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# Qualitative analysis of some bioactive components of methanolic leaf extract of *M. citrifolia* (Noni)

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## ABSTRACT

Medicinal plants offer endless opportunities for new drugs discovery due to their supremacy for the possession of phytochemical compounds known for diverse antimicrobial activities. The world ever increasing demand for therapeutic drugs from natural products with particular interest in edible plants for safety purposes is now catching researchers' attention. This study therefore aimed at determining the presence of some bioactive phytochemical components of methanolic leaf extract of *M. citrifolia* L. Qualitative screening of leaf extract has confirmed the existence of Tannins, steroids, saponins, flavonoids and alkaloids in the mixture. And these bioactive compounds correspond to phytochemicals with antimicrobial, nematocidal, pesticidal, antioxidant, ant-inflammatory, cytotoxic, anti-allergy, and anti-carcinogenic properties (bioactive compounds) earlier documented by previous researchers.

**KEYWORDS:** Bioactive compounds, *M. citrifolia*, Methanol, Phytochemicals

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## INTRODUCTION

The use of synthetic chemical for plant protection is highly effective but the over dependency on these synthetic agrochemicals is predisposing consumers to mycotoxins and build-up of chemical residue in the environment [1]. Medicinal plants are sources of phytochemicals with antimicrobial effect which may serve as safe alternative biopesticide with low toxicity to human and the environment [2]. These phytochemical compounds are plants' defensive/health enhancing compounds, for self-protection against biotic and abiotic stress [3] and [4]. They are regarded as alternative bases for broad spectrum natural biopesticides for the management of plant pathogens, owing to their possession of complex mechanisms of action against pathogenic organisms [5].

According to [6] the world is currently shifting away from the synthetic pesticides to thousands of alternative naturally occurring bioactive compounds, because they are environmentally friendly, broadly effective and safe for both terrestrial and aquatic animals due to low level of acute toxicity. Botanical extracts are normally mixtures of composite bioactive compounds which need to be identified [7].

Qualitative phytochemical screening assays is a vital tool used in bioactive compound analyses, it is inexpensive, quick, and simple, procedure that provides researchers with swift idea of the phytochemical components in a plant extract. This technique uses known and specific tests to prove the presence of a certain compound [8] and [6]. The major steps to identification and utilization of plant base biologically active compound are extraction, screening, and characterization of the compounds, then followed by toxicological assessment. Therefore, this study aimed at extraction and determination of bioactive components; tannins, steroids, saponins, flavonoids, and alkaloids in *M. citrifolia* leaf powder using phytochemical screening assay (chemical tests).

## MATERIALS AND METHODS

### Collection and Preparation of Plant Materials

Fresh *M. citrifolia* leaves were collected around the Universiti Putra Malaysia main library. Collected leaves were identified/confirmed as *M. citrifolia* L. by a botanist at the Herbarium of Biodiversity Unit, Institute of Bioscience (IBS), Universiti Putra Malaysia, (Reference number UPM/IBS/UB/H90/17 and

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Voucher number SK 3255/17) (Appendix A). Thereafter leaves were brought to the Biocontrol Laboratory, Department of Plant Protection, Faculty of Agriculture, Universiti Putra Malaysia (UPM), washed under running tap water to get rid of dust and debris and rinsed in sterile distilled water. Leaves were first allowed to stay for 6 hrs under the laminar air flow to dry up the wet leaves surface. And finally dried at 40- 45°C in a mechanical convection oven (Memmert, Germany). An electrical grinder Retsch SK100 standard Gußeisen, 2002 was used to grind the dried leaves into the powdered form for use [5].

### Test for Tannins

To determine the presence of Tannins, Braemer's test was performed. Following the method described by [6], where 2 g of the powdered *M. citrifolia* leaf was dissolved in 10 ml of methanol, then macerated and filtered by means of cotton wool and funnel. Thereafter, 2 ml of the filtrate was added to 2 ml of 10 % alcoholic ferric chloride (1:1). Formation of greenish grey coloration of the mixture indicates the presence of tannins.

### Test for Steroids

To test for the presence of steroids, Lieberman Burchardt test was used. For the test, 2 g of the leaf powder was added to 20 ml of methanol in a 150 ml conical flask and covered for 30 min, mixture was filtered using cotton wool and funnel. Filtrate was poured into a 50 ml beaker and placed on a water bath until filtrate was completely evaporated. 6 ml of chloroform was added to the evaporated extract and mixed thoroughly. Then 2 ml of the chloroform mixture was transferred into a test tube where few drops of acetic anhydride was added, followed by addition of few drops of H<sub>2</sub>SO<sub>4</sub>, which was added slowly to the wall of the test tube. Formation of dark green colour designated the presence of steroids [9]. Plants steroids are derivatives of cyclization of the triterpene squalene [10] and [11].

### Test for Saponins

To determine the presence of saponin in the phytochemical components of *M. citrifolia* leaf. 70 ml of sterile distilled water was placed in a beaker containing 3 g of plant powdered leaf, mixed then boiled for 2 min. Mixture was filtered into a new test tube using cotton wool and funnel to produce an aqueous extract. 2 ml of the aqueous extract was discharged into a graduated test tube, and vigorously agitated. The formation of 1cm foam that persists for 3 minutes designated the presence of saponins [12].

### Test for Flavonoid

Ammonium test was employed to test for the presence of Flavonoid in leaf extract of *M. citrifolia*, following the method as described by [13] and [14]. To achieve this, 0.2 g of *M. citrifolia* leaf powder was added to 10 ml of ethyl acetate in a 100 ml conical flask. Mixture was then heated for 5 min in a water bath, allowed to cool and filtered. 4 ml of the filtrate was discharged into a test tube where 1 ml of diluted ammonia

solution was added to the mixture, agitated and kept at room temperature for a few seconds then observed. Appearance of layer of yellow coloration at the bottom of the test tube indicates the presence of flavonoid

### Test for Alkaloids

Dragendroff reagent test: For this test, 0.2 g of *M. citrifolia* leaf powder was added to 20 ml of diluted H<sub>2</sub>SO<sub>4</sub> in methanol in a conical flask. Mixture was boiled for 5 minutes in a water bath, cool and filtered. Three drops of dragendroff reagent were added to the filtrate. Formation of creamy, orange solution indicates the presence of alkaloids [15].

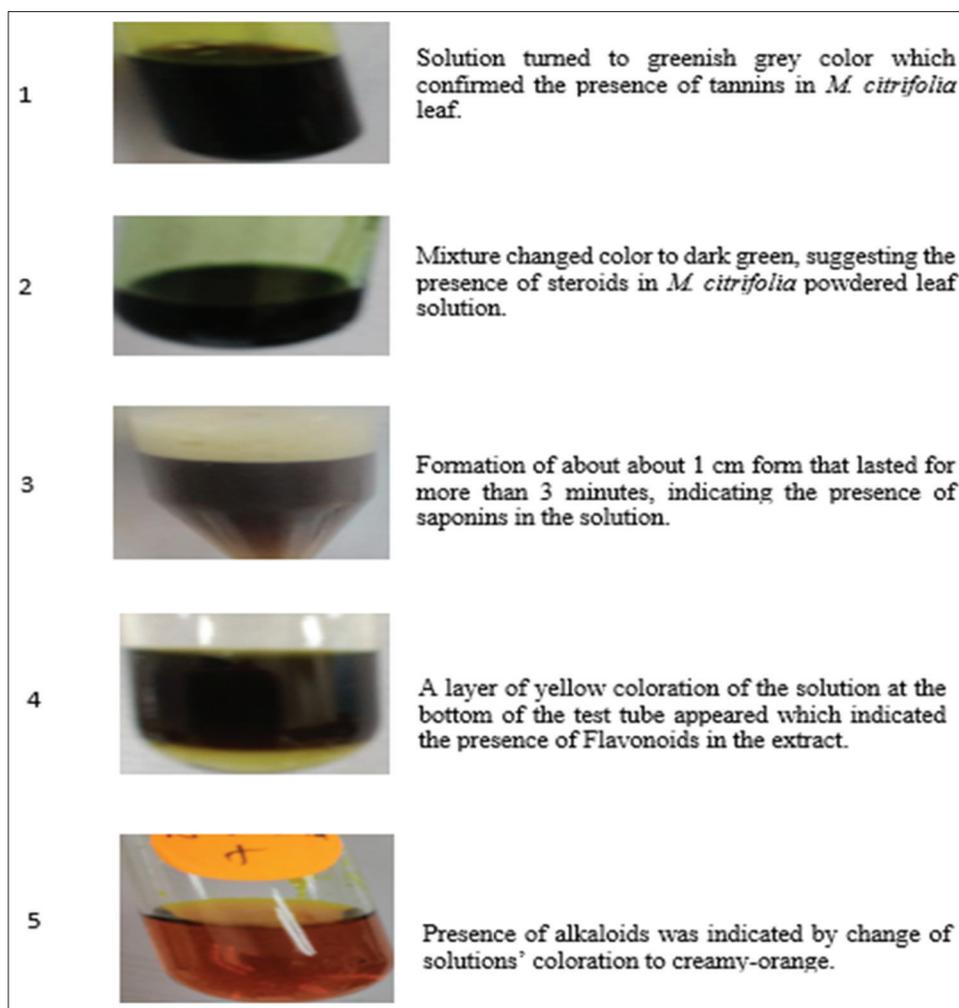
## RESULTS AND DISCUSSION

The qualitative analysis of phytochemical components of *M. citrifolia* leaf extract using the conventional phytochemical screening assay (chemical tests) detected the presence of tannins, steroids, saponins, flavonoids, and alkaloids (Table 1 and Figure 1) which is in agreement with the findings of [16] and [17]. During the Braemer's test, the *M. citrifolia* leaf extract turned greenish grey which was an indication for the presence of Tannins in the solution according to the reports of [18] and [8]. Tannins belong to the class polyphenol compounds with an astringent property, soluble in acetone, alcohol, and water. Similarly, Lieberman test (Burchardt test) of the leaf extract solution turned dark green in colour (Figure 2) which was an indication for the presence of steroids. This finding is in line with the report of [8]. The Frothing test also showed the formation of 1 cm foam height above the mixture which persisted for more than 3 min. According to [6], the appearance of up to 1 cm foam height above mixture that lasts for up to 2-3 minutes is an indication for the presence of saponins (Figure 3). Saponins are naturally produced by many plants for defense against pest and pathogens, they are easily converted to drugs, cosmetics and taste modifiers and are therefore considered economically viable compounds [19] and [20]. Result for the Ammonium test (test for flavonoids) showed layer of yellow coloration at the bottom of the test tube, which is an indication for the presence of flavonoid in the leaf extract, according to [21] and [14] (Figure 4). Flavonoids is another member of the compounds class polyphenols which are known for their Pesticidal, antimicrobial, antioxidant, chemotherapy activities, and their mechanism of action against microorganisms includes; complex activities with the cell wall, cell lysis, membrane disruption, inactivation of enzymes and death [22]. Findings of the Dragendroff test (test

**Table 1: Phytochemical constituents of *M. Citrifolia* leaf powder**

| Phytochemical compound | Chemical test    | Indicator                                  | Result |
|------------------------|------------------|--|--------|
| Tannins                | Braemer's test   | Greenish grey                              | +      |
| Steroids               | Burchardt test   | Dark green colour                          | +      |
| Saponins               | Froth test       | 1 cm foam that lasted for 3 minutes,       | +      |
| Flavonoids             | Ammonium test    | A layer of yellow coloration at the bottom | +      |
| Alkaloids              | Dragendroff test | Creamy orange coloration                   | +      |

Key: (+) indicates the presence of a phytochemical compound



**Figure 1:** Pictorial summary of phytochemical screening assay tests for tannins, steroids, saponins, flavonoids and alkaloids components of *M. citrifolia* leaf extract.

for Alkaloids) shows formation of layer of creamy-orange solution (Figure 5). This result is in agreement with the report of [15], that by this test, the formation of creamy-orange coloration of the test solutions suggests the presence of alkaloids in the test solution. Alkaloids have a toxic effect on microbials, in human medication and or as biopesticides [22].

## CONCLUSION

The methanolic leaf extract of *M. citrifolia* has the following significant/potent bioactive compounds; tannins, steroids, saponins, flavonoids and alkaloids. These compounds also correspond to those with antimicrobial, antioxidant, pesticidal, anticancer, anti-inflammatory, Hepatoprotective activities earlier reported by [23] and [7]. Therefore, *M. citrifolia* leaf extract can be utilized as a new source of drugs and pesticides for both agrochemical industries and pharmaceutical industries.

## REFERENCES

- Bokhari, N. A., Siddiqui, I., Siddique, K. P. I., Rizwana, H., and Soliman, D. A. W. Management of anthracnose of banana by UV irradiation. *Journal of Animal and Plant Sciences*. 2013; 23:1211-12114.
- Madhumitha, G., Rajakumar, G., Roopan, S. M., Rahuman, A. A., Priya, K. M., Saral, A. M., and Kamaraj, C. Acaricidal, insecticidal, and larvicidal efficacy of fruit peel aqueous extract of *Annona squamosa* and its compounds against blood-feeding parasites. *Parasitology Research*. 2012;111:(5),2189-2199.
- Singh, S. Enhancing phytochemical levels, enzymatic and antioxidant activity of spinach leaves by chitosan treatment and an insight into the metabolic pathway using DART-MS technique. *Food Chemistry*. 2016;199:176-184.
- Cos, P., Vlietinck, A. J., Berghe, D. V., and Maes, L. Anti-infective potential of natural products: how to develop a stronger in vitro 'proof-of-concept'. *Journal of Ethnopharmacology*. 2006; 106 (3):290-302.
- Idris, F. M., Ibrahim, A. M., and Forsido, S. F. Essential oils to control *Colletotrichum musae* in vitro and in vivo on banana fruits. *American-Eurasian Journal of Agricultural and Environmental Science*. 2015;15(3):291-302.
- Sasidharan, S., Chen, Y., Saravanan, D., Sundram, K. M., and Latha, L. Y. Extraction, isolation and characterization of bioactive compounds from plants' extracts. *African Journal of Traditional, Complementary and Alternative Medicines*. 2011; 8(1):1-10.
- Kumar, S., Samydrurai, R., and Nagarajan, N. Gas chromatography and mass spectrometry analysis of bioactive constituents of *Adiantum Capillus-veneris* L. *International Journal of Pharmacy and Pharmaceutical Sciences*. 2014;6: 60-63.
- Compean, K. L., and Ynalvez, R. A. Antimicrobial activity of plant

- Secondary metabolites: A Review. Research Journal of Medicinal Plants. 2014;8(5):204-13.
9. Kumar, G. S., Jayaveera, K. N., Kumar, C. K., Sanjay, U. P., Swamy, B. M., and Kumar, D. V. Antimicrobial effects of Indian medicinal plants against acne-inducing bacteria. Tropical Journal of Pharmaceutical Research. 2007; 6(2): 717-723.
  10. Burčová, Z., Kreps, F., Greifová, M., Jablonský, M., Ház, A., Schmidt, Š. and Šurina, I., Antibacterial and antifungal activity of phytosterols and methyl dehydroabietate of Norway spruce bark extracts. Journal of Biotechnology. 2018; 282:18-24.
  11. Mubiu, J.K., Ndwigah, S.N., Abuga, K.O. and Ongarora, D.S. Antimicrobial activity of extracts and phytosterols from the root bark of Lonchocarpus eriocalyx. East and Central African Journal of Pharmaceutical Sciences. 2017;20(1-3):13-16.
  12. Parekh, J., and Chanda, S. Antibacterial and phytochemical studies on twelve species of Indian medicinal plants. African Journal of Biomedical Research. 2007;10(2): 175-181.
  13. Onwukaeme DN, Ikuogbvweha TB, Asonye CC. Evaluation of phytochemical constituents, antibacterial activities and effect of exudate of *Pycnanthus Angolensis* weld warb (Myristicaceae) on corneal ulcers in rabbits. Tropical Journal of Pharmaceutical Research. 2007;6(2):725-30.
  14. Harborne, J. B. Twenty-five years of chemical ecology. Natural product reports. 2001; 18(4): 361-379.
  15. Gul, R., Jan, S. U., Faridullah, S., Sherani, S., and Jahan, N. Preliminary Phytochemical Screening, Quantitative Analysis of Alkaloids, and Antioxidant Activity of Crude Plant Extracts from *Ephedra intermedia* Indigenous to Balochistan. The Scientific World Journal. 2017; 2017(Special Issue).
  16. Yahia EM, editor. Postharvest biology and technology of tropical and subtropical fruits: Mangosteen to white sapote. Elsevier; 2011 Jun 30.
  17. Wang, M. Y., and Su, C. Cancer preventive effect of *Morinda citrifolia* (Noni). Annals of the New York Academy of Sciences. 2001; 952(1): 161-168.
  18. Nair, R. and Chanda, S. Antibacterial activities of some medicinal plants of the western region of India. Turkish Journal of Biology. 2007;31(4):231-236.
  19. Somani, S. J., Modi, K. P., Majumdar, A. S., and Sadarani, B. N. Phytochemicals and their potential usefulness in inflammatory bowel disease. Phytotherapy Research, 2015; 29(3): 339-35.
  20. Sharma G, Prakash D, Gupta C, Prakash D, Sharma G. Phytochemicals of nutraceutical importance: do they defend against diseases. Phytochemicals of Nutraceutical Importance. 2014;28;1.
  21. Morrissey, J. P., and Osbourn, A. E. Fungal resistance to plant antibiotics as a mechanism of pathogenesis. Microbiology and Molecular Biology Reviews. 1999; 63(3):708-724.
  22. Okwu D. E. Phytochemicals, Vitamins and Mineral contents of two Nigeria Medicinal plants. International Journal of Molecular Medicine and Advance Sciences. 2005;1(4): 375-381.
  23. Biresaw G, Mittal KL. Surfactants in Tribology, 2 Volume Set. CRC Press; 2011 Jun 17.