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Ethnomedicinal and pharmacological potential of marine macroalgae for CNS disorders: An overview

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ABSTRACT

Marine macroalgae or seaweeds have created a favourable implication in the area of biomedical sciences, due to the present of potential bioactive substances. Extensive studies are reported on neuropharmacological effects of terrestrial plants and their constituents but there is inadequate information on the potential application of marine macroalgae for behavioural and neurological disorders. This review will emphasize on recent studies and/or updates on bioactive compounds or extracts from marine macroalgae and their potential toward CNS disorders.

KEYWORDS: Marine macroalgae, Seaweed, CNS disorders

INTRODUCTION

Most medicines are obtained from natural sources and researchers are still searching the tropical rainforest for potentially high-priced medicinal products. Many products of sea origin were used as food supplement since early 20th century, like cod liver and shark liver oil. It was only in early years of 1950s that researchers began to scientifically probe marine flora for medicines. Till now, around 10,000 bioactive compounds have been explored from marine sources. The quest still continues to discover new bioactive compounds from oceans (Colwell, 2002; Proksch *et al.*, 2002).

Marine plants involve marine algae, mangroves, sea grasses and sand dune plants. Almost 90 percent of marine plants are marine algae (Dhargalkar & Pereira, 2005). Marine algae are of two types, macroalgae (seaweeds) and microalgae. Marine macroalgae are found in the coastal area between high tide to low tide or in the subtidal region. Marine macroalgae are of three types: Green algae (Chlorophyta), Brown algae (Phaeophyta) and Red algae (Rhodophyta) (Garson, 1989; El Gamal, 2010). Green algae (Chlorophyta) found in the fresh as well as marine habitats. It contains chlorophyll a & b as photosynthetic pigments. The photosynthetic product of these algae is starch, with ulvan being the major polysaccharide component. Brown algae (Phaeophyta) are found only in marine habitat. Photosynthetic pigments of these algae are carotenoid, fucoxanthin (pigment responsible for brown colour), chlorophyll a & c, carotene and xanthophylls.

The cell walls are made of cellulose and alginic acid. The photosynthesis of brown algae produced Laminarian, fucane and Manitol. Red algae (Rhodophyta) are exclusively marine (except for few species). Red algae contain phycoerythrin and phycothyanin (pigment responsible for red colour), chlorophyll a and b-carotene. The primary polysaccharides of these algae are agars and carrageenans (Bold & Wynne, 1978).

The macroalgae are used for human consumption in China and Japan. The countries like Singapore, Malaysia, Thailand, Indonesia, Korea used marine macroalgae in soup, salad or jelly. Marine macroalgae are loaded with proteins, soluble dietary fibers, minerals, polyunsaturated fatty acids, vitamins, antioxidants and essential amino acids (Dhargalkar & Pereira, 2005). Extensive studies are reported on the therapeutic prospective of marine macroalgae toward various diseases. We have previously published a review paper on therapeutic potential of various *Sargassum* species and their health benefits (Yende *et al.*, 2014). In present review article, CNS potential of various marine macroalgae are explored.

CNS Potential of Marine Macroalgae

As per WHO report, out of approximately 450 million people suffering from a mental or behavioural disorder, small number of them receive basic treatment, this accounts for 12.3 percent of the global burden of disease and will increase to 15 percent by the year 2020 (Herrera-Ruiz *et al.*, 2006). From last few

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decades, synthetic drugs have been increasingly prescribed as most effective in the treatment and management of neurological and psychological disorders, although extensive use of these synthetic drugs lead to a variety of unwanted pharmacological effects. However, synthetic drugs primarily alleviate the symptoms and give temporary relief. Therefore, development of cheap, effective and safe agents from plants and other natural sources becomes necessary to overcome these untoward effects. For long time, natural products have abundant use throughout the world for treatment and prevention of human diseases. The medicines from natural product obtained from various sources like marine plants and organisms, terrestrial plants, microorganisms, vertebrates and invertebrates (Newman *et al.*, 2000). Large numbers of studies are reported on neuropharmacological effects of terrestrial plants and their constituents but there is inadequate information on the potential application of components from marine plants for neuropsychiatric disorders. Recently, in the field of marine pharmacognosy, the development of marine natural products is promising as an important field for neuropharmacology. Currently, several drugs derived from marine organisms (worm and snail) are in the phase of clinical trials or used for treatment of schizophrenia, neuropathic pain and Alzheimer's disease (Alicino *et al.*, 2012). By considering various pharmacologically active components and previous reports on neuropharmacological effects of marine herbs (Smit, 2004), they can be considered as a promising source for development of novel neuropsychological drugs.

Effects of Marine Macroalgae in Behavioural Diseases

CNS depressant effects of the *Himanthalia elongata* extract was studied in mice (Table 1). *H. elongata* extract at dose of 300 mg/kg significantly prolonged sleep duration treated with barbiturate and delay pentylenetetrazol induced mortality. It also showed significant reductions in motor activity (Anca *et al.*, 1990).

Kamat *et al.* (1994) evaluated CNS activity of 69 marine plants of Indian coastal region. Among these, extract of *Caulerpa sertularioides* and *Sargassum tenerrimum* possesses CNS depressant activity. The methanolic extract of *C. sertularioides* and *S. tenerrimum* prolonged pentobarbital induced sleep and decreased spontaneous locomotor activities. Further, authors reported that *Ulva fasciata*, *Chnoospora implexa*, *Gelidiella acerosa*, *Centroceras clavulatum*, *Hypnea musciformis* and *Hypneac ericornis* showed increase spontaneous locomotor score and possess CNS stimulant activity.

Methanol extract of *Laminaria ochroleuca* was studied for CNS activities. The result of the study revealed that, *L. ochroleuca* significantly and dose dependently reduced spontaneous motor activity, reduced exploratory behaviour, reduced time on rota rod, reduced d-amphetamine-induced hyper-motility, decreased rectal temperature and decreased the reaction time in the hot plate analgesic test. Also, the extract showed significant enhancement in sleeping time induced by pentobarbital. Whereas, extract doesn't showed any protection against pentylenetetrazole induced convulsions (Vázquez-Freire *et al.*,

1994). Further, neuropharmacological effect of methanol extract of brown seaweed *Cystoseira usneoides* was studied. The extract significantly decreased spontaneous locomotor activity, reduced exploratory behaviour, reduced d- amphetamine induced hypermotility, reduced body temperature. Also, extracts showed significant effect on pentobarbital induced sleep and pentylenetetrazole induced seizures. Further, administration of extracts shows modification in level of dopamine, 3, 4-dihydroxy phenylacetic acid, 3-methoxytyramine and homovanilic acid in rat striatum (Vázquez-Freire *et al.*, 1995).

The effects of phlorotannins and eckstolonol (a major phlorotannin ingredient) obtained from ethanol extract of *Ecklonia cava* were evaluated for sleep wake profiles in C57BL/6N mice by measuring electroencephalograms (EEG) and Electromyograms (EMG) (Figure 1). Further, hypnotic mechanism of eckstolonol (1) was investigated by injecting Flumazenil. The results of this study revealed that, eckstolonol administration showed significant decrease in sleep latency and increase in non-rapid eye movement sleep (NREMS). Hypnotic effect of eckstolonol was completely abolished by pre-treatment with flumazenil. In conclusions, phlorotannins enhanced NREMS by modulating GABA_A receptor at benzodiazepine site. The results obtained in this study propose that phlorotannins can be used as potential sedative-hypnotics to cure insomnia (Cho *et al.*, 2014). Further, Yoon and Cho (2018) conducted similar study on triphlorethol A (2), major phlorotannin constituents and compared with zolpidem, a distinguished hypnotic drug. The result of this study showed significant sleep-promoting effect of triphlorethol A at a dose of 50 mg/kg.

Cho *et al.* (2012) reported that, ethanol extract of *Ecklonia cava* at 1000 mg/kg showed increased in pentobarbital-induced sleep. Also, at 500 mg/kg it showed marked anticonvulsant activity in mice against picrotoxin induced seizure. The anxiolytic effects of sulphated polysaccharides isolated from *Gracilaria cornea* (red algae) was evaluated using elevated plus maze (EPM), hole board, open field and rotarod test. Also, its mechanism through benzodiazepine receptor was evaluated. It observed that, isolated sulphated polysaccharides at 10 mg/kg showed significant modification all the parameters observed in the EPM test. Further, it showed non-significant change in motor activity observed in open field test and rotarod test. Also, at 10 mg/kg it significantly increased number of head dips when tested using the hole board test. An increased expression of alpha 2-gamma-aminobutyric acid type A (alpha 2-GABA(A)) receptor in the hippocampus was observed in diazepam treated as well as with isolated sulphated polysaccharides treated group. In conclusion, sulphated polysaccharides from *G. cornea* (10 mg/kg) showed an anxiolytic effect, observed via alpha 2-GABA(A) receptor (Monteiro *et al.*, 2016).

The neuropharmacological properties of seaweeds from the genus *Turbinaria*, *Sargassum*, *Dictyota*, *Ulva* and *Padina* were studied using behavioural and electrophysiological techniques. The extracts of these seaweeds were evaluated as anti-cholinergic and anti-glutamatergic agents using snail receptors. Further, results showed sedative as well as anxiolytic action of *Dictyota*'s extracts and anticonvulsant effects for *Sargassum*, *Turbinaria*, *Dictyota* and *Padina* extract (Nuñez *et al.*, 2006).

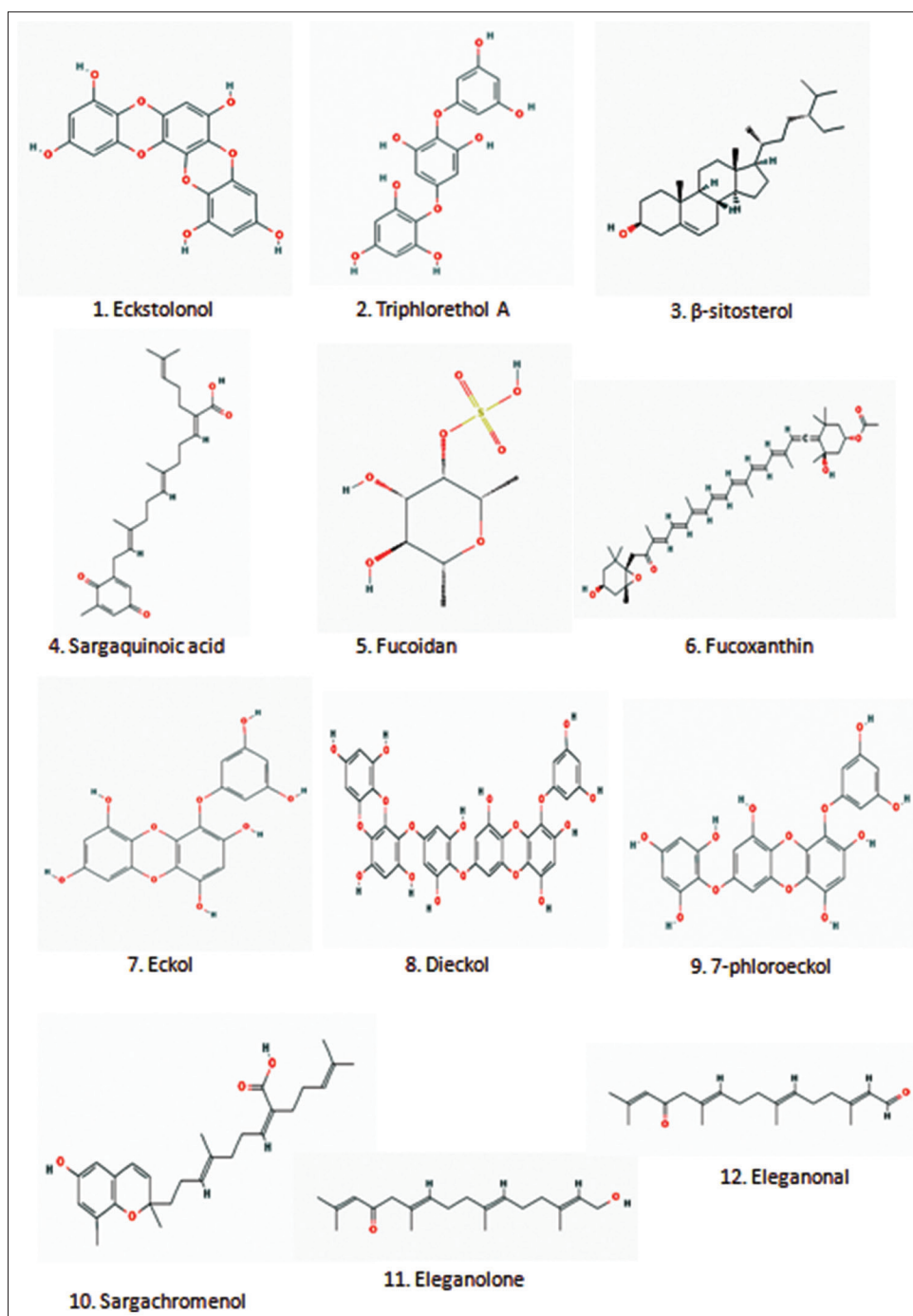


Figure 1: Chemical structures of some phytoconstituents of marine macroalgae (Source-<https://pubchem.ncbi.nlm.nih.gov/>)

Antidepressant activity, anticonvulsant activity and anxiolytic effect of extracts of *Sargassum ilicifolium* was screened using animal experimental models. The results of these studies showed that ethanol and chloroform extract at 400 mg/kg and 600 mg/kg exhibited significant antidepressant, anxiolytic and anticonvulsant activity (Yende & Harle, 2013; Yende *et al.*, 2016a; Yende *et al.*, 2018). Further, cognitive enhancing and antioxidant activities of *Sargassum ilicifolium* and *Padina tetraströmatica* are reported in scopolamine treated mice (Yende *et al.*, 2021). Yende *et al.* (2016b) reported anticonvulsant activity and anxiolytic effect of *Padina tetraströmatica*. Ethanol extract

at 400 and 600 mg/kg and chloroform extract at 600 mg/kg of *P. tetraströmatica* showed significant anxiolytic effect in elevated plus maze and light/dark exploration tests. Whereas, chloroform extract at 600 mg/kg showed significant activity against maximal electroshock and pentylenetetrazole induced convulsion in mice.

A study was carried out as antidepressant like effect of *Sargassum swartzii*, *Stoehospermum marginatum* and *Nizamuddinina zanardinii* using forced swimming test in rats. It was found that, oral injection of methanolic extracts (30–60 mg/kg) showed reduction in immobility time, without any significant change in locomotion in open field

Table 1: CNS activity of extracts or components of marine macroalgae

Algal source	Extract/Component	Activity	Reference
<i>H. elongata</i>		CNS depressant effects Sedative – Hypnotic activity	(Anca <i>et al.</i> , 1990)
<i>C. sertularioides</i> , <i>S. tenerrimum</i> <i>U. fasciata</i> , <i>C. implexa</i> , <i>G. acerosa</i> , <i>C. clavulatum</i> , <i>H. musciformis</i> <i>H. ervicornis</i> <i>L. ochroleuca</i> <i>C. usneoides</i> <i>E. cava</i>	Methanolic extract	CNS depressant activity, Sedative – Hypnotic activity CNS stimulant activity	(Kamat <i>et al.</i> , 1994)
	Methanol extract	CNS depressant activity	(Vázquez-Freire <i>et al.</i> , 1994)
	Methanol extract	Neuropharmacological effect	(Vázquez-Freire <i>et al.</i> , 1995)
	Phlorotannins and eckstolonol	Hypnotic activity	(Cho <i>et al.</i> , 2014)
	Triphlorethol A	Hypnotic activity	(Yoon & Cho, 2018)
	Ethanol extract	Sedative- hypnotic activity, Anticonvulsant activity	(Cho <i>et al.</i> , 2012)
<i>G. cornea</i> Genus <i>Turbinaria</i> , <i>Sargassum</i> , <i>Dictyota</i> , <i>Ulva</i> and <i>Padina</i> <i>S. ilicifolium</i>	Sulphated polysaccharides	Anxiolytic effect Neuropharmacological activities	(Monteiro <i>et al.</i> , 2016) (Nuñez <i>et al.</i> , 2006)
	Ethanol and Chloroform extract	Antidepressant activity, Anticonvulsant activity, Anxiolytic activity	(Yende <i>et al.</i> , 2016a; Yende & Harle, 2013; Yende <i>et al.</i> , 2018)
<i>P. tetrastromatica</i>	Ethanol and Chloroform extract	Anticonvulsant activity, Anxiolytic activity	(Yende <i>et al.</i> , 2016a; 2016b)
<i>S. swartzii</i> , <i>S. marginatum</i> , and <i>N. azanardinii</i> <i>S. horneri</i> <i>U. pinnatifida</i> <i>G. gracilis</i>	Methanolic extract	Antidepressant activity	(Siddiqui <i>et al.</i> , 2017)
	Total sterols and β -sitosterol	Antidepressant activity	(Zhao <i>et al.</i> , 2016)
	Ethanol extract	Neuroprotective activity	(Choi <i>et al.</i> , 2018)
	Methanolextract	Anti AChE activity, Anti BuChE activity	(Natarajan <i>et al.</i> , 2009)
<i>S. sagamianum</i>	Farnesylacetone derivatives, sargaquinoic acid and sargachomenol	Anti AChE activity, Anti BuChE activity	(Choi <i>et al.</i> , 2007)
<i>S. fusiforme</i> <i>S. siliquastrum</i> , <i>S. horneri</i> <i>E. stolonifera</i>	Fucoxanthin	Neuroprotective activity	(Hu <i>et al.</i> , 2016)
	Fucoxanthin	Antioxidant activity, anti AChE activity	(Heo <i>et al.</i> , 2008)
	Eckol, dieckol, eckstolonol, phlorofucofuroeckol, 2-phloroecol and 7-phloroecol	Anti AChE activity, Anti BuChE activity	(Yoon <i>et al.</i> , 2008)
<i>E. cava</i>	Dieckol, phlorofucofuroeckol and phlorotannins	Anti AChE activity	(Myung <i>et al.</i> , 2005)
<i>S. macrocarpum</i> , <i>S. fulvellum</i>	Sargachromenol, sargaquinoic acid, Pheophytin A	Neuroprotective activity, Enhanced nerve growth factor in rat pheochromocytoma (PC12) cell line	(Kamei & Tsang 2003; Ina <i>et al.</i> , 2007)
<i>U. conglobata</i> <i>B. bifurcate</i>	Methanol extracts Eleganolone and Eleganonal	Neuroprotective activity Antioxidant and neuroprotective activity	(Jin <i>et al.</i> , 2006) (Silva <i>et al.</i> , 2019)

test. Further this study extended by investigating involvement of monoaminergic system using inhibitors of brain serotonin, dopamine and adrenaline. The results of this study suggested the involvement of monoamine system in antidepressant like activity of the methanolic extracts (Siddiqui *et al.*, 2017). A similar study was reported by Zhao *et al.* (2016) where total sterols and β -sitosterol (3) was isolated from brown algae *Sargassum horneri* and its antidepressant-like activity was investigated. It was found that, total sterols and β -sitosterol significantly decreased immobility time when evaluated using forced swimming test and tail suspension test. Further, it significantly increased brain norepinephrin, serotonin and the serotonin metabolite, 5- hydroxyindoleacetic acid level.

Effects of Marine Macroalgae on Neurological Disorders

It was reported that ethanol extracts of *U. pinnatifida* enhanced cognitive impairment when evaluated using passive

avoidance test. The extract increased latency time and restored spine density and morphology in hippocampal neurons of scopolamine-induced rats (Choi *et al.*, 2018). Natarajan et al, reported that methanol extract of *Sargassum* and *Gracilaria gracilis* showed strong inhibition on acetylcholinesterase (AChE) and butyrylcholinesterase (BuChE) activity (Natarajan *et al.*, 2009). Also, two farnesylacetone derivatives from *Sargassum sagamianum* possess moderate inhibition on acetylcholinesterase and butyrylcholinesterase activity (Ryu *et al.*, 2003). Further sargaquinoic acid (4) and sargachomenol (meroterpenes) isolated from *S. sagamianum* Yendo showed marked anti acetylcholinesterase and anti butyrylcholinesterase activity (Choi *et al.*, 2007).

Fucoxanthin (5) (sulphated polysaccharides) isolated from brown algae, was shown protective effect on A β -induced cholinergic neuronal loss in rat. It may be due to inhibition of the A β -

induced ROS generation, or due to activation of caspase-9 and caspase-3 (mediators of terminal stages of neuronal apoptosis) (Jhamandas *et al.*, 2005). A fucoidan extracted from *Sargassum fusiforme*, showed improvement in spatial learning and memory when screened using scopolamine and ethanol induced cognitive deficit (Hu *et al.*, 2016).

Fucoxanthin (6), a carotenoid extracted from *Sargassum siliquastrum* significantly inhibit free radical formation and apoptosis induced by H₂O₂. It shows reduction in oxidative damage and inflammation in microglial cells (Heo *et al.*, 2008). Recently, Lin *et al.* reported that, Fucoxanthin obtained from *Sargassum horneri*, significantly reversed impairments of memory induced by scopolamine. Further, it significantly increased choline acetyltransferase activity and Brain derived neurotrophic fact (BDNF) expression. Also, fucoxanthin shows anti AChE activity in scopolamine treated mice (Lin *et al.*, 2016).

Phlorotannins, algal polyphenols, are prominent therapeutically active compounds. Yoon *et al.* (2008) isolated Eckol (7), dieckol (8), eckstolonol (1), phlorofuocofuroeckol, 2-phloroeckol and 7-phloroeckol (9) from *Ecklonia stolonifera* Okamura, these compounds exhibit dose-dependent inhibition in AChE and BuChE. Also, anti-AChE activity of Dieckol, phlorofuocofuroeckol and phlorotannins from *Ecklonia cava* was reported (Myung *et al.*, 2005). Further, anti neuro inflammatory activity of dieckol extracted from *E. Cava* was also evaluated (Jung *et al.*, 2009).

Tsang *et al.* (2005) isolated sargachromenol (10), a plastoquinone (terpens) from *Sargassum macrocarpum*. It was found that, sargachromenol enhanced nerve growth factor (NGF) dependent neurogenesis. This study was carried out in rat pheochromocytoma (PC12) cell line, which is an experimental model for the neuroprotective study of neurotrophic factors, like NGF. Further, it was reported that sargaquinoic acid (4) extracted from *S. macrocarpum*, increased neuritis outgrowth when studied in PC12D cells (Kamei & Tsang, 2003). Also, Pheophytin A (Chlorophylls) obtained from *Sargassum fulvellum* significantly increased neuritis outgrowth in PC12 cells (Ina *et al.*, 2007).

The neuroprotective activity of *Ulva conglobata* on glutamate-induced neurotoxicity in the murine hippocampal HT22 cell line and the anti-inflammatory effects on interferon gamma (IFN- γ)-induced microglial activation in BV2 cells was evaluated. It was found that, methanol extracts of *U. conglobata* showed significant attenuation in the neurotoxicity induced by glutamate in HT22 cells and inhibit nitric oxide production induced by IFN- γ in BV2 cells. Further, it suppressed the expression of the cyclooxygenase 2 and nitric oxide synthase (pro inflammatory enzyme), which suggest that *U. conglobata* possesses neuroprotective activity associated with neuroinflammation (Jin *et al.*, 2006).

Seven fractions (F1–F7) and two major diterpenes, eleanolone (11) and eleanonal were extracted from dichloromethane extract of *Bifurcaria bifurcata* and evaluated for antioxidant and neuroprotective activity. The neuroprotective activity was evaluated by 6-hydroxydopamine induced neurotoxicity model using human neuroblastoma cell line. It was observed that, fractions F4 and F5

showed the potent neuroprotective and antioxidant activities, whereas, fractions F4, eleanolone (11) and eleanonal (12) showed exhibited antioxidant activity (Silva *et al.*, 2019).

CONCLUSION

In extensive studies of marine macroalgae, CNS depressant/stimulant activities, sedative-hypnotic activities, anticonvulsant, antidepressant, anxiolytic effects, antioxidant, anticholinesterase and neuroprotective activities have also been reported. This scientific evidence shows that marine macroalgae have significant CNS potential. Further, the extensive pharmacological actions coupled with compounds derived from these seaweeds such as phlorotannins, alginates, β -sitosterol, fucoidan, sargachromenol, sargaquinoic acid, fucoxanthin *etc* increases the prospective to expand the neuropharmacological potential and health beneficial values of marine macroalgae. In conclusion, marine macroalgae are a valuable resource for the formation of novel ingredients which can be useful for the treatment of CNS disorders.

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REFERENCES

- Alicino, I., Giglio, M., Manca, F., Bruno, F., & Puntillo, F. (2012). Intrathecal combination of ziconotide and morphine for refractory cancer pain: A rapidly acting and effective choice. *Pain*, 153(1), 245–249. <https://doi.org/10.1016/j.pain.2011.10.002>
- Anca, J. M., Lamela, M., Cadavid, I., & Calleja, J. M. (1990). Effects of *Himantalia elongata* on the central nervous system of mice. *Journal of Ethnopharmacology*, 29(2), 225–231. [https://doi.org/10.1016/0378-8741\(90\)90059-3](https://doi.org/10.1016/0378-8741(90)90059-3)
- Bold, H. C., & Wynne, M. J. (1978). *Introduction to the Algae : Structure and Reproduction*. Prentice-Hall.
- Cho, S., Yang, H., Jeon, Y. J., Lee, C. J., Jin, Y. H., Baek, N. I., Kim, D., Kang, S. M., Yoon, M., Yong, H., Shimizu, M., & Han, D. (2012). Phlorotannins of the edible brown seaweed *Ecklonia cava* Kjellman induce sleep via positive allosteric modulation of gamma-aminobutyric acid type A-benzodiazepine receptor: A novel neurological activity of seaweed polyphenols. *Food Chemistry*, 132(3), 1133–1142. <https://doi.org/10.1016/j.foodchem.2011.08.040>
- Cho, S., Yoon, M., Pae, A. N., Jin, Y. H., Cho, N. C., Takata, Y., Urade, Y., Kim, S., Kim, J. S., Yang, H., Kim, J., Kim, J., Han, J. K., Shimizu, M., & Huang, Z. L. (2014). Marine polyphenol phlorotannins promote non-rapid eye movement sleep in mice via the benzodiazepine site of the GABAA receptor. *Psychopharmacology*, 231(14), 2825–2837. <https://doi.org/10.1007/s00213-014-3445-1>
- Choi, B. W., Ryu, G., Park, S. H., Kim, E. S., Shin, J., Roh, S. S., Shin, H. C., & Lee, B. H. (2007). Anticholinesterase activity of plastoquinones from *Sargassum sagamianum*: lead compounds for alzheimer's disease therapy. *Phytotherapy Research*, 21(5), 423–426. <https://doi.org/10.1002/ptr.2090>
- Choi, J. Y., Mohibbullah, M., Park, I. S., Moon, I. S., & Hong, Y. K. (2018). An ethanol extract from the phaeophyte *Undaria pinnatifida* improves learning and memory impairment and dendritic spine morphology in hippocampal neurons. *Journal of Applied Phycology*, 30(1), 129–136. <https://doi.org/10.1007/s10811-017-1116-4>
- Colwell, R. R. (2002). Fulfilling the promise of biotechnology. *Biotechnology Advances*, 20(3), 215–228. [https://doi.org/https://doi.org/10.1016/S0734-9750\(02\)00011-3](https://doi.org/https://doi.org/10.1016/S0734-9750(02)00011-3)
- Dhargalkar, V. K., & Pereira, N. (2005). Seaweed : Promising Plant of the Millennium. *Science and Culture*, 71(3–4), 60–66.

- El Gamal, A. A. (2010). Biological importance of marine algae. *Saudi Pharmaceutical Journal*, 18(1), 1–25. <https://doi.org/10.1016/j.jsps.2009.12.001>
- Garson, M. J. (1989). Biosynthetic studies on marine natural products. *Natural Product Reports*, 6(2), 143–170. <https://doi.org/10.1039/NP9890600143>
- Heo, S., Ko, S. C., Kang, S. M., Kang, H. S., Kim, J. P., Kim, S., Lee, K., Cho, M., & Jeon, Y. (2008). Cytoprotective effect of fucoxanthin isolated from brown algae *Sargassum siliquastrum* against H₂O₂-induced cell damage. *European Food Research and Technology*, 228(1), 145–151. <https://doi.org/10.1007/s00217-008-0918-7>
- Herrera-Ruiz, M., García-Beltrán, Y., Mora, S., Díaz-Véliz, G., Viana, G. S. B., Tortoriello, J., & Ramírez, G. (2006). Antidepressant and anxiolytic effects of hydroalcoholic extract from *Salvia elegans*. *Journal of Ethnopharmacology*, 107(1), 53–58. <https://doi.org/10.1016/j.jep.2006.02.003>
- Hu, P., Li, Z., Chen, M., Sun, Z., Ling, Y., Jiang, J., & Huang, C. (2016). Structural elucidation and protective role of a polysaccharide from *Sargassum fusiiforme* on ameliorating learning and memory deficiencies in mice. *Carbohydrate Polymers*, 139, 150–158. <https://doi.org/10.1016/j.carbpol.2015.12.019>
- Ina, A., Hayashi, K. I., Nozaki, H., & Kamei, Y. (2007). Pheophytin a, a low molecular weight compound found in the marine brown alga *Sargassum fulvellum*, promotes the differentiation of PC12 cells. *International Journal of Developmental Neuroscience*, 25(1), 63–68. <https://doi.org/10.1016/j.ijdevneu.2006.09.323>
- Jhamandas, J. H., Wie, M. B., Harris, K., MacTavish, D., & Kar, S. (2005). Fucoidan inhibits cellular and neurotoxic effects of β -amyloid (A β) in rat cholinergic basal forebrain neurons. *European Journal of Neuroscience*, 27(10), 2649–2659. <https://doi.org/10.1111/j.1460-9568.2005.04111.x>
- Jin, D. Q., Lim, C. S., Sung, J. Y., Choi, H. G., Ha, I., & Han, J.-S. (2006). *Ulva conglobata*, a marine algae, has neuroprotective and anti-inflammatory effects in murine hippocampal and microglial cells. *Neuroscience Letters*, 402(1), 154–158. <https://doi.org/10.1016/j.neulet.2006.03.068>
- Jung, W. K., Heo, S. J., Jeon, Y. J., Lee, C. M., Park, Y. M., Byun, H. G., Choi, Y. H., Park, S. G., & Choi, I. W. (2009). Inhibitory Effects and Molecular Mechanism of Dieckol Isolated from Marine Brown Alga on COX-2 and iNOS in Microglial Cells. *Journal of Agricultural and Food Chemistry*, 57(10), 4439–4446. <https://doi.org/10.1021/jf9003913>
- Kamat, S. Y., Wahidulla, S., D'Souza, L., Naik, C. G., Ambiyee, V., Bhakuni, D. S., Jain, S., Goel, A. K., Srimal, R. C. (1994). Bioactivity of marine organisms: Part VII—Effect of seaweed extract on central nervous system. *Indian Journal of Experimental Biology*, 32(6), 418–422.
- Kamei, Y., & Tsang, C. K. (2003). Sargaquinoic acid promotes neurite outgrowth via protein kinase A and MAP kinases-mediated signaling pathways in PC12D cells. *International Journal of Developmental Neuroscience*, 21(5), 255–262. [https://doi.org/10.1016/S0736-5748\(03\)00068-6](https://doi.org/10.1016/S0736-5748(03)00068-6)
- Lin, J., Huang, L., Yu, J., Xiang, S., Wang, J., Zhang, J., Yan, X., Cui, W., He, S., & Wang, Q. (2016). Fucoxanthin, a Marine Carotenoid, Reverses Scopolamine-Induced Cognitive Impairments in Mice and Inhibits Acetylcholinesterase in Vitro. *Marine Drugs*, 14(4), 67. <https://doi.org/10.3390/md14040067>
- Monteiro, V. S., Teles, F. B., Coura, C. O., Souza, R. B., Lima, C. N. de C., Costa, D. V. da S., Honório Junior, E. R., Escudeiro, S. de S., Chaves, E. M. C., Vasconcelos, S. M. M., & Benevides, N. M. B. (2016). Involvement of the GABAergic system in the anxiolytic effect of sulfated polysaccharides from the red seaweed *Gracilaria cornea*. *Journal of Applied Phycology*, 28(3), 1997–2004. <https://doi.org/10.1007/s10811-015-0724-0>
- Myung, C. S., Shin, H. C., Bao, H. Y., Yeo, S. J., Lee, B. H., & Kang, K. J. (2005). Improvement of Memory by Dieckol and Phlorofucoxanthin in Ethanol-Treated Mice Possible Involvement of the Inhibition of Acetylcholinesterase. *Archives of Pharmacological Research*, 28(6), 691–698. <https://doi.org/10.1007/bf02969360>
- Natarajan, S., Shanmugiahthevar, K. P., & Kasi, P. D. (2009). Cholinesterase inhibitors from *Sargassum* and *Gracilaria gracilis*: Seaweeds inhabiting South Indian coastal areas (Hare Island, Gulf of Mannar). *Natural Product Research*, 23(4), 355–369. <https://doi.org/10.1080/14786410802156036>
- Newman, D. J., Cragg, G. M., & Snader, K. M. (2000). The influence of natural products upon drug discovery. *Natural Product Reports*, 17(3), 215–234. <https://doi.org/10.1039/A902202C>
- Nuñez, R., Garateix, A., Laguna, A., Fernández, M. D., Ortiz, E., Llanio, M., Valdés, O., Rodríguez, A., & Menéndez, R. (2006). Caribbean marine biodiversity as a source of new compounds of biomedical interest and others industrial applications. *Pharmacologyonline*, 3, 111–119. <https://www.oceandocs.org/handle/1834/3649>
- Proksch, P., Edrada, R., & Ebel, R. (2002). Drugs from the seas – current status and microbiological implications. *Applied Microbiology and Biotechnology*, 59(2), 125–134. <https://doi.org/10.1007/s00253-002-1006-8>
- Ryu, G., Park, S. H., Kim, E. S., Choi, B. W., Ryu, S. Y., & Lee, B. H. (2003). Cholinesterase inhibitory activity of two farnesylacetone derivatives from the brown alga *Sargassum sagamianum*. *Archives of Pharmacological Research*, 26(10), 796–799. <https://doi.org/10.1007/BF02980022>
- Siddiqui, P. J. A., Khan, A., Uddin, N., Khalique, S., Rasheed, M., Nawaz, S., Hanif, M., & Dar, A. (2017). Antidepressant-like deliverables from the sea: evidence on the efficacy of three different brown seaweeds via involvement of monoaminergic system. *Bioscience, Biotechnology, and Biochemistry*, 81(7), 1369–1378. <https://doi.org/10.1080/09168451.2017.1313697>
- Silva, J., Alves, C., Freitas, R., Martins, A., Pinteus, S., Ribeiro, J., Gaspar, H., Alfonso, A., & Pedrosa, R. (2019). Antioxidant and Neuroprotective Potential of the Brown Seaweed *Bifurcaria bifurcata* in an in vitro Parkinson's Disease Model. *Marine Drugs*, 17(2), 85. <https://doi.org/10.3390/md17020085>
- Smit, A. J. (2004). Medicinal and pharmaceutical uses of seaweed natural products: A review. *Journal of Applied Phycology*, 16(4), 245–262. <https://doi.org/10.1023/B:JAPH.0000047783.36600.ef>
- Tsang, C. K., Ina, A., Goto, T., & Kamei, Y. (2005). Sargachromenol, a novel nerve growth factor-potentiating substance isolated from *Sargassum macrocarpum*, promotes neurite outgrowth and survival via distinct signaling pathways in PC12D cells. *Neuroscience*, 132(3), 633–643. <https://doi.org/10.1016/j.neuroscience.2005.01.028>
- Vázquez-Freire, M. J., Castro, E., Lamela, M., & Calleja, J. M. (1995). Neuropharmacological effects of *Cystoseira usneoides* extract. *Phytotherapy Research*, 9(3), 207–210. <https://doi.org/10.1002/ptr.2650090311>
- Vázquez-Freire, M. J., Lamela, M., & Calleja, J. M. (1994). *Laminaria ochroleuca*: A preliminary study of its effect on the central nervous system. *Phytotherapy Research*, 8(7), 422–425. <https://doi.org/10.1002/ptr.2650080709>
- Yende, S., Harle, U., & Chaugule, B. (2014). Therapeutic potential and health benefits of *Sargassum* species. *Pharmacognosy Reviews*, 8(15), 1–7. <https://doi.org/10.4103/0973-7847.125514>
- Yende, S. R., Harle, U. N., & Ittadwar, A. M. (2016a). Anxiolytic activity of marine macroalgae *Sargassum ilicifolium* and *Padina tetrastratica* in mice. *International Journal of Pharmacy and Pharmaceutical Sciences*, 8(5), 97–101.
- Yende, S. R., Harle, U. N., & Ittadwar, A. M. (2016b). Insignificant anticonvulsant activity of *Padina tetrastratica* (Brown macroalgae) in mice. *Journal of Pharmaceutical Negative Results*, 7(1), 33. <https://doi.org/10.4103/0976-9234.177061>
- Yende, S. R., Arora, S. K., & Ittadwar, A. M. (2021). Antioxidant and Cognitive Enhancing Activities of *Sargassum ilicifolium* and *Padina tetrastratica* in Scopalamine Treated Mice. *Journal of Biologically Active Products from Nature*, 11(1), 11–21. <https://doi.org/10.1080/22311866.2020.1871071>
- Yende, S. R., & Harle, U. N. (2013). Antidepressant-like effect of *Sargassum ilicifolium* in mice model of depression. *Advances in Pharmacology & Toxicology*, 14(3), 7–13.
- Yende, S. R., Harle, U. N., Arora, S. K., & Pande, V. B. (2018). Phytochemical screening and anticonvulsant activity of *Sargassum ilicifolium* (brown algae) in mice. *The Journal of Phytopharmacology*, 7(1), 25–28.
- Yoon, M., & Cho, S. (2018). Triphlorethol A, a Dietary Polyphenol from Seaweed, Decreases Sleep Latency and Increases Non-Rapid Eye Movement Sleep in Mice. *Marine Drugs*, 16(5), 139. <https://doi.org/10.3390/md16050139>
- Yoon, N. Y., Chung, H. Y., Kim, H. R., & Choi, J. S. (2008). Acetyl- and butyrylcholinesterase inhibitory activities of sterols and phlorotannins from *Ecklonia stolonifera*. *Fisheries Science*, 74(1), 200–207. <https://doi.org/10.1111/j.1444-2906.2007.01511.x>
- Zhao, D., Zheng, L., Qi, L., Wang, S., Guan, L., Xia, Y., & Cai, J. (2016). Structural Features and Potent Antidepressant Effects of Total Sterols and β -sitosterol Extracted from *Sargassum horneri*. *Marine Drugs*, 14(7), 123. <https://doi.org/10.3390/md14070123>