid-19): Molecular Docking Study. *Chem Rxiv*, 2(2), 1-8. <https://doi.org/10.26434/chemrxiv.12055716>
- Bouzabata, A., & Boukhari, A. (2014). Variation in the Traditional Knowledge of *Curcuma longa* L. in North-Eastern Algeria. *International Journal of Pharmacological and Pharmaceutical Sciences*, 8(11), 1227-1231.
- Bulut, G., & Tuzlaci, E. (2015). An ethnobotanical study of medicinal plants in Bayramiç (Çanakkale-Turkey). *Marmara Pharmaceutical Journal*, 19(3), 268-282. <https://doi.org/10.12991/mpj.201519392830>
- Cai, Z., Zhang, G., Tang, B., Liu, Y., Fu, X., & Zhang, X. (2015). Promising anti-influenza properties of active constituent of *Withania somnifera* ayurvedic herb in targeting neuraminidase of H1N1 influenza: computational study. *Cell Biochemistry and Biophysics*, 72(3), 727-739. <https://doi.org/10.1007/s12013-015-0524-9>
- Camejo-Rodrigues, J., Ascensao, L., Bonet, M. Á., & Valles, J. (2003). An ethnobotanical study of medicinal and aromatic plants in the Natural Park of "Serra de São Mamede"(Portugal). *Journal of Ethnopharmacology*, 89(2-3), 199-209. [https://doi.org/10.1016/S0378-8741\(03\)00270-8](https://doi.org/10.1016/S0378-8741(03)00270-8)
- Chantrill, B. H., Coulthard, C. E., Dickinson, L., Inkley, G. W., Morris, W., & Pyle, A. H. (1952). The action of plant extracts on a bacteriophage of *Pseudomonas pyocyannea* and on influenza A virus. *Microbiology*, 6(1-2), 74-84. <https://doi.org/10.1099/00221287-6-1-2-74>
- Chavan, R., & Chowdhary, A. (2014). *In vitro* inhibitory activity of *Justicia adhatoda* extracts against influenza virus infection and hemagglutination. *International Journal of Pharmaceutical Sciences Review and Research*, 25(2), 231-236.
- Chen, C. H., Chou, T. W., Cheng, L. H., & Ho, C. W. (2011). *In vitro* antiadenoviral activity of five *Allium* plants. *Journal of the Taiwan Institute of Chemical Engineers*, 42(2), 228-232. <https://doi.org/10.1016/j.jtice.2010.07.011>
- Chen, D. Y., Shien, J. H., Tiley, L., Chiou, S. S., Wang, S. Y., Chang, T. J., & Hsu, W. L. (2010). Curcumin inhibits influenza virus infection and haemagglutination activity. *Food Chemistry*, 119(4), 1346-1351. <https://doi.org/10.1016/j.foodchem.2009.09.011>
- Chen, J. X., Xue, H. J., Ye, W. C., Fang, B. H., Liu, Y. H., Yuan, S. H., & Wang, Y. Q. (2009). Activity of andrographolide and its derivatives against influenza virus *in vivo* and *in vitro*. *Biological and Pharmaceutical Bulletin*, 32(8), 1385-1391. <https://doi.org/10.1248/bpb.32.1385>
- Chen, N., Zhou, M., Dong, X., Qu, J., Gong, F., Han, Y., Qiu, Y., Wang, J., Liu, Y., Wei, Y., Xia, J., Yu, T., Zhang, X., & Zhang, L. (2020). Epidemiological and clinical characteristics of 99 cases of 2019 novel coronavirus pneumonia in Wuhan, China: a descriptive study. *Lancet*, 395(10223),

- 507-513. [https://doi.org/10.1016/S0140-6736\(20\)30211-7](https://doi.org/10.1016/S0140-6736(20)30211-7)
- Chiang, L. C., Chiang, W., Chang, M. Y., & Lin, C. C. (2003). *In vitro* cytotoxic, antiviral and immunomodulatory effects of *Plantago major* and *Plantagoasiatica*. *The American Journal of Chinese Medicine*, 31(02), 225-234. <https://doi.org/10.1142/S0192415X03000874>
- Chiang, L. C., Chiang, W., Chang, M. Y., Ng, L. T., & Lin, C. C. (2002). Antiviral activity of *Plantago major* extracts and related compounds *in vitro*. *Antiviral Research*, 55(1), 53-62. [https://doi.org/10.1016/S0166-3542\(02\)00007-4](https://doi.org/10.1016/S0166-3542(02)00007-4)
- Chiang, L. C., Ng, L. T., Cheng, P. W., Chiang, W., & Lin, C. C. (2005). Antiviral activities of extracts and selected pure constituents of *Ocimumbasilicum*. *Clinical and Experimental Pharmacology and Physiology*, 32(10), 811-816. <https://doi.org/10.1111/j.1440-1681.2005.04270.x>
- Chollom, S. C., Agada, G. O. A., Gotep, J. G., Mwankon, S. E., Dus, P. C., Bot, Y. S., & Okwor, A. E. J. (2012). Investigation of aqueous extract of *Moringa oleifera* lam seed for antiviral activity against newcastle disease virus *in ovo*. *Journal of Medicinal Plants Research*, 6(22), 3870-3875. <https://doi.org/10.5897/JMPR12.394>
- Chung, T. H., Kim, J. C., Lee, C. Y., Moon, M. K., Chae, S. C., Lee, I. S., & Lee, I. P. (1997). Potential antiviral effects of *Terminalia chebula*, *Sanguisorba officinalis*, *Rubuscoreanus* and *Rheum palmatum* against duck hepatitis B virus (DHBV). *Phytotherapy Research*, 11(3), 179-182. [https://doi.org/10.1002/\(SICI\)1099-1573\(199705\)11:3<179::AID-MTR11>3.0.CO;2-1](https://doi.org/10.1002/(SICI)1099-1573(199705)11:3<179::AID-MTR11>3.0.CO;2-1)
- Cinatl, J., Morgenstern, B., Bauer, G., Chandra, P., Rabenau, H., & Doerr, H. W. (2003). Glycyrrhizin, an active component of liquorice roots, and replication of SARS-associated coronavirus. *The Lancet*, 361(9374), 2045-2046. [https://doi.org/10.1016/S0140-6736\(03\)13615-x](https://doi.org/10.1016/S0140-6736(03)13615-x)
- Davidson-Hunt, I. (2000). Ecological ethnobotany: stumbling toward new practices and paradigms. *MASA Journal*, 16(1), 1-13.
- De Clercq, E. (2000). Current lead natural products for the chemotherapy of human immunodeficiency virus (HIV) infection. *Medicinal Research Reviews*, 20(5), 323-349. [https://doi.org/10.1002/1098-1128\(200009\)20:5<323::AID-MED1>3.0.CO;2-A](https://doi.org/10.1002/1098-1128(200009)20:5<323::AID-MED1>3.0.CO;2-A)
- De Clercq, E., & Li, G. (2016). Approved antiviral drugs over the past 50 years. *Clinical Microbiology Reviews*, 29(3), 695-747. <https://doi.org/10.1128/CMR.00102-15>
- De Oliveira, A., Adams, S. D., Lee, L. H., Murray, S. R., Hsu, S. D., Hammond, J. R., & Chu, T. C. (2013). Inhibition of herpes simplex virus type 1 with the modified green tea polyphenol palmitoyl-epigallocatechin gallate. *Food and Chemical Toxicology*, 52, 207-215. <https://doi.org/10.1016/j.fct.2012.11.006>
- Devaraj, S., Ismail, S., Ramanathan, S., Marimuthu, S., & Fei, Y. M. (2010). Evaluation of the hepatoprotective activity of standardized ethanolic extract of *Curcuma xanthorrhiza* Roxb. *Journal of Medicinal Plants Research*, 4(23), 2512-2517. <https://doi.org/10.5897/JMPR10.453>
- Divya, C. S., & Pillai, M. R. (2006). Antitumor action of curcumin in human papilloma virus associated cells involves downregulation of viral oncogenes, prevention of NFkB and AP-1 translocation, and modulation of apoptosis. *Molecular Carcinogenesis: Published in cooperation with the University of Texas MD Anderson Cancer Center*, 45(5), 320-332. <https://doi.org/10.1002/mc.20170>
- Dolatkhahi, M., Dolatkhahi, A., & Nejad, J. B. (2014). Ethnobotanical study of medicinal plants used in Arjan-Parishan protected area in Fars Province of Iran. *Avicenna Journal of Phytomedicine*, 4(6), 402-412.
- Dutta, K., Ghosh, D., & Basu, A. (2009). Curcumin protects neuronal cells from Japanese encephalitis virus-mediated cell death and also inhibits infective viral particle formation by dysregulation of ubiquitin-proteasome system. *Journal of Neuroimmuno Pharmacology*, 4(3), 328-337. <https://doi.org/10.1007/s11481-009-9158-2>
- Dwivedi, V. D., Bharadwaj, S., Afroz, S., Khan, N., Ansari, M. A., Yadava, U., & Kang, S. G. (2021). Anti-dengue infectivity evaluation of bioflavonoid from *Azadirachtaindica* by dengue virus serine protease inhibition. *Journal of Biomolecular Structure and Dynamics*, 39(4), 1417-1430. <https://doi.org/10.1080/07391102.2020.1734485>
- Eggink, P. M., Maliepaard, C., Tikunov, Y., Haanstra, J. P. W., Bovy, A. G., & Visser, R. G. F. (2012). A taste of sweet pepper: Volatile and non-volatile chemical composition of fresh sweet pepper (*Capsicum annuum*) in relation to sensory evaluation of taste. *Food Chemistry*, 132(1), 301-310. <https://doi.org/10.1016/j.foodchem.2011.10.081>
- Enmozhí, S. K., Raja, K., Sebastine, I., & Joseph, J. (2020). Andrographolide as a Potential Inhibitor of SARS-CoV-2 Main Protease: an *in silico* approach. *Journal of Biomolecular Structure and Dynamics*, 39(9), 3092-3098. <https://doi.org/10.1080/07391102.2020.1760136>
- Faccin-Galhardi, L. C., Yamamoto, K. A., Ray, S., Ray, B., Linhares, R. E. C., & Nozawa, C. (2012). The *in vitro* antiviral property of *Azadirachtaindica* polysaccharides for poliovirus. *Journal of Ethnopharmacology*, 142(1), 86-90. <https://doi.org/10.1016/j.jep.2012.04.018>
- Farnsworth, N. R., & Soejarto, D. D. (1991). Global importance of medicinal plants. *The Conservation of Medicinal Plants*, 26, 25-51.
- Fernandes, M. J. B., Barros, A. V., Melo, M. S., & Simoni, I. C. (2012). Screening of Brazilian plants for antiviral activity against animal herpes viruses. *Journal of Medicinal Plants Research*, 6(12), 2261-2265. <https://doi.org/10.5897/JMPR10.040>
- Gebreyohannes, G., & Gebreyohannes, M. (2013). Medicinal values of garlic: A review. *International Journal of Medicine and Medical Sciences*, 5(9), 401-408. <https://doi.org/10.5897/IJMMS2013.0960>
- Gholamnezhad, Z., Boskabady, M. H., & Hosseini, M. (2014). Effect of *Nigella sativa* on immune response in treadmill exercised rat. *BMC Complementary and Alternative Medicine*, 14(1), 1-11.
- Grover, A., Agrawal, V., Shandilya, A., Bisaria, V. S., & Sundar, D. (2011). Non-nucleosidic inhibition of Herpes simplex virus DNA polymerase: mechanistic insights into the anti-herpetic mode of action of herbal drug withaferin A. In *BMC Bioinformatics*, 12(13), 1-9. <https://doi.org/10.1186/1471-2105-12-S13-S22>
- Guan, W.-J., Ni, Z.-Y., Hu, Y., Liang, W.-H., Ou, C.-Q., He, J.-X., Liu, L., Shan, H., Lei, C.-L., Hui, D. S. C., Du, B., Li, L.-J., Zeng, G., Yuen, K. Y., Chen, R.-C., Tang, C.-L., Wang, T., Chen, P.-Y., Xiang, J.,...,Zhong, N. S. (2020). Clinical characteristics of coronavirus disease 2019 in China. *The New England Journal of Medicine*, 382(18), 1708-1720. <https://doi.org/10.1056/NEJMoa2002032>
- Guha, S., Ghosal, S., & Chattopadhyay, U. (1996). Antitumor, immunomodulatory and anti-HIV effect of mangiferin, a naturally occurring glucosylxanthone. *Cancer Chemotherapy*, 42(6), 443-451. <https://doi.org/10.1159/000239478>
- Guo, Y. R., Cao, Q. D., Hong, Z. S., Tan, Y. Y., Chen, S. D., Jin, H. J., & Yan, Y. (2020). The origin, transmission and clinical therapies on coronavirus disease 2019 (COVID-19) outbreak—an update on the status. *Military Medical Research*, 7(1), 1-10. <https://doi.org/10.1186/s40779-020-00240-0>
- Gupta, A., Kumar, S., Dole, S., Deshpande, S., Deshpande, V., Singh, S., & Sasibhushan, V. (2017). Evaluation of Cyavanaprásā on health and immunity related parameters in healthy children: a two arm, randomized, open labeled, prospective, multicenter, clinical study. *Ancient Science of Life*, 36(3), 141-150. <https://doi.org/10.4103/asl.ASL817>
- Gupta, G. D., Sujatha, N., Dhanik, A., & Rai, N. P. (2010). Clinical evaluation of Shilajatu Rasayana in patients with HIV infection. *An International Quarterly Journal of Research in Ayurveda*, 31(1), 28-32. <https://doi.org/10.4103/0974-8520.68205>
- Gürdal, B., & Kültür, S. (2013). An ethnobotanical study of medicinal plants in Marmaris (Muğla, Turkey). *Journal of Ethnopharmacology*, 146(1), 113-126. <https://doi.org/10.1016/j.jep.2012.12.012>
- Hamming, I., Timens, W., Bulthuis, M. L. C., Lely, A. T., Navis, G. J., & van Goor, H. (2004). Tissue distribution of ACE2 protein, the functional receptor for SARS coronavirus. A first step in understanding SARS pathogenesis. *The Journal of Pathology and Bacteriology*, 203, 631-637. <https://doi.org/10.1002/path.1570>
- Hanif, S. M., Meher, M. M., & Anower, M. (2016). Field study on efficacy of red pepper (*Capsicum annum*) along with antibiotics against newcastle disease in broiler at Narail Sadar Upazilla, Bangladesh. *Wayamba Journal of Animal Science*, 8, 1460-1466.
- He, W., Han, H., Wang, W., & Gao, B. (2011). Anti-influenza virus effect of aqueous extracts from dandelion. *Virology Journal*, 8(1), 1-11. <https://doi.org/10.1186/1743-422X-8-538>
- Ho, H. Y., Cheng, M. L., Weng, S. F., Leu, Y. L., & Chiu, D. T. Y. (2009). Antiviral effect of epigallocatechin gallate on enterovirus 71. *Journal of Agricultural and Food Chemistry*, 57(14), 6140-6147. <https://doi.org/10.1021/jf901128u>
- Hossain, M. S., Hanif, A., Khan, M., Bari, S., Jahan, R., & Rahmatullah, M. (2009). Ethnobotanical survey of the Tripura tribe of Bangladesh. *American Eurasian Journal of Sustainable Agriculture*, 3(2), 253-261. <https://doi.org/10.1111/j.1440-1681.2005.04270.x>
- Hu, B., Ge, X., Wang, L. F., & Shi, Z. (2015). Bat origin of human coronaviruses. *Virology Journal*, 12(1), 1-10. <https://doi.org/10.1186/s12985-015-0422-1>
- Hussain, A., Shadma, W., Maksood, A., & Ansari, S. H. (2013). Protective effects of *Picrorhizakurroa* on cyclophosphamide-induced

- immunosuppression in mice. *Pharmacognosy Research*, 5(1), 30-35. <https://doi.org/10.4103/0974-8490.105646>
- Hussain, K., Shahzad, A., & Zia-ul-Hussnain, S. (2008). An ethnobotanical survey of important wild medicinal plants of Hattar district Haripur, Pakistan. *Ethnobotanical Leaflets*, 2008(1), 1-9. <https://opensiuc.lib.siu.edu/ebli/vol2008/iss1/5>
- Ikram, M. (1975). A review on the chemical and pharmacological aspects of genus *Berberis*. *Planta Medica*, 28(8), 353-358. <https://doi.org/10.1055/s-0028-1097869>
- Ishtiaq, M., Hanif, W., Khan, M. A., Ashraf, M., & Butt, A. M. (2007). An ethnomedicinal survey and documentation of important medicinal folklore food phytonims of flora of Samahni valley, (Azad Kashmir) Pakistan. *Pakistan Journal of Biological Sciences*, 10(13), 2241-2256. <https://doi.org/10.3923/pjbs.2007.2241.2256>
- Jabeen, S., Shah, M. T., Khan, S., & Hayat, M. Q. (2010). Determination of major and trace elements in ten important folk therapeutic plants of Haripur basin, Pakistan. *Journal of Medicinal Plants Research*, 4(7), 559-566. <https://doi.org/10.5897/JMPR10.015>
- Jadhav, P., Kapoor, N., Thomas, B., Lal, H., & Kshirsagar, N. (2012). Antiviral potential of selected Indian medicinal (ayurvedic) plants against herpes simplex virus 1 and 2. *North American Journal of Medical Sciences*, 4(12), 641-647. <https://doi.org/10.4103/1947-2714.104316>
- Jain, D. L., Baheti, A. M., Jain, S. R., & Khandelwal, K. R. (2010). Use of medicinal plants among tribes in Satpuda region of Dhule and Jalgaon districts of Maharashtra - an ethnobotanical survey. *Indian Journal of Traditional Knowledge*, 9(1), 152-157.
- Jarić, S., Mačukanović-Jocić, M., Djurdjević, L., Mitrović, M., Kostić, O., Karadžić, B., & Pavlović, P. (2015). An ethnobotanical survey of traditionally used plants on Suva planina mountain (south-eastern Serbia). *Journal of Ethnopharmacology*, 175, 93-108. <https://doi.org/10.1016/j.jep.2015.09.002>
- Jarić, S., Popović, Z., Mačukanović-Jocić, M., Djurdjević, L., Mijatović, M., Karadžić, B., & Pavlović, P. (2007). An ethnobotanical study on the usage of wild medicinal herbs from Kopaonik Mountain (Central Serbia). *Journal of Ethnopharmacology*, 111(1), 160-175. <https://doi.org/10.1016/j.jep.2006.11.007>
- Javed, T., Ashfaq, U. A., Riaz, S., Rehman, S., & Riazuddin, S. (2011). In-vitro antiviral activity of *Solanum nigrum* against Hepatitis C virus. *Virology Journal*, 8(1), 1-7. <http://www.virologyj.com/content/8/1/26>
- Jayati, B. A., Kumar, A., Goel, A., Gupta, S., & Rahal, A. (2013). In vitro antiviral potential of *Ocimum sanctum* leaves extract against New Castle Disease Virus of poultry. *International Journal of Microbiology and Immunology Research*, 2(7), 51-55.
- Jena, A. B., Kanungo, N., Nayak, V., Chainy, G. B. N., & Dandapat, J. (2021). Catechin and curcumin interact with S protein of SARS-CoV2 and ACE2 of human cell membrane: insights from computational studies. *Scientific Reports*, 11, 2043. <https://doi.org/10.1038/s41598-021-81462-7>
- Joshi, G., Sharma, S., Acharya, J., & Parida, M. (2014). Assessment of In vitro antiviral activity of *Ocimum sanctum* (Tulsi) against pandemic swine flu H1N1 virus infection. *World Research Journal of Antimicrobial Agents*, 3(1), 62-67.
- Kambizi, L. G. B. M., Goosen, B. M., Taylor, M. B., & Afolayan, A. J. (2007). Anti-viral effects of aqueous extracts of *Aloe ferox* and *Withania somnifera* on herpes simplex virus type 1 in cell culture: research in action. *South African Journal of Science*, 103(9), 359-360.
- Kaushik, R. (2018). Trikatu-A combination of three bioavailability enhancers. *International Journal of Green Pharmacy*, 12(03), 437-441. <https://doi.org/10.22377/ijgp.v12i03.2002>
- Kaushik, S., Jangra, G., Kundu, V., Yadav, J. P., & Kaushik, S. (2020). Anti-viral activity of *Zingiber officinale* (Ginger) ingredients against the Chikungunya virus. *Virus Disease*, 31(3), 270-276. <https://doi.org/10.1007/s13337-020-00584-0>
- Keyerta, E., Vijgen, L., Pannecouque, C., Van Damme, E., Peumans, W., Egberink, H., Balzarini, J., & Van Ranst, M. (2007). Plant lectins are potent inhibitors of coronaviruses by interfering with two targets in the viral replication cycle. *Antiviral Research*, 75(3), 179-187. <https://doi.org/10.1016/j.antiviral.2007.03.003>
- Khan, A. A., Ali, F., Ihsan, M., Hayat, K., & Nabi, G. (2015). Ethnobotanical study of the medicinal plants of Tehsil Charbagh, district Swat, Khyber Pakhtunkhwa, Pakistan. *American Eurasian Journal of Agriculture and Environmental Sciences*, 15, 1464-1474. <https://doi.org/10.5829/idosi.aejas.2015.15.7.94235>
- Kharisma, V. D., Septiadi, L., & Syafrudin, S. (2018). Prediction of novel bioactive compound from *Zingiber officinale* as non-nucleoside reverse transcriptase inhibitors (NNRTIs) of HIV-1 through computational study. *Bioinformatics and Biomedical Research Journal*, 1(2), 49-55. <https://doi.org/10.11594/bbrj.01.02.05>
- Kichu, M., Malewska, T., Akter, K., Imchen, I., Harrington, D., Kohen, J., & Jamie, J. F. (2015). An ethnobotanical study of medicinal plants of Chungtia village, Nagaland, India. *Journal of Ethnopharmacology*, 166, 5-17. <https://doi.org/10.1016/j.jep.2015.02.053>
- Kim, H. J., Yoo, H. S., Kim, J. C., Park, C. S., Choi, M. S., Kim, M., & Ahn, J. K. (2009). Antiviral effect of *Curcuma longa* Linn extract against hepatitis B virus replication. *Journal of Ethnopharmacology*, 124(2), 189-196. <https://doi.org/10.1016/j.jep.2009.04.046>
- Kim, H. Y., Shin, H. S., Park, H., Kim, Y. C., Yun, Y. G., Park, S., & Kim, K. (2008). *In vitro* inhibition of coronavirus replications by the traditionally used medicinal herbal extracts, *Cimicifuga rhizoma*, *Meliaecortex*, *Coptidisrhizoma*, and *Phellodendron cortex*. *Journal of Clinical Virology*, 41(2), 122-128. <https://doi.org/10.1016/j.jcv.2007.10.011>
- Kim, J. M., Chung, Y. S., Jo, H. J., Lee, N. J., Kim, M. S., Woo, S. H., & Han, M. G. (2020). Identification of coronavirus isolated from a patient in Korea with COVID-19. *Osong Public Health and Research Perspectives*, 11(1), 3-7. <https://doi.org/10.24171/j.phrp.2020.11.1.02>
- Korkmaz, M., Karakuş, S., Özgelik, H., & Selvi, S. (2016). An ethnobotanical study on medicinal plants in Erzincan, Turkey. *Indian Journal of Traditional Knowledge*, 15(2), 192-202.
- Krishnasamy, R., Baba, M., Bharath, M. V., Phuntsho, J., Arunachalam, D., Natarajan, K., & Ramasamy, M. (2020). *In silico* analysis of active compounds from siddha herbal infusion of AmmaiyoKoondhalKudineer (Akk) against SARS-CoV-2 spike protein and its ACE2 receptor complex. *Preprint*. <http://doi.org/10.2139/ssrn.3578294>
- Kultur, S. (2007). Medicinal plants used in Kırklareli province (Turkey). *Journal of Ethnopharmacology*, 111(2), 341-364. <https://doi.org/10.1016/j.jep.2006.11.035>
- Kumar, A. H. (2020). Molecular docking of natural compounds from tulsi (*Ocimum sanctum*) and neem (*Azadirachtaindica*) against SARS-CoV-2 protein targets. *Biology, Engineering, Medicine and Science Reports*, 6(1), 1-9. <https://doi.org/10.21203/rs.3.rs-27151/v1>
- Kumar, K. M., & Ramaiah, S. (2011). Pharmacological importance of *Echinacea purpurea*. *International Journal of Pharma and Bio Sciences*, 2(4), 304-314.
- Kumar, S. A. & Pandey, S. A. (2015). An ethnobotanical study of local plants and their medicinal importance in Tons river area, Dehradun, Uttarakhand. *Indian Journal of Tropical Biodiversity*, 23(2), 227-231.
- Kumar, S., & Singh, A. (2019). An ethnobotanical, phytochemical and antioxidant activity of *Spinacia oleracea*, L. *Journal of Pharmaceutical Sciences and Research*, 11(7), 251-2525.
- Kurapati, K. R. V., Atluri, V. S., Samikkannu, T., Garcia, G., & Nair, M. P. (2016). Natural products as anti-HIV agents and role in HIV-associated neurocognitive disorders (HAND): a brief overview. *Frontiers in Microbiology*, 12(6), 1444. <https://doi.org/10.3389/fmicb.2015.01444>
- Kutluay, S. B., Doroghazi, J., Roemer, M. E., & Triezenberg, S. J. (2008). Curcumin inhibits herpes simplex virus immediate-early gene expression by a mechanism independent of p300/CBP histone acetyltransferase activity. *Virology*, 373(2), 239-247. <https://doi.org/10.1016/j.virol.2007.11.028>
- Lan, J., Ge, J., Yu, J., Shan, S., Zhou, H., Fan, S., & Wang, X. (2020). Structure of the SARS-CoV-2 spike receptor-binding domain bound to the ACE2 receptor. *Nature*, 581(7807), 215-220. <https://doi.org/10.1038/s41586-020-2180-5>
- Lee, C. D., Ott, M., Thyagarajan, S. P., Shafritz, D. A., Burk, R. D., & Gupta, S. (1996). *Phyllanthus amarus* down-regulates hepatitis B virus mRNA transcription and replication. *European Journal of Clinical Investigation*, 26(12), 1069-1076. <https://doi.org/10.1046/j.1365-2362.1996.410595>
- Lee-Huang, S., Zhang, L., Huang, P. L., Chang, Y. T., & Huang, P. L. (2003). Anti-HIV activity of olive leaf extract (OLE) and modulation of host cell gene expression by HIV-1 infection and OLE treatment. *Biochemical and Biophysical Research Communications*, 307(4), 1029-1037. [https://doi.org/10.1016/s0006-291x\(03\)01292-0](https://doi.org/10.1016/s0006-291x(03)01292-0)
- Li, S., Hattori, T., & Kodama, E. N. (2011). Epigallocatechin gallate inhibits the HIV reverse transcription step. *Antiviral Chemistry and Chemotherapy*, 21(6), 239-243. <https://doi.org/10.3851/IMP1774>
- Ling, J. X., Wei, F., Li, N., Li, J. L., Chen, L. J., Liu, Y. Y., & Yang, Z. Q. (2012). Amelioration of influenza virus-induced reactive oxygen species formation by epigallocatechin gallate derived from green tea. *Acta*

- Pharmacologica Sinica*, 33(12), 1533-1541. <https://doi.org/10.1038/aps.2012.80>
- Liu, G., Xiong, S., Xiang, Y. F., Guo, C. W., Ge, F., Yang, C. R., & Kitazato, K. (2011). Antiviral activity and possible mechanisms of action of pentagalloylglucose (PGG) against influenza A virus. *Archives of Virology*, 156(8), 1359-1369. <https://doi.org/10.1007/s00705-011-0989-9>
- Lu, G., Wang, Q., Gao, G. F. (2015) Bat-to-human: spike features determining 'host jump' of coronaviruses SARS-CoV, MERS-CoV, and beyond. *Trends in Microbiology*, 23(8), 468-478. <https://doi.org/10.1016/j.tim.2015.06.003>
- Lung, J., Lin, Y. S., Yang, Y. H., Chou, Y. L., Shu, L. H., Cheng, Y. C. & Wu, C. Y. (2020). The potential chemical structure of anti-SARS-CoV-2 RNA-dependent RNA polymerase. *Journal of Medical Virology*, 92(6), 693-697. <https://doi.org/10.1002/jmv.25761>
- Majumdar, D. D. (2013). Recent updates on pharmaceutical potential of plant protease inhibitors. *International Journal of Medicine and Pharmaceutical Sciences*, 3(4), 101-120.
- Manganelli, R. E. U., Zaccaro, L., & Tomei, P. E. (2005). Antiviral activity *in vitro* of *Urticadioica* L., *Parietariadiffusa* M. et K. and *Sambucus nigra* L. *Journal of Ethnopharmacology*, 98(3), 323-327. <https://doi.org/10.1016/j.jep.2005.01.021>
- Maurya, D. K. (2020). Evaluation of Yashtimadhu (*Glycycorrhiza glabra*) active phytochemicals against novel coronavirus (SARS-CoV-2). *Bioinformatics*. <https://doi.org/10.21203/rs.3.rs-26480/v1>
- Mir, A., Ismatullah, H., Rauf, S., & Niazi, U. H. (2016). Identification of bioflavonoid as fusion inhibitor of dengue virus using molecular docking approach. *Informatics in Medicine Unlocked*, 3, 1-6. <https://doi.org/10.1016/j.imu.2016.06.001>
- Mounce, B. C., Cesaro, T., Carrau, L., Vallet, T., & Vignuzzi, M. (2017). Curcumin inhibits Zika and chikungunya virus infection by inhibiting cell binding. *Antiviral Research*, 142, 148-157. <https://doi.org/10.1016/j.antiviral.2017.03.014>
- Mukherjee, R., Dash, P.K., & Ram, G. C. (2005). Immunotherapeutic potential of *Ocimum sanctum* (L) in bovine subclinical mastitis. *Research in Veterinary Science*, 79(1), 37-43. <https://doi.org/10.1016/j.rvsc.2004.11.001>
- Nadkarni, K. M., & Nadkarni, A. K. (1976). *Indian Materia Medica*. Bombay, India: Popular Prakashan Pvt. Ltd.
- Narayan, D., & Singh, P. K. (2017). Ethnobotanical importance and herbal medicine in Vindhya region of Eastern Uttar Pradesh, India. *Journal of Medicinal Plants Research*, 11(25), 403-413. <https://doi.org/10.5897/JMPR2017.6351>
- Narayanan, M. R., Mithunlal, S., Sujanapal, P., Kumar, N. A., Sivadasan, M., Alfarhan, A. H., & Alatar, A. A. (2011). Ethnobotanically important trees and their uses by Kattunaikka tribe in Wayanad Wildlife Sanctuary, Kerala, India. *Journal of Medicinal Plants Research*, 5(4), 604-612.
- Nasab, F. K., & Khosravi, A. R. (2014). Ethnobotanical study of medicinal plants of Sirjan in Kerman Province, Iran. *Journal of Ethnopharmacology*, 154(1), 190-197. <https://doi.org/10.1016/j.jep.2014.04.003>
- Nekooeian, A. A., Moatari, A., & Motamedifar, M. (2006). The antiinfluenza virus activity of hydroalcoholic extract of Olive leaves. *Iranian Journal of Pharmaceutical Sciences*, 2(3), 163-168.
- Niranjan Reddy, V. L., Malla Reddy, S., Ravikanth, V., Krishnaiah, P., Venkateswar Goud, T., Rao, T. P., & Venkateswarlu, Y. (2005). A new bis-andrographolide ether from *Andrographis paniculata* and evaluation of anti-HIV activity. *Natural Product Research*, 19(3), 223-230. DOI: 10.1080/14786410410001709197
- Notka, F., Meier, G., & Wagner, R. (2004). Concerted inhibitory activities of *Phyllanthus amarus* on HIV replication *in vitro* and *ex vivo*. *Antiviral Research*, 64(2), 93-102. <https://doi.org/10.1016/j.antiviral.2004.06.010>
- Nworu, C. S., Okoye, E. L., Ezeifeka, G. O., & Esimone, C. O. (2013). Extracts of *Moringa oleifera* Lam. showing inhibitory activity against early steps in the infectivity of HIV-1 lentiviral particles in a viral vector-based screening. *African Journal of Biotechnology*, 12(30), 4866-4873. <https://doi.org/10.5897/AJB2013.12343>
- Obata, O. O., & Aigbokhan, E. I. (2012). Ethnobotanical practices among the people of Oka-Akoko, Nigeria. *Plant Archives*, 12(2), 627-638.
- Oh, E. G., Kim, K. L., Shin, S. B., Son, K. T., Lee, H. J., Kim, T. H., & Kim, J. H. (2013). Antiviral activity of green tea catechins against feline calicivirus as a surrogate for norovirus. *Food Science and Biotechnology*, 22(2), 593-598. <https://doi.org/10.1007/s10068-013-0119-4>
- Olivieri, F., Prasad, V., Valbonesi, P., Srivastava, S., Ghosal-Chowdhury, P., Barbieri, L., & Stirpe, F. (1996). A systemic antiviral resistance-inducing protein isolated from *Clerodendrum inerme* Gaertn. is a polynucleotide: adenosine glycosidase (ribosome-inactivating protein). *FEBS letters*, 396(2-3), 132-134. [https://doi.org/10.1016/0014-5793\(96\)01089-7](https://doi.org/10.1016/0014-5793(96)01089-7)
- Onifade, A. A., Jewell, A. P., & Adeledeji, W. A. (2013). *Nigella sativa* concoction induced sustained seroreversion in HIV patient. *African Journal of Traditional, Complementary and Alternative Medicines*, 10(5), 332-335. <https://doi.org/10.4314/ajtcam.v10i5.18>
- Ooi, L. S., Ho, W. S., Ngai, K. L., Tian, L., Chan, P. K., Sun, S. S., & Ooi, V. E. (2010). *Narcissus tazetta* lectin shows strong inhibitory effects against respiratory syncytial virus, influenza A (H1N1, H3N2, H5N1) and B viruses. *Journal of Biosciences*, 35(1), 95-103. <https://doi.org/10.1007/s12038-010-0012-8>
- Ou, X., Liu, Y., Lei, X., Li, P., Mi, D., Ren, L., & Qian, Z. (2020). Characterization of spike glycoprotein of SARS-CoV-2 on virus entry and its immune cross-reactivity with SARS-CoV. *Nature communications*, 11(1), 1-12. <https://doi.org/10.1038/s41467-020-15562-9>
- Panvilai, S., Napaswad, C., Limthongkul, J., & Akkarawongsapat, R. (2021). Aqueous extracts of Thai medicinal plants possess anti-HIV-1 activity. *Journal of Herbs, Spices & Medicinal Plants*, 27(1), 1-10. <https://doi.org/10.1080/10496475.2020.1753276>
- Partha, P. (2014). Ethnobotany of the Laleng (Patra) Community in Bangladesh. *Journal of Pharmacognosy and Phytochemistry*, 2(6), 173-184.
- Patwardhan, B., Warude, D., Pushpangadan, P., & Bhatt, N. (2005). Ayurveda and traditional Chinese medicine: a comparative overview, *Evidence-based Complementary and Alternative Medicine*, 2(4), 465-473. <https://doi.org/10.1093/ecam/neh140>
- Penzak, S. R., Robertson, S. M., Hunt, J. D., Chairez, C., Malati, C. Y., Alfaro, R. M., Stevenson, J. M., & Kovacs, J. A. (2010). *Echinacea purpurea* significantly induces cytochrome P450 3A activity but does not alter lopinavir-ritonavir exposure in healthy subjects. *Pharmacotherapy: The Journal of Human Pharmacology and Drug Therapy*, 30(8), 797-805. <https://doi.org/10.1592/phco.30.8.797>
- Pleschka, S., Stein, M., Schoop, R., & Hudson, J. B. (2009). Anti-viral properties and mode of action of standardized *Echinacea purpurea* extract against highly pathogenic avian influenza virus (H5N1, H7N7) and swine-origin H1N1 (S-OIV). *Virology Journal*, 6(1), 1-9. <https://doi.org/10.1186/1743-422X-6-197>
- Polat, R., & Satil, F. (2012). An ethnobotanical survey of medicinal plants in Edremit Gulf (Balikesir-Turkey). *Journal of Ethnopharmacology*, 139(2), 626-641. <https://doi.org/10.1016/j.jep.2011.12.004>
- Pompeii, R., Flore, O., Marcialis, M. A., Pani, A., & Loddo, B. (1979). Glycyrrhetic acid inhibits virus growth and inactivates virus particles. *Nature*, 281, 689-690. <https://doi.org/10.1038/281689a0>
- Pongtuluran, O. B., & Rofaani, E. (2015). Antiviral and immunostimulant activities of *Andrographis paniculata*. *HAYATI Journal of Biosciences*, 22(2), 67-72. <https://doi.org/10.4308/hjb.22.2.67>
- Pruthivish, R., & Gopinatha, S. M. (2018). Antiviral prospective of *Tinosporacordifolia* on HSV-1. *International Journal of Current Microbiology and Applied Sciences*, 7(1), 3617-3624. <https://doi.org/10.20546/ijcmas.2018.701.425>
- Rahayu, S. M., & Andini, A. S. (2019). Ethnobotanical Study on Medicinal Plants in Sesao Forest, Narmada, West Lombok, Indonesia. *Biosaintika: Journal of Biology & Biology Education*, 11(2), 234-242. <https://doi.org/10.15294/biosaintika.v11i2.19314>
- Rajput, S. B., Tonge, M. B., & Karuppayil, S. M. (2014). An overview on traditional uses and pharmacological profile of *Acorus calamus* Linn. (Sweet flag) and other *Acorus* species. *Phytomedicine*, 21(3), 268-276. <https://doi.org/10.1016/j.phymed.2013.09.020>
- Ramya, S., Neethirajan, K., & Jayakumararaj, R. (2012). Profile of bioactive compounds in *Syzygiumcumini*-a review. *Journal of Pharmacy research*, 5(8), 4548-4553.
- Ranjan, P., Mohapatra, B., & Das, P. (2020). A rational drug designing: What bioinformatic approach tells about the wisdom of practicing traditional medicines for screening the potential of Ayurvedic and natural compounds for their inhibitory effect against COVID-19 Spike, Indian strain Spike, Papain-like protease and Main Protease protein. *Preprint*. <https://doi.org/10.21203/rs.3.rs-30366/v1>
- Rashid, A., & Marwat, S. K. (2006). Ethnobotanical study of important wild plants of Bahadur Khel tract (tehsil Banda Daud Shah) in Karak district. *Gomal Univ Journal of Biological Research*, 2(2), 165-172.

- Rashid, S., Ahmad, M., Zafar, M., Sultana, S., Ayub, M., Khan, M. A., & Yaseen, G. (2015). Ethnobotanical survey of medicinally important shrubs and trees of Himalayan region of Azad Jammu and Kashmir, Pakistan. *Journal of Ethnopharmacology*, 166, 340-351. <https://doi.org/10.1016/j.jep.2015.03.042>
- Rathinavel, T., Palanisamy, M., Palanisamy, S., Subramanian, A., & Thangaswamy, S. (2020). Phytochemical 6-Gingerol-A promising Drug of choice for COVID-19. *International Journal on Advanced Science, Engineering and Information Technology*, 6(4), 1482-9. <https://doi.org/10.29294/IJASE.6.4.2020.1482-1489>
- Ratia, K., Pegan, S., Takayama, J., Sleeman, K., Coughlin, M., Baliji, S., & Mesecar, A. D. (2008). A noncovalent class of papain-like protease/deubiquitinase inhibitors blocks SARS virus replication. *Proceedings of the National Academy of Sciences*, 105(42), 16119-16124. <https://doi.org/10.1073/pnas.0805240105>
- Ravikumar, Y. S., Ray, U., Nanditha, M., Perveen, A., Naika, H. R., Khanna, N., & Das, S. (2011). Inhibition of hepatitis C virus replication by herbal extract: *Phyllanthus amarus* as potent natural source. *Virus Research*, 158(1-2), 89-97. <https://doi.org/10.1016/j.virusres.2011.03.014>
- Ravishankar, B., & Shukla, V. J. (2007). Indian systems of medicine: a brief profile. *African Journal of Traditional, Complementary and Alternative Medicines*, 4(3), 319-337.
- Rehman, S., Ashfaq, U. A., Riaz, S., Javed, T., & Riazuddin, S. (2011). Antiviral activity of *Acacia nilotica* against Hepatitis C virus in liver infected cells. *Virology Journal*, 8(1), 1-6. <https://doi.org/10.1186/1743-422X-8-220>
- Robinson Jr, W. E. (1998). L-chicoric acid, an inhibitor of human immunodeficiency virus type 1 (HIV-1) integrase, improves on the *in vitro* anti-HIV-1 effect of Zidovudine plus a protease inhibitor (AG1350). *Antiviral Research*, 39(2), 101-111. [https://doi.org/10.1016/S0166-3542\(98\)00037-0](https://doi.org/10.1016/S0166-3542(98)00037-0)
- Romeilah, R. M., Fayed, S. A., & Mahmoud, G. I. (2010). Chemical compositions, antiviral and antioxidant activities of seven essential oils. *Journal of Applied Sciences Research*, 6(1), 50-62.
- Roschek Jr, B., Fink, R. C., McMichael, M. D., Li, D., & Alberte, R. S. (2009). Elderberry flavonoids bind to and prevent H1N1 infection *in vitro*. *Phytochemistry*, 70(10), 1255-1261. <https://doi.org/10.1016/j.phytochem.2009.06.003>
- Sagar, V., & Kumar, A. H. (2020). Efficacy of natural compounds from *Tinospora cordifolia* against SARS-CoV-2 protease, surface glycoprotein and RNA polymerase. *Virology*, 6(1), 1-10. <https://doi.org/10.21203/rs.3.rs-27375/v1>
- Saha, M. R., Kar, P., Sen, A., & De Sarker, D. (2014). Ethnobotany of Chanchal Block of Malda District of West Bengal (India): plants used in local healthcare. *Pleione*, 8(2), 381-390.
- Saha, S., & Ghosh, S. (2012). *Tinospora cordifolia*: one plant, many roles. *Ancient Science of Life*, 31(4), 151-159. <https://doi.org/10.4103/0257-7941.107344>
- Sahu, P. K., Kumari, A., Sao, S., Singh, M., & Pandey, P. (2013). Sacred plants and their Ethno-botanical importance in central India: A mini review. *International Journal of Pharmacy and Life Sciences*, 4(8), 439-446.
- Saikia, S., Bordoloi, M., Sarmah, R., & Kolita, B. (2019). Antiviral compound screening, peptide designing, and protein network construction of Influenza A virus (strain a/Puerto Rico/8/1934 H1N1). *Drug Development Research*, 80(1), 106-124. <https://doi.org/10.1002/ddr.21475>
- Sairam, K., Hemalatha, S., Kumar, A., Srinivasan, T., Ganesh, J., Shankar, M., & Venkataraman, S. (2003). Evaluation of anti-diarrhoeal activity in seed extracts of *Mangifera indica*. *Journal of Ethnopharmacology*, 84(1), 11-15. [https://doi.org/10.1016/S0378-8741\(02\)00250-7](https://doi.org/10.1016/S0378-8741(02)00250-7)
- Samy, R. P., Thwin, M. M., Gopalakrishnakone, P., & Ignacimuthu, S. (2008). Ethnobotanical survey of folk plants for the treatment of snakebites in Southern part of Tamil Nadu, India. *Journal of Ethnopharmacology*, 115(2), 302-312. <https://doi.org/10.1016/j.jep.2007.10.006>
- San Chang, J., Wang, K. C., Yeh, C. F., Shieh, D. E., & Chiang, L. C. (2013). Fresh ginger (*Zingiber officinale*) has anti-viral activity against human respiratory syncytial virus in human respiratory tract cell lines. *Journal of Ethnopharmacology*, 145(1), 146-151. <https://doi.org/10.1016/j.jep.2012.10.043>
- Saotoing, P., Vroumsia, P., Tchobolsala, Tchuenguem, F. F., Njan, N. A., & Messi, J. (2011). Medicinal plants used in traditional treatment of malaria in Cameroon. *Journal of Ecology and the Natural Environment*, 3(3), 104-117.
- Sathy, V., & Gopalakrishnan, V. K. (2013). *In-silico* ADMET prediction of phytochemicals in *Camellia sinensis* and *Citrus sinensis*. *International Journal of Pharmaceutical Sciences and Research*, 4(4), 1635-1637.
- Sen, P. (1993). Therapeutic potentials of Tulsi: from experience to facts. *Drugs News & Views*, 1(2), 15-21.
- Senthivel, P., Lavanya, P., Kumar, K. M., Swetha, R., Anitha, P., Bag, S., Sarveswari, S., Vijayakumar, V., Ramaiah, S., & Anbarasu, A. (2013). Flavonoid from *Carica papaya* inhibits NS2B-NS3 protease and prevents Dengue 2 viral assembly. *Bioinformation*, 9(18), 889-895. <https://doi.org/10.6026/97320630009889>
- Serkedjieva, J., Manolova, N., Zgórniak-Nowosielska, I., Zawilińska, B., & Grzybek, J. (1990). Antiviral activity of the infusion (SHS-174) from flowers of *Sambucus nigra* L., aerial parts of *Hypericum perforatum* L., and roots of *Saponaria officinalis* L. against influenza and herpes simplex viruses. *Phytotherapy Research*, 4(3), 97-100. <https://doi.org/10.1002/ptr.2650040305>
- Shapoo, G. A., Kaloo, Z. A., Ganie, A. H., & Singh, S. (2013). Ethnobotanical survey and documentation of some orchid species of Kashmir Himalaya, J&K-India. *International Journal of Pharmaceutical and Biological Research*, 4(2), 32-40.
- Sharma, N. (2019). Efficacy of Garlic and Onion against virus. *International Journal of Pharmaceutical Sciences Research*, 10(4), 3578-3586. <https://doi.org/10.26452/ijprs.v10i4.1738>
- Sharma, N., Mishra, K. P., Chanda, S., Bhardwaj, V., Tanwar, H., Ganju, L., & Singh, S. B. (2019). Evaluation of anti-dengue activity of *Carica papaya* aqueous leaf extract and its role in platelet augmentation. *Archives of Virology*, 164(4), 1095-1110. <https://doi.org/10.1007/s00705-019-04179-z>
- Shende, J. J., Rajurkar, B. M., Mhaiskar, M. N., & Dalal, L. P. (2014). Ethnobotanical studies of Samudrapur Tahsil of Wardha district. *IOSR Journal of Pharmacy and Biological Science*, 9(6), 16-23. <https://doi.org/10.9790/3008-09651623>
- Si, X., Wang, Y., Wong, J., Zhang, J., McManus, B. M., & Luo, H. (2007). Dysregulation of the ubiquitin-proteasome system by curcumin suppresses coxsackievirus B3 replication. *Journal of Virology*, 81(7), 3142-3150. <https://doi.org/10.1128/JVI.02028-06>
- Silva, J. K. R. D., Figueiredo, P. L. B., Byler, K. G., & Setzer, W. N. (2020). Essential oils as antiviral agents, potential of essential oils to treat SARS-CoV-2 infection: an *in-silico* investigation. *International Journal of Molecular Sciences*, 21(10), 3426. <https://doi.org/10.3390/ijms21103426>
- Singh, A., & Singh, P. K. (2009). An ethnobotanical study of medicinal plants in Chandauli District of Uttar Pradesh, India. *Journal of Ethnopharmacology*, 121(2), 324-329. <https://doi.org/10.1016/j.jep.2008.10.018>
- Singh, H. (2008). Importance of local names of some useful plants in ethnobotanical study. *Indian Journal of Traditional Knowledge*, 7(2), 365-370.
- Singh, H., Husain, T., Agnihotri, P., Pande, P. C., & Khatoon, S. (2014). An ethnobotanical study of medicinal plants used in sacred groves of Kumaon Himalaya, Uttarakhand, India. *Journal of Ethnopharmacology*, 154(1), 98-108. <https://doi.org/10.1016/j.jep.2014.03.026>
- Singh, Y. N. (1986). Traditional medicine in Fiji: some herbal folk cures used by Fiji Indians. *Journal of Ethnopharmacology*, 15(1), 57-88. [https://doi.org/10.1016/0378-8741\(86\)90104-2](https://doi.org/10.1016/0378-8741(86)90104-2)
- Song, W. Y., Ma, Y. B., Bai, X., Zhang, X. M., Gu, Q., Zheng, Y. T., & Chen, J. J. (2007). Two new compounds and anti-HIV active constituents from *Illicium verum*. *Planta Medica*, 73(04), 372-375. <https://doi.org/10.1055/s-2007-967162>
- Sood, R., Swarup D, Bhatia S., Kulkarni, D. D., Dey, S., Saini, M., & Dubey, S. C. (2012) Antiviral activity of crude extracts of *Eugenia jambolana* Lam. against highly pathogenic avian influenza (H5N1) virus. *Indian Journal of Experimental Biology*, 50(3), 179-186.
- Sorice, A., Guerriero, E., Capone, F., Colonna, G., Castello, G., & Costantini, S. (2014). Ascorbic acid: its role in immune system and chronic inflammation diseases. *Mini Reviews in Medicinal Chemistry*, 14(5), 444-452. <https://doi.org/10.2174/1389557514666140428112602>
- Srivastava, A. K., Kumar, A., & Misra, N. (2020). On the inhibition of COVID-19 protease by Indian herbal plants: an *in silico* investigation. *arXiv (Preprint)*, 2004, 03411. <https://doi.org/10.48550/arXiv.2004.03411>
- Sui, Z., Salto, R., Li, J., Craik, C., & de Montellano, P. R. O. (1993). Inhibition

- of the HIV-1 and HIV-2 proteases by curcumin and curcumin boron complexes. *Bioorganic & Medicinal Chemistry*, 1(6), 415-422. [https://doi.org/10.1016/S0968-0896\(00\)82152-5](https://doi.org/10.1016/S0968-0896(00)82152-5)
- Sujarwo, W., & Caneva, G. (2015). Ethnobotanical study of cultivated plants in home gardens of traditional villages in Bali (Indonesia). *Human Ecology*, 43(5), 769-778. <https://doi.org/10.1007/s10745-015-9775-8>
- Sun, Z., Yu, C., Wang, W., Yu, G., Zhang, T., Zhang, L., & Wei, K. (2018). Aloe polysaccharides inhibit influenza A virus infection—a promising natural anti-flu drug. *Frontiers in Microbiology*, 9, 2338-2338. <https://doi.org/10.1007/s10745-015-9775-8>
- Surkar, A., Lavania, S. C., Pandey, D. N., & Pant, M. C. (1994). Changes in the blood lipid profile after administration of *Ocimum sanctum* (Tulsi) leaves in the normal albino rabbits. *Indian Journal of Physiology and Pharmacology*, 38, 311-312.
- Tallei, T. E., Tumilaar, S. G., Niode, N. J., Fatimawali, Kepel, B. J., Idroes, R., Effendi, Y., Sakib, S. A., & Emran, T. B. (2020). Potential of plant bioactive compounds as SARS-CoV-2 main protease (Mpro) and spike (S) glycoprotein inhibitors: a molecular docking study. *Scientifica*, 2020, 6307457. <https://doi.org/10.1155/2020/6307457>
- Tamm, C. O. (1972). Survival and flowering of some perennial herbs. II. The behaviour of some orchids on permanent plots. *Oikos*, 23(1), 23-28. <https://doi.org/10.2307/3543923>
- Tang, L. I., Ling, A. P., Koh, R. Y., Chye, S. M., & Voon, K. G. (2012). Screening of anti-dengue activity in methanolic extracts of medicinal plants. *BMC Complementary and Alternative Medicine*, 12, 3. <https://doi.org/10.1186/1472-6882-12-3>
- Thomas, B., Mathews, R. P., Rajendran, A., & Kumar, K. P. (2013). Ethnobotanical observations on tribe Arnatans of nilambur forest, Western Ghats region of Kerala, India. *Research in Plant Biology*, 3(2), 12-17.
- Thompson, K. D. (1998). Antiviral activity of Viraceae against acyclovir susceptible and acyclovir resistant strains of herpes simplex virus. *Antiviral Research*, 39(1), 55-61. [https://doi.org/10.1016/S0166-3542\(98\)00027-8](https://doi.org/10.1016/S0166-3542(98)00027-8)
- Thuy, B. T. P., My, T. T. A., Hai, N. T. T., Hieu, L. T., Hoa, T. T., Loan, H. T. P., Triet, N. T., Anh, T. T. V., Quy, P. T., Tat, P. V., Hue, N. V., Quang, D. T., Trung, N. T., Tung, V. T., Huynh, L. K., & Nhung, N. T. A. (2020). Investigation into SARS-CoV-2 resistance of compounds in garlic essential oil. *ACS Omega*, 5(14), 8312-8320. <https://doi.org/10.1021/acsomega.0c00772>
- Tiwari, D., Upadhyay, S., & Paliwal, A. (2016). Survey of ethnomedicinal plants of Bharsar, *International Journal of Agricultural Sciences*, 6(6), 329-349.
- Tiwari, V., Darmani, N. A., Yue, B. Y., & Shukla, D. (2010). In vitro antiviral activity of neem (*Azadirachta indica* L.) bark extract against herpes simplex virus type-1 infection. *Phytotherapy Research*, 24(8), 1132-1140. <https://doi.org/10.1002/ptr.3085>
- Tsai, Y., Cole, L. L., Davis, L. E., Lockwood, S. J., Simmons, V., & Wild, G. C. (1985). Antiviral properties of garlic: *in vitro* effects on influenza B, herpes simplex and coxsackie viruses. *Planta Medica*, 51(05), 460-461. <https://doi.org/10.1055/s-2007-969553>
- Tyrrell, D. A., & Myint, S. H. (1996). Coronaviruses. *Medical Microbiology*. (4th ed.). Galveston: University of Texas Medical Branch at Galveston
- Ugulu, I. (2011). Traditional ethnobotanical knowledge about medicinal plants used for external therapies in Alasehir, Turkey. *International Journal of Medicinal and Aromatic Plants*, 1(2), 101-106.
- Ul Qamar, M. T., Maryam, A., Muneer, I., Xing, F., Ashfaq, U. A., Khan, F. A., & Siddiqi, A. R. (2019). Computational screening of medicinal plant phytochemicals to discover potent pan-serotype inhibitors against dengue virus. *Scientific reports*, 9(1), 1-16. <https://doi.org/10.1016/j.jpha.2020.03.009>
- Ulasli, M., Gurses, S. A., Bayraktar, R., Yumrutas, O., Oztuzcu, S., Igci, M., & Arslan, A. (2014). The effects of *Nigella sativa* (Ns), *Anthemishyalina* (Ah) and *Citrus sinensis* (Cs) extracts on the replication of coronavirus and the expression of TRP genes family. *Molecular Biology Reports*, 41(3), 1703-1711. <https://doi.org/10.1007/s11033-014-3019-7>
- Upadhyay, B., Dhaker, A. K., & Kumar, A. (2010). Ethnomedicinal and ethnopharmacological-statistical studies of Eastern Rajasthan, India. *Journal of Ethnopharmacology*, 129(1), 64-86. <https://doi.org/10.1016/j.jep.2010.02.026>
- van Damme, E. J., Balzarini, J., Smeets, K., van Leuven, F., & Peumans, W. J. (1994). The monomeric and dimeric mannose-binding proteins from the Orchidaceae species *Listera ovata* and *Epipactis helleborine*: sequence homologies and differences in biological activities. *Glycoconjugate Journal*, 11(4), 321-332.
- Varshney, A., Balkrishna, A., & Singh, J. (2020). Withanone from *Withaniasomnifera* May Inhibit Novel Coronavirus (COVID-19) Entry by Disrupting Interactions between Viral S-Protein Receptor Binding Domain and Host ACE2 Receptor. <https://doi.org/10.21203/rs.3.rs-17806/v1>
- Venkateswaran, P. S., Millman, I., & Blumberg, B. S. (1987). Effects of an extract from *Phyllanthus niruri* on hepatitis B and woodchuck hepatitis viruses: *in vitro* and *in vivo* studies. *Proceedings of the National Academy of Sciences*, 84(1), 274-278. <https://doi.org/10.1073/pnas.84.1.274>
- Vijayan, A., Liju, V. B., Reena John, J. V., Parthipan, B., & Renuka, C. (2007). Traditional remedies of Kani tribes of Kottoor reserve forest, Agasthyavanam, Thiruvananthapuram, Kerala. *Indian Journal of Traditional Knowledge*, 6(4), 589-594.
- Vimalanathan, S., & Hudson, J. (2012). Anti-Influenza virus activities of commercial oregano oils and their carriers. *Journal of Applied Pharmaceutical Science*, 2(7), 214-218. <https://doi.org/10.7324/JAPS.2012.2734>
- Wang, X. S., Dong, Q., Zuo, J. P., & Fang, J. N. (2003). Structure and potential immunological activity of a pectin from *Centella asiatica* (L.) Urban. *Carbohydrate Research*, 338(22), 2393-2402. [https://doi.org/10.1016/S0008-6215\(03\)00380-X](https://doi.org/10.1016/S0008-6215(03)00380-X)
- Weniger, B., Rouzier, M., Daguilh, R., Henrys, D., Henrys, J. H., & Anton, R. (1986). Traditional medicine in the Central Plateau of Haiti. 2. Ethnopharmacological inventory. *Journal of Ethnopharmacology*, 17(1), 13-30. [https://doi.org/10.1016/0378-8741\(86\)90070-x](https://doi.org/10.1016/0378-8741(86)90070-x)
- Wiart, C., Kumar, K., Yusof, M. Y., Hamimah, H., Fauzi, Z. M., & Sulaiman, M. (2005). Antiviral properties of ent-labdene diterpenes of *Andrographis paniculata* Nees, inhibitors of herpes simplex virus type 1. *Phytotherapy Research*, 19(12), 1069-1070. <https://doi.org/10.1002/ptr.1765>
- Wolkerstorfer, A., Kurz, H., Bachhofner, N., & Szolar, O. H. (2009). Glycyrrhizin inhibits influenza A virus uptake into the cell. *Antiviral Research*, 83(2), 171-178. <https://doi.org/10.1016/j.antiviral.2009.04.012>
- Wrapp, D., Wang, N., Corbett, K. S., Goldsmith, J. A., Hsieh, C. L., Abiona, O., & McLellan, J. S. (2020). Cryo-EM structure of the 2019-nCoV spike in the prefusion conformation. *Science*, 367(6483), 1260-1263. <https://doi.org/10.1126/science.abb2507>
- Xu, B., Wang, L., Gonzalez-Molleda, L., Wang, Y., Xu, J., & Yuan, Y. (2014). Antiviral activity of (+)-rutamarin against Kaposi's sarcoma-associated herpesvirus by inhibition of the catalytic activity of human topoisomerase II. *Antimicrobial Agents and Chemotherapy*, 58(1), 563-573. <https://doi.org/10.1128/AAC.01259-13>
- Yang, F., Zhang, Y., Tariq, A., Jiang, X., Ahmed, Z., Zhihao, Z., Idrees, M., Azizullah, A., Adnan, M., & Bussmann, R. W. (2020). Food as medicine: A possible preventive measure against coronavirus disease (COVID-19). *Phytotherapy Research*, 34(12), 3124-3136. <https://doi.org/10.1002/ptr.6770>
- Yang, Q. Y., Tian, X. Y., & Fang, W. S. (2007). Bioactive coumarins from *Boenninghausenia sessiliflora*. *Journal of Asian Natural Products Research*, 9(1), 59-65. <https://doi.org/10.1080/10286020500382397>
- Yasmin, A. R., Chia, S. L., Looi, Q. H., Omar, A. R., Noordin, M. M., & Ideris, A. (2020). Herbal extracts as antiviral agents. *Feed additives*, 2020, 115-132. <https://doi.org/10.1016/B978-0-12-814700-9.00007-8>
- Yousaf, T., Rafique, S., Wahid, F., Rehman, S., Nazir, A., Rafique, J., & Shah, S. M. (2018). Phytochemical profiling and antiviral activity of *Ajuga bracteosa*, *Ajuga parviflora*, *Berberis lycium* and *Citrus lemon* against Hepatitis C Virus. *Microbial Pathogenesis*, 118, 154-158. <https://doi.org/10.1016/j.micpath.2018.03.030>
- Yu, Y. B. (2004). The extracts of *Solanum nigrum* L. for inhibitory effects on HIV-1 and its essential enzymes. *Korean Journal of Oriental Medicine*, 10(1), 119-126.
- Zaher, K. S., Ahmed, W. M., & Zerizer, S. N. (2008). Observations on the biological effects of black cumin seed (*Nigella sativa*) and green tea (*Camellia sinensis*). *Global Veterinaria*, 2(4), 198-204.
- Zandi, K., Ramedani, E., Mohammadi, K., Tajbakhsh, S., Deilami, I., Rastian, Z., Fouladvand, M., Yousefi, F., & Farshadpour, F. (2010). Evaluation of antiviral activities of curcumin derivatives against HSV-1 in Vero cell line. *Natural Product Communications*, 5(12), 1935-1938.
- Zandi, K., Zadeh, M. A., Sartavi, K., & Rastian, Z. (2007). Antiviral activity of *Aloe vera* against herpes simplex virus type 2: an *in vitro* study. *African Journal of Biotechnology*, 6(15), 1770-1773.
- Zhang, L., Lin, D., Sun, X., Curth, U., Drosten, C., & Sauerhering, L.

- & Hilgenfeld, R. (2020). Crystal structure of SARS-CoV-2 main protease provides a basis for design of improved α -ketoamide inhibitors. *Science*, 368(6489), 409-412. <https://doi.org/10.1126/science.abb3405>
- Zhang, X. L., Guo, Y. S., Wang, C. H., Li, G. Q., Xu, J. J., Chung, H. Y., & Wang, G. C. (2014). Phenolic compounds from *Origanum vulgare* and their antioxidant and antiviral activities. *Food Chemistry*, 152, 300-306. <https://doi.org/10.1016/j.foodchem.2013.11.153>
- Zhen, H., Fang, F., Ye, D. Y., Shu, S. N., Zhou, Y. F., Dong, Y. S., & Li, G. (2006). Experimental study on the action of allitridin against human cytomegalovirus *in vitro*: Inhibitory effects on immediate-early genes. *Antiviral Research*, 72(1), 68-74. <https://doi.org/10.1016/j.antiviral.2006.03.017>
- Zhu, X. M., Song, J. X., Huang, Z. Z., Wu, Y. M., & Yu, M. J. (1993). Antiviral activity of mangiferin against herpes simplex virus type 2 *in vitro*. *Acta Pharmacologica Sinica*, 14(5), 452-454.