



ISSN: 2455-0485

Walsura robusta Roxb. (Meliaceae), a little-known tree with a rich limonoid profile

Christian Bailly*

OncoWitan, Lille (Wasquehal), 59290, France

ABSTRACT

The plant *Walsura robusta* Roxb. (Meliaceae) is a robust tree largely distributed in south-east Asia, including provinces of southern China. A few traditional usages of the plant have been mentioned, notably for the treatment of microbial infections. But experimental studies using different types of plant extracts only revealed modest antibacterial effects, and no major antiparasitic activity. *Walsura robusta* Roxb. is a rich source of secondary metabolites. Several series of limonoids have been isolated from the leaves or the fruits of the plant, such as walsuronoid A-I, walsurins A-E, walsunoids A-I, walrobsins A-R and other cedrelone- or dihydrocedrelone-type limonoids, in addition to a few other terpenoids. All information about *Walsura robusta* Roxb. have been collated in this brief review. The analysis underlines the presence of two limonoids endowed with significant anticancer activities, walsuronoid B and cedrelone. They both activate the production of reactive oxygen species in cancer cells, modulate mitochondrial activities and induce apoptosis of cancer cells. Their molecular targets and mechanism of action are discussed. *Walsura robusta* Roxb. has a potential for the development of anticancer natural products. The use of the plant extracts could be further considered for the treatment of diseases with a cell proliferation component.

KEY WORDS: Anticancer agents, limonoids, Meliaceae, natural products, *Walsura robusta*.

Received: April 08, 2021

Accepted: May 19, 2021

Published: July 03, 2021

*Corresponding author:

Christian Bailly

Email: christian.bailly@

oncowitan.com

WALSURA ROBUSTA ROXB.: A ROBUST TREE

The plant genus *Walsura* comprises about 50 different names but there are only 4 accepted specie names: *W. pinnata* Hassk., *W. trichostemon* Miq., *W. villosa* Wall. ex Hiern. and *W. robusta* Roxb (<http://www.theplantlist.org>). There are other commonly used names, such as *W. cochinchinensis* (Baill.) Harms and *W. yunnanensis* C.Y. Wu but they are both synonyms for *Walsura pinnata* Hassk. These plants, belonging to the *Meliaceae* family, are usually found in subtropical regions of Southern China, India, Indonesia, and the Asean member countries.

The specie *Walsura robusta* Roxb. (synonym: *Surwala robusta* (Roxb.) M. Roem.) is an evergreen large tree which can usually grow 20-30 meters tall, with large leaves (Fig. 1). The outer bark is grey-brown whereas the inner bark is pink-red, with a specific wood anatomy. The wood of *W. robusta* presents a much larger vessel pore density (308/sq.mm) than the wood of *W. trifoliolata*, *W. tubulata*, and *W. chrysogyne* (at 81-154/sq.mm). The fruits of *W. robusta* Roxb. are non-edible, in contrast to those of *W. pinnata* Hassk. which are sweet-tasting brown-orange fleshy berries. From a taxonomic point of view, *W. robusta* belongs to a distinct subgroup compared to other *Walsura* species (Clark,

1994). The tree can be found in south-east Asia, notably in Vietnam (where it is called *lông tong*), Cambodia, Thailand, Laos, Myanmar, and Malaysia. There is an old mention that the tree is commonly grown in Chittagong (Ghosh, 1968) and in the Rajkandi reserve forest of Moulvibazar, (Bangladesh) (Haque *et al.*, 2018). There may be confusion about the name of the plant because several studies mention that the vernacular Bengla name for *W. robusta* is in *bonlichu* (or *bonlichu*), whereas other studies refer to *bonlichu* as a totally different tree, *Xerospermum laevigatum* Radlk. (Pasha and Uddi, 2019; Abdullah *et al.*, 2020). *W. robusta* Roxb. is found also in the south of China, notably in the Yunnan, Guangxi, Guangdong, and Hainan provinces (Sakkat, 2007).

EXTRACTS OF WALSURA ROBUSTA ROXB. AND THEIR USES

There are a few reports mentioning the traditional use of extracts of *W. robusta* Roxb. to treat various diseases and conditions. In Thailand, extracts of the leaves and twigs of the plant (called *bonlichu*) are used to treat microbial infections and diarrhea. In China, it may be named name “*geshesu*” as mentioned in a study (Hou *et al.*, 2013), although it is possible that this Chinese

Copyright: © The authors. This article is open access and licensed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0/>) which permits unrestricted, use, distribution and reproduction in any medium, or format for any purpose, even commercially provided the work is properly cited. Attribution — You must give appropriate credit, provide a link to the license, and indicate if changes were made.



Figure 1: The plant *Walsura robusta* Roxb. (Meliaceae) and its geographic breakdown. (a) the tree. (b) leaves and fruits (<http://tracuuduoclieu.vn/walsura-robusta-roxb.html>). (c) The plant can be found in Vietnam, Laos, Cambodia, Thailand, Myanmar, Malaysia, Bhutan, Bangladesh, and in provinces of south China (Yunnan, Guangxi, Guangdong, and Hainan) (<http://www.plantsoftheworldonline.org/taxon/urn:lsid:ipni.org:names:579946-1>)

name refers in fact to *W. yunnanensis*, as cited in another study (Luo *et al.*, 2006). *W. robusta* seem to be used in traditional medicine in diverse countries in Asia, but there is a lack of robust information about this traditional usage. Moreover, there is also little experimental evidence to support the traditional medicinal use of *W. robusta*. Initially, a modest antibacterial activity was reported with an ethanolic plant extract found to inhibit the growth of a clinical isolate of methicillin-resistant *Staphylococcus aureus* (MRSA) and the *S. aureus* ATCC strain

25923 with a minimum inhibitory concentration (MIC) of 1.6 mg/ml (Voravuthikunchai and Kitpip, 2005a, 2005b). A better level of activity was observed using wood extracts of the plant against the enterohaemorrhagic *Escherichia coli* strain O157:H7, with MIC of 0.09 and 0.19 mg/ml for the aqueous and ethanolic extracts, respectively (Voravuthikunchai *et al.*, 2004) but a later study reported a marginal bacteriostatic and bactericidal activities with a similar extract (Voravuthikunchai and Limsuwan, 2006). In another study, an aqueous extract of *W. robusta* was found to mildly inhibit the growth of Gram-positive and some Gram-negative bacteria, including *E. coli*. In the same study, the plant extract was tested against trophozoites of the intestinal parasite *Giardia intestinalis* but no anti-giardial activity was observed (Voravuthikunchai *et al.*, 2010). Clearly, there is not enough experimental data to support the traditional use of the plant as an anti-infectious agent.

PHYTOCHEMICAL ANALYSES OF WALSURA ROBUSTA ROXB.

Diverse types of secondary metabolites have been isolated from *W. robusta*, in particular limonoids which are heavily oxygenated, modified triterpenes frequently encountered in Meliaceae and Rutaceae (*Citrus* species) and to a lesser extent in Cneoraceae and Simaroubaceae (Tan and Luo, 2011; Tundis *et al.*, 2014; Zhang and Xu, 2017). The major limonoids in *W. robusta* are named walsuronoids A-C, initially isolated from the leaves of the plant (Yin *et al.*, 2007). Walsuronoid A (Fig. 2) bears a seco limonoid skeleton with a 3,4-peroxide bridge and walsuronoids B and C possess an 18(13→14)-abeo-limonoid skeleton. Walsuronoid C has been considered as an oxidation product of walsuronoid B because its furyl ring is easily oxidized when exposed to air in chloroform. A modest growth inhibition of the malaria parasite *Plasmodium falciparum* was observed with walsuronoids A and B (40% inhibition at the dose of 40 μ M) (Yin *et al.*, 2007). Walsuronoid B was also identified from the fruits of the plant together with a series of linonoids, known as walsuronoid F-I and walsurins A-E (Fig. 2) and many other terpenoids (Zhang *et al.*, 2017).

Walsuronoid B deserves a special mention because it has been shown to exert potent anticancer effects *in vitro* and *in vivo* (Geng *et al.*, 2017). The compound dose-dependently inhibited the proliferation of several types of cancer cells, with IC_{50} in the range of 3-4 μ M. Walsuronoid B blocked cell cycle progression of HepG2 and Bel-7402 liver cancer cells (G_2/M arrest), with a concomitant up-regulation of cyclin B1 and down-regulation of (phospho)-cdc55C. The compound was found to trigger a massive apoptosis of these malignant cells, with activation of caspases and a drug-induced dysfunction of mitochondria and lysosomes (Fig. 3a). The molecular target of the drug is not known at present, but the compound enhanced the production of reactive oxygen species (ROS) and elevated the expression of the tumor-suppressor protein p53. Interestingly, walsuronoid B (at the intraperitoneal dose of 4 mg/kg) displayed a marked anticancer activity in mice with xenografted HepG2 liver tumors, with an efficacy comparable to that of the standard drug cisplatin (Geng *et al.*, 2017). The activation of the ROS/

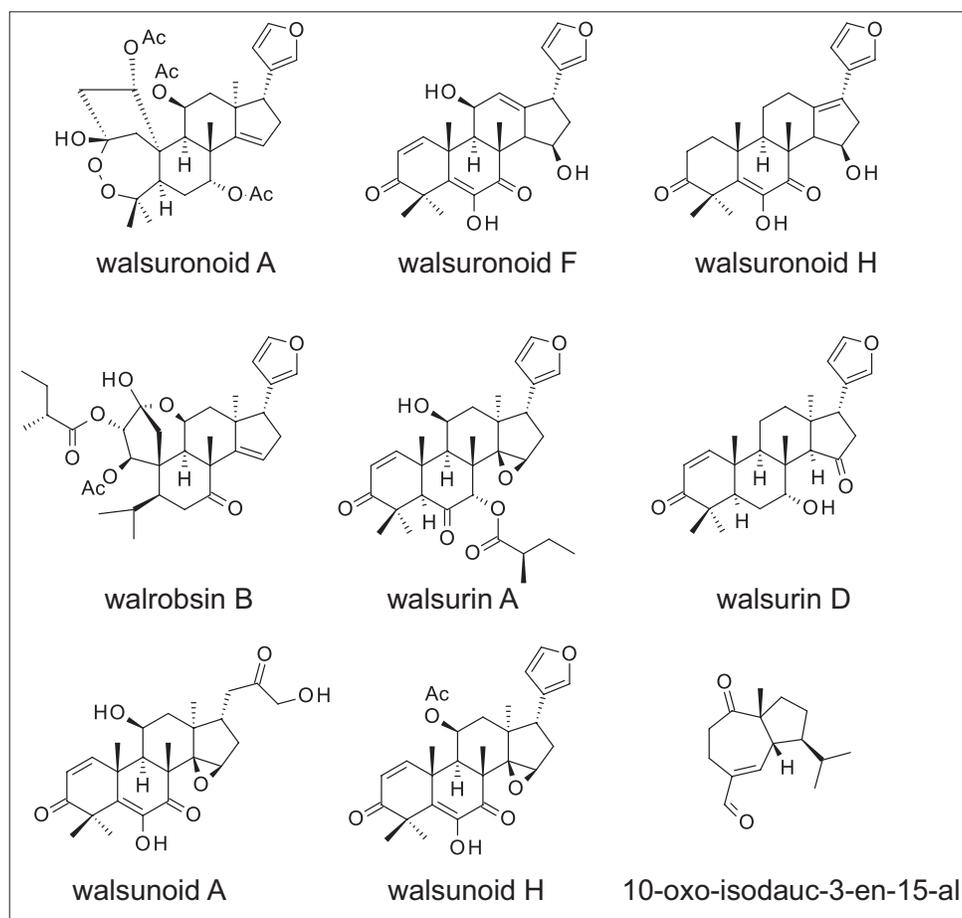


Figure 2: Chemical structures of selected natural products isolated from *Walsura robusta* Roxb

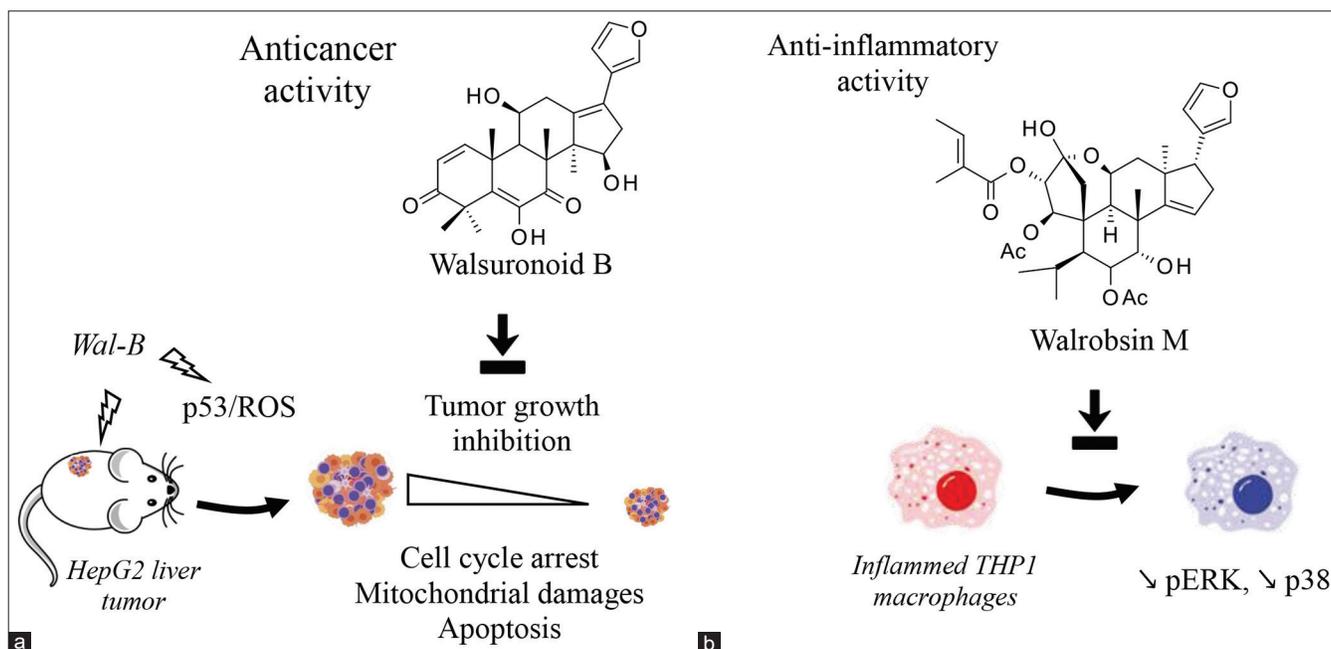


Figure 3: Schematic representation of (a) the anticancer activity of walsuronoid B and (b) the anti-inflammatory action of walrobsin M. Walsuronoid B activates the production of reactive oxygen species (ROS) and the tumor suppressor protein p53, leading to apoptotic cell death (Geng *et al.*, 2017). Walrobsin M represses the expression of the phosphorylated signaling proteins ERK (extracellular signal-regulated kinase) and p38 in THP-1-like macrophages (stimulated with the skin pathogen *Propionibacterium acnes*), thereby inhibiting the MAPK signaling pathway (An *et al.*, 2019)

p53 signaling pathway is central to the mechanism of action of walsuronoid B, as it is the case for other anticancer plant natural products like the cardiac glycoside odoroside A from the plant *Nerium oleander* Linn. (Chen *et al.*, 2019), betulinic acid (Shen *et al.*, 2017) and gambogic acid (Liang and Zhang, 2016) for examples. There is no doubt that walsuronoid B warrants further studies to better define its mode of anticancer action, tumor selectivity and molecular targets.

Another small group of limonoids isolated from *W. robusta* has been named walsunoids A-I, isolated together with different cedrelone-type limonoids such as 11 β -hydroxyisowalsuranolide and 11 β -hydroxydihydrocedrelone (Wang *et al.*, 2016). These compounds were tested against the human 11 β -hydroxysteroid dehydrogenase type 1 (11 β -HSD1), the enzyme which reduces inert cortisone into active cortisol, and which is frequently implicated in metabolic diseases (obesity, diabetes). A modest inhibition of human 11 β -HSD1 was observed with walsunoid H (Fig. 2) *in vitro* (IC₅₀ = 9.9 μ M) (Wang *et al.*, 2016).

Two other analogues with an atypical 5-oxatricyclo[5.4.1^{1,4}] hendecane ring system have been isolated from the root bark of the plant. They were named walrobsins A and B (Fig. 2). These two compounds showed no cytotoxic activity, but a modest anti-inflammatory activity was reported with walrobsin A. The compound was found to inhibit the release of the pro-inflammatory cytokine interleukin-1 β from lipopolysaccharide-activated RAW264.7 macrophages and to dose-dependently inhibit the expression of the inducible nitric oxide synthase (iNOS) enzyme, the enzyme responsible for nitric oxide (NO) production (An *et al.*, 2017). A subsequent phytochemical analysis from the root bark of *W. robusta* led to the identification of other compounds with a 5-oxatricyclohendecane ring system and named walrobsins C-to-R (An *et al.*, 2019). The bioactivity of these compounds has not been deeply investigated but a preliminary evaluation indicated that walrobsin M (Fig. 3b) presents an interesting anti-inflammatory profile, with a drug-induced down-regulation of the expression of protein phospho-ERK and phospho-p38 in inflamed THP-1 macrophage cells (An *et al.*, 2019).

The fruits of the plant also contain a series of compounds structurally similar to cedrelone (Fig. 2) and dihydrocedrelone, which were found to potentiate the cytotoxicity of the anticancer drug doxorubicin in human MCF-7/DOX breast cancer cells, normally resistant to the drug. These compounds could efficiently reverse the multidrug resistance (MDR) phenotype *in vitro* (Zhang *et al.*, 2017). Cedrelone limonoids have been isolated from *W. robusta*, *W. yunnanensis* (Ji *et al.*, 2014), *Toona sinensis* (Jiang *et al.*, 2020) and from the leaves of *Trichilia americana*, another plant of the Meliaceae family (Ji *et al.*, 2015). Acetylation of the cedrelone core affords cedrelone acetate, a synthetic molecule presenting enhanced cytotoxic properties and efficient to revert the malignant phenotype of cancer cells (Becceneri *et al.*, 2020). Cedrelone itself is an interesting anticancer agent because this compound was found to activate the expression of the protein PBLD (phenazine biosynthesis-like domain-containing protein) which is often down-regulated in hepatocellular carcinoma (HCC) (Wu

et al., 2019). PBLD functions as a tumor suppressor. An increase of the expression of PBLD could reduce HCC cell growth and invasion via inactivation of several tumorigenesis-related signaling pathways (Li *et al.*, 2016). Through the activation of PBLD in cells, cedrelone regulates the Ras and Ras-proximate-1 (Rap1) signaling pathways and this signaling action triggers apoptosis of cancer cell, while reducing cell proliferation and the epithelial-mesenchymal transition (Wu *et al.*, 2019). The antitumor capacity of cedrelone has been characterized in different experimental models, notably using drug-resistant human glioma cells (Cao *et al.*, 2019) and leukemia cells (Wang *et al.*, 2019). In both cases, the compound selectively affected the ERK/MAPK signaling pathway. A direct interaction of cedrelone with the ERK1 kinase has been postulated, based on a computational docking analysis. The compound was predicted to bind to a small pocket on the protein, leading to the kinase activation (Fig. 4). Similarly, a combination of docking and molecular dynamic simulations has suggested that cedrelone could bind to the multi domain ceramide transfer protein (CERT), a protein implicated in sphingolipid metabolism and which allows the transport of ceramide from the endoplasmic reticulum to the Golgi apparatus (Fig. 4). Cedrelone would bind to the linked PH and START domains of CERT, thereby inhibiting the protein (Ghoula *et al.*, 2020). Inhibition of CERT is considered as an appropriate mechanism to re-sensitize cancer cells to chemotherapy (Palau *et al.*, 2018; Kumagai and Hanada, 2019). The anticancer effects of cedrolone have been evidenced using other cell types, including MDA-MB-231 breast cancer cells, NCI-H460 non-small cell lung cancer cells and A375 melanoma cells (Cazal *et al.*, 2010; Fuzer *et al.*, 2013). Cedrolone is an interesting natural product which also displays anti-fungal and insecticidal activities (Govindachari *et al.*, 1995, 2000). It exhibits a sub-micromolar activity against the protozoan parasite *Cryptosporidium parvum in vitro* (EC₅₀ = 0.27 μ M) (Jin *et al.*, 2019) and causes lethal and sublethal effects on the armyworm *Spodoptera frugiperda* (Giongo *et al.*, 2016).

Other terpenes and sesquiterpenes have been isolated from *W. robusta*, such as the carotane sesquiterpene 10-oxo-isodauc-3-en-15-al (Fig. 2) and its nitro derivative, both isolated from a methanolic extract of the plant leaves. These two compounds showed no antimicrobial activity against *Staphylococcus aureus* and different MRSA strains (Hou *et al.*, 2013). It is worth to note that this isodaucane type sesquiterpenoid can also be found in the leaves of the sunflower crop (*Helianthus annuus* L.) and it has revealed a significant herbicidal activity. Indeed, 10-oxo-isodauc-3-en-15-al was found to inhibit coleoptile elongation and is considered as a useful allelopathic compound (Fuentes-Gandara *et al.*, 2019). It has been found in diverse plants, such as *Senecio crassiflorus* (Pox.) De Candolle (Jares and Pomillo, 1989), *Uvaria lucida* (Moriyasu *et al.*, 2012), *Chromolaena laevigata* (Misra *et al.*, 1985) and *Conza linifolia* (Hussein *et al.*, 1995). Surprisingly, this compound and its C-10 epimer which is known as sinulin A, can be found also in a marine organism, the Xisha soft coral *Sinularia* sp. (Qin *et al.*, 2018).

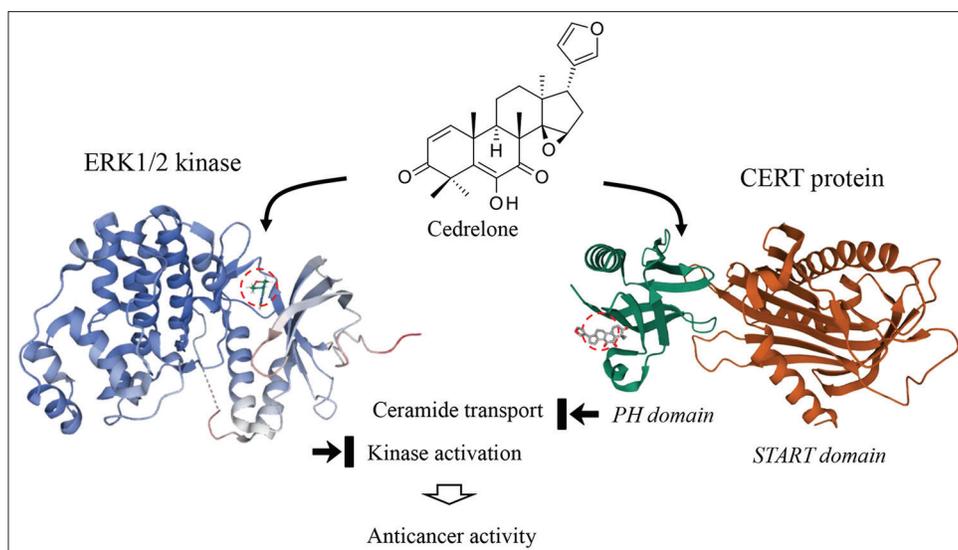


Figure 4: Two potential protein targets of the limonoid cedrelone, which can be isolated from *Walsura robusta* Roxb. The molecular model (left) shows a drug fragment bound to the ERK1/2 kinase, to modulate the phosphorylation and catalytic activity of the enzyme (PDB: 6G91) (Heightman *et al.*, 2018). A molecular modeling analysis has predicted that cedrelone can bind to the same site of the protein, thereby activating the kinase (Wang *et al.*, 2019). Molecular modeling has also predicted that cedrelone can bind to the PH domain of the multi domain ceramide transfer protein (CERT) to inhibit its transport activity (Ghoula *et al.*, 2020). Binding of the compound to ERK1/2, CERT and other proteins likely contributes to the anticancer activity of this limonoid

CONCLUSION

Plants of the *Walsura* genus have been little studied thus far. For examples, the PubMed data bank comprises 30 references with the name *Walsura*, compared to >2000, 300 and 150 for the genera *Trichilia*, *Aglaiia* and *Xylocarpus*, which also belong to the *Meliaceae* family (as of April 2021). Moreover, in the *Walsura* genus, the specie *W. pinnata* Hassk (synonyms: *W. cochinchinensis* (Baill.) Harms and *W. yunmanensis* C.Y. Wu) is more frequently studied than *W. robusta* Roxb. This is perhaps not surprising because the plant is not so frequently used in traditional medicine, except in very local areas. *W. robusta* trees should be investigated further, for the usefulness of their woods and the diversity of the secondary metabolites which can be isolated from the bark, leaves and fruits of the plant.

As presented here, *W. robusta* Roxb. provides a rich reservoir of limonoids, a huge family of natural products widely distributed in nature. Limonoids can be isolated from many plants, and some of them can be (semi) synthesized (Zhang and Xu, 2017; Fu and Liu, 2020). They are particularly abundant in *Meliaceae* (Tan and Luo, 2011). The great majority of the new natural products isolated from *W. robusta* over the past few years are limonoids, such as the walsuronoids, walrobsins, walsurins, and walsunoids. Beyond the isolation and characterization of these compounds, the bioactivity measurements have been limited mainly to a few preliminary *in vitro* tests, notably to evaluate their antimicrobial activity. No outstanding activity has been reported, but occasionally a few compounds revealed a modest antibacterial or anti-inflammatory effect. Two compounds emerge from this literature survey: walsuronoid B and cedrelone, because they both present noticeable anticancer properties against multiple cancer cell lines. Walsuronoid B stands as a

potent anticancer agent active in a xenograft model *in vivo* and cedrelone also displays marked anticancer activities. They both impact mitochondrial activities, cause an increase of ROS in cells, and trigger apoptosis of cancer cells (Geng *et al.*, 2017; Cao *et al.*, 2019). They could be considered as prototypes for the design of novel anticancer derivatives (Becceneri *et al.*, 2020).

Plants of the *Walsura* genus, and the specie *W. robusta* Roxb. in particular, have received little attention thus far. This robust tree, well distributed in south Asia, should be considered further in the search of natural products of medicinal interest. We can certainly recommend the investigation of extracts of the plant and its abundant fruits, for the treatment of cancers, and possibly other diseases with a cell proliferation component. The specie deserves its named “*robusta*”, as a robust provider of bioactive limonoids.

FUNDING

This research did not receive any specific grant from funding agencies in the public, commercial, or not-for-profit sectors.

DECLARATION OF COMPETING INTEREST

The author declares no conflict of interest associated with this publication and there has been no significant financial support for this work that could have influenced its outcome.

REFERENCES

- Abdullah, M. R., Haque, M., Sarwar, A. G., Ashrafuzzaman, M., & Rahman, M. (2020). Diversity of underutilized fruits and their uses in Karnaphuli range, Rangamati, Bangladesh. *International Journal of*

- Forestry, Ecology and Environment*, 1, 10-20.
- An, F., Wang, X., Yang, M., Luo, J., & Kong, L. (2019). Bioactive A-ring rearranged limonoids from the root barks of *Walsura robusta*. *Acta Pharmaceutica Sinica B*, 9, 545-556.
- An, F. L., Sun, D. M., Li, R. J., Zhou, M. M., Yang, M. H., Yin, Y., Kong, L. Y., & Luo, J. (2017). Walrobins A and B, Two Anti-inflammatory Limonoids from root barks of *Walsura robusta*. *Organic Letters*, 19, 4568-4571.
- Becceneri, A. B., Fuzer, A. M., Popolin, C. P., Cazal, C. M., Domingues, V. C., Fernandes, J. B., Vieira, P. C., & Cominetti, M. R. (2020). Acetylation of cedrelone increases its cytotoxic activity and reverts the malignant phenotype of breast cancer cells in 3D culture. *Chemico-Biological Interactions*, 316, 108920.
- Cao, Y., Zhang, L., & Wang, Y. (2019). Antitumor activity of Cedrelone in temozolomide-resistant human glioma cells is accompanied by mitochondrial mediated apoptosis, inhibition of angiogenesis, cell cycle disruption and modulation of ERK/MAPK signalling pathway. *Journal of the Balkan Union of Oncology*, 24, 1204-1209.
- Cazal, C. M., Choosang, K., Severino, V. G., Soares, M. S., Sarria, A. L., Fernandes, J. B., Silva, M. F., Vieira, P. C., Pakkong, P., Almeida, G. M., Vasconcelos, M. H., Nascimento, M. S., & Pinto, M. M. (2010). Evaluation of effect of triterpenes and limonoids on cell growth, cell cycle and apoptosis in human tumor cell line. *Anti-cancer Agents in Medicinal Chemistry*, 10(10), 769-776.
- Chen, Y. Y., Wen, S. Y., Deng, C. M., Yin, X. F., Sun, Z. H., Jiang, M. M., & He, Q. Y. (2019). Proteomic analysis reveals that Odoroside A triggers G2/M arrest and apoptosis in colorectal carcinoma through ROS-p53 pathway. *Proteomics*, 19(15), e1900092
- Clark, T. P. (1994). The species of *Walsura* and *Pseudoclausena* genus novum (Meliaceae). *Blumea*, 38, 247-302.
- Fu S, Liu B. (2020). Recent progress in the synthesis of limonoids and limonoid-like natural products. *Organic Chemistry Frontiers*, 7, 1903-1947.
- Fuentes-Gandara, F., Torres, A., Fernández-Ponce, M. T., Casas, L., Mantell, C., Varela, R., Martínez de la Ossa-Fernández, E. J., & Macías, F. A. (2019). Selective fractionation and isolation of allelopathic compounds from *Helianthus annuus* L. leaves by means of high-pressure techniques. *Journal of Supercritical Fluids*, 143, 32-41.
- Fuzer, A. M., Filho, J. C., Becceneri, A. B., Dos Santos, D. A., da Silva, M. F., Vieira, P. C., Fernandes, J. B., Selistre-de-Araujo, H. S., Cazal, C. M., & Cominetti, M. R. (2013). Effects of limonoid cedrelone on MDA-MB-231 breast tumor cells *in vitro*. *Anti-cancer Agents in Medicinal Chemistry*, 13(10), 1645-1653.
- Geng, Y. D., Zhang, C., Lei, J. L., Yu, P., Xia, Y. Z., Zhang, H., Yang, L., & Kong, L. Y. (2017). Walsuronoid B induces mitochondrial and lysosomal dysfunction leading to apoptotic rather than autophagic cell death via ROS/p53 signaling pathways in liver cancer. *Biochemical Pharmacology*, 142, 71-86.
- Ghosh, R. B. (1968). Studies on the morphology of somatic chromosomes in *Walsura piscidia* Roxb. *Caryologia*, 21, 111-114.
- Ghoula, M., Le Marec, A., Magnan, C., Le Stunff, H., & Taboureau, O. (2020). Identification of the interactions interference between the PH and START domain of CERT by limonoid and HPA inhibitors. *Frontiers in Molecular Biosciences*, 7, 603983.
- Giongo, A. M., Vendramim, J. D., Freitas, S. D., & Silva, M. F. (2016). Toxicity of secondary metabolites from Meliaceae against *Spodoptera frugiperda* (J. E. Smith) (Lepidoptera: Noctuidae). *Neotropical Entomology*, 45(6), 725-733.
- Govindachari, T. R., Narasimhan, N. S., Suresh, G., Partho, P. D., Gopalakrishnan, G., & Krishna Kumari, G. N. (1995). Structure-related insect antifeedant and growth regulating activities of some limonoids. *Journal of Chemical Ecology*, 21(10), 1585-1600.
- Govindachari, T. R., Suresh, G., Gopalakrishnan, G., Masilamani, S., & Banumathi, B. (2000). Antifungal activity of some tetranortriterpenoids. *Fitoterapia*, 71, 317-320.
- Haque, A. K. M. K., Khan, S. A., Uddin, S. N., & Shetu, S. S. (2018). An annotated checklist of the angiospermic flora of Rajkandi reserve forest of Moulvibazar, Bangladesh. *Bangladesh Journal of Plant Taxonomy*, 25, 187-207.
- Heightman, T. D., Berdini, V., Braithwaite, H., Buck, I. M., Cassidy, M., Castro, J., Courtin, A., Day, J., East, C., Fazal, L., Graham, B., Griffiths-Jones, C. M., Lyons, J. F., Martins, V., Muench, S., Munck, J. M., Norton, D., O'Reilly, M., Palmer, N., Pathuri, P., Reader, M., Rees, D. C., Rich, S. J., Richardson, C., Saini, H., Thompson, N. T., Wallis, N. G., Walton, H., Wilsher, N. E., Woolford, A. J., Cooke, M., Cousin, D., Onions, S., Shannon, J., Watts, J., & Murray, C. W. (2018). Fragment-Based Discovery of a Potent, Orally Bioavailable Inhibitor That Modulates the Phosphorylation and Catalytic Activity of ERK1/2. *Journal of Medicinal Chemistry*, 61, 4978-4992.
- Hou, L., Tang, G. H., Zhang, Y., Hao, X. J., Zhao, Q., & He, H-P. (2013). A new carotane sesquiterpene from *Walsura robusta*. *Chinese Journal of Natural Medicines*, 11, 84-86.
- Hussein, N. S., Ahmed, A. A., Spaller, M., & Mabry, T. J. (1995). Terpenoids from *Conza linifolia*. *Asian Journal of Chemistry*, 7, 433-435.
- Jares, E. A., & Pomillo, A. B. (1989). Isolation of Sesquiterpenes from *Senecio crassiflorus* Combined Dry Column and High-Performance Liquid Chromatography. *Journal of Separation Science*, 12, 565-568.
- Ji, K. L., Zhang, P., Hu, H. B., Hua, S., Liao, S. G., & Xu, Y. K. (2014). Limonoids from the Leaves and Twigs of *Walsura yunnanensis*. *Journal of Natural Products*, 77, 1764-1769.
- Ji, K. L., Zhang, P., Li, X. N., Guo, J., Hu, H. B., Xiao, C. F., Xie, X. Q., & Xu, Y. K. (2015). Cytotoxic limonoids from *Trichilia americana* leaves. *Phytochemistry*, 118, 61-67.
- Jiang, H. U., Jia-Xun, L. I., Qiang, L. I., Tian-Feng, P., Xiao-Dong, S., Tao, L., Jian, C., Nian-Hua, J., Wei-Xing, Y., & Wei, W. (2020). [Terpenoids from *Toona sinensis*]. *Zhongguo Zhong Yao Za Zhi*, 45, 4411-4415.
- Jin, Z., Ma, J., Zhu, G., & Zhang, H. (2019). Discovery of Novel Anticryptosporidial Activities From Natural Products by *in vitro* High-Throughput Phenotypic Screening. *Frontiers in Microbiology*, 10, 1999.
- Kumagai, K., Hanada, K. (2019). Structure, functions and regulation of CERT, a lipid-transfer protein for the delivery of ceramide at the ER-Golgi membrane contact sites. *FEBS Letters*, 593, 2366-2377.
- Li, A., Yan, Q., Zhao, X., Zhong, J., Yang, H., Feng, Z., Du, Y., Wang, Y., Wang, Z., Wang, H., Zhou, Y., Liu, S., & Nie, Y. (2016). Decreased expression of PBLD correlates with poor prognosis and functions as a tumor suppressor in human hepatocellular carcinoma. *Oncotarget*, 7, 524-537.
- Liang, L., & Zhang, Z. (2016). Gambogic Acid Inhibits Malignant Melanoma Cell Proliferation Through Mitochondrial p66shc/ROS-p53/Bax-Mediated Apoptosis. *Cellular Physiology and Biochemistry*, 38, 1618-1630.
- Luo, X.D., Wu, D. G., Cai, X. H., & Kennelly, E. J. (2006). New antioxidant phenolic glycosides from *Walsura yunnanensis*. *Chemistry and Biodiversity*, 3, 224-230.
- Misra, L. N., Jakupovic, J., Bohlmann, F., & Schemda-Hirschmann, G. (1985). Isodaucane derivatives, norsesquiterpenes and clerodanes from *chromolaena laevigata*. *Tetrahedron*, 41, 5353-5356.
- Moriyasu, M., Takeuchi, S., Ichimaru, M., Nakatani, N., Nishiyama, Y., Kato, A., Mathenge, S. G., Juma, F. D., & ChaloMutiso, P. B. (2012). Pyrenes and pyrendiones from *Uvaria lucida*. *Journal of Natural Medicines*, 66, 453-458.
- Palau, V. E., Chakraborty, K., Wann, D., Lightner, J., Hilton, K., Brannon, M., Stone, W., & Krishnan, K. (2018). γ -Tocotrienol induces apoptosis in pancreatic cancer cells by upregulation of ceramide synthesis and modulation of sphingolipid transport. *BMC Cancer*, 18, 564.
- Pasha, M. K., Uddi, S. B. (2019). Minor edible fruits of Bangladesh. *Bangladesh Journal of Plant Taxonomy*, 26, 299-313.
- Qin, G. F., Tang, X. L., Sun, Y. T., Luo, X. C., Zhang, J., van Otwegen, L., Sung, P. J., Li, P. L., & Li, G. Q. (2018). Terpenoids from the Soft Coral *Sinularia* sp. Collected in Yongxing Island. *Marine Drugs*, 16, 127.
- Sakkat, P. (2007). Estimation of number and density, and random distribution testing of important plant species in Ban Pong Forest, Sansai District, Chiang Mai Province, Thailand using T-Square sampling. *Maejo International Journal of Science and Technology*, 1, 64-72.
- Shen, H., Liu, L., Yang, Y., Xun, W., Wei, K., & Zeng, G. (2017). Betulinic acid inhibits cell proliferation in human oral squamous cell carcinoma via modulating ROS-regulated p53 signaling. *Oncology Research*, 25, 1141-1152.
- Tan, Q. G., & Luo, X. D. (2011). Meliaceae limonoids: chemistry and biological activities. *Chemical Reviews*, 111, 7437-7522.
- Tundis, R., Loizzo, M. R., & Menichini, F. (2014). An overview on chemical aspects and potential health benefits of limonoids and their derivatives. *Critical Review in Food Science and Nutrition*, 54, 225-250.
- Voravuthikunchai, S., Lortheeranuwat, A., Jeeju, W., Sriirak, T., Phongpaichit, S., & Supawita, T. (2004). Effective medicinal plants against enterohaemorrhagic *Escherichia coli* O157:H7. *Journal of*

- Ethnopharmacology*, 94, 49-54.
- Voravuthikunchai, S. P., Kanchanapoom, T., Sawangjaroen, N., & Hutadilok-Towatana, N. (2010). Antioxidant, antibacterial and antiangiogenic activities of *Walsura robusta* Roxb. *Natural Products Research*, 24, 813-824.
- Voravuthikunchai, S. P., Kitpip, L. (2005a). Activity of medicinal plant extracts against hospital isolates of methicillin-resistant *Staphylococcus aureus*. *Clinical Microbiology and Infection*, 11, 510-551.
- Voravuthikunchai, S. P., & Kitpip, L. (2005b). Antibacterial activity of crude extracts of Thai medicinal plants against clinical isolates of methicillin-resistant *Staphylococcus aureus*. *Journal of Science and Technology*, 27, 525-534.
- Voravuthikunchai, S. P., & Limsuwan, S. (2006). Medicinal plant extracts as anti-*Escherichia coli* O157:H7 agents and their effects on bacterial cell aggregation. *Journal of Food Protection*, 69, 2336-2341.
- Wang, G. C., Yu, J. H., Shen, Y., Leng, Y., Zhang, H., & Yue, J. M. (2016). Limonoids and Triterpenoids as 11 β -HSD1 Inhibitors from *Walsura robusta*. *Journal of Natural Products*, 79, 899-906.
- Wang, N., Fan, Y., Yuan, C. M., Song, J., Yao, Y., Liu, W., Gajendran, B., Zacksenhaus, E., Li, Y., Liu, J., Hao, X. J., & Ben-David, Y. (2019). Selective ERK1/2 agonists isolated from *Melia azedarach* with potent anti-leukemic activity. *BMC Cancer*, 19, 764.
- Wu, J., Niu, Q., Yuan, J., Xu, X., & Cao, L. (2019). Novel compound cedrelone inhibits hepatocellular carcinoma progression via PBLD and Ras/Rap1. *Experimental and Therapeutic Medicine*, 18, 4209-4220.
- Yin, S., Wang, X. N., Fan, C. Q., Liao, S. G., and Yue, J. M. (2007). The first limonoid peroxide in the Meliaceae family: walsuronoid A from *Walsura robusta*. *Organic Letters*, 9, 2353-2356.
- Zhang, Y., An, F. L., Huang, S. S., Yang, L., Gu, Y. C., Luo, J., & Kong, L. Y. (2017). Diverse triterpenoids from the fruits of *Walsura robusta* and their reversal of multidrug resistance phenotype in human breast cancer cells. *Phytochemistry*, 136, 108-118.
- Zhang, Y., & Xu, H. (2017). Recent progress in the chemistry and biology of limonoids. *RSC Advances*, 7, 35191-35220.