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## **Research Article**

## Natural plant colorants widely used in Vietnam traditional food culture

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#### Abstract

The artificial colorants have gradually been being replaced by natural pigments which are becoming increasingly important in Vietnam and other parts of the world due to the potential noxiousness of man-made food dyes to human health. This research covers colorant plants and sources used commonly in the food culture of ethnic communities in Vietnam with current trend toward natural pigments and coloring foods. As a result, we reported 49 species which can be used as natural food pigments and of these, 7 colorant plants used mostly in Vietnam traditional food culture were detailed.

Keywords: Natural pigments, food coloring, colorant plant, Vietnam

#### Introduction

In recent years, consumers are averting foods containing artificial colorants by reason of associations with possible harmful effects to human health. As a result, food industries have been substituting them by natural pigments and there is a risen world-wide trend towards usage of natural colors in food, pharmaceutical and personal care industries. Moreover, the food industry and nutrition scientists have dedicated numerous studies to discovering natural alternatives for the evolution of natural food colorants. The results confirmed that using natural food colorants brings many benefits. Natural colorants for food made from renewable sources do not cause health hazards. A number of synthetic dyes such as tartrazine and carmoisine are commonly added to foods, but their use has been questioned because of their suspected toxic properties such as damaging liver function and causing oxidative stress. Hence, interest has increased considerably in searching naturally occurring pigments.

Many natural dyes are used as both food coloring and substances that protect health and well being by precluding or even treating diseases. Moreover, the use of natural pigments offers no toxic properties (Kumar and Sinha, 2004). However, endorsement of such new pigment sources as "novel" food colorants mostly demands stringent food safety assessments, even if their long-term use in the respective country of origin has been previously described to be harmless or even to be associated with health-protective properties. Because of the suspected toxic properties of artificial dyes, all countries have promulgated rigorous regulations about the permitted colors to be used as food additives (Kapoor, 2006). This review aims to cover and display knowledge on colorant plants and sources used commonly in the food culture of ethnic communities in Vietnam

#### Materials and Methodology

The study was conducted to collect real-life interviews and literature references and field survey in the ethnic tribes and communities in almost provinces of northern Vietnam. Field investigations and sample collecting were performed in random chosen regions from 2013 to 2017. We focus on high-density ethnic minorities areas: Tay-Nung people in the Cao Bang and Lao Cai Province; Tai people in the Son La, Lang Son and Tuyen Quang Province; H' Mong people in the Yen Bai, Lai Chau and Dien Bien Province; Muong people in Hoa Binh; Trai-San Diu people in Thai Nguyen; San Chay people in Bac Kan. A data-base was collected from participant observation and semi-structured interviews with questionnaires regarding the local knowledge on utilization of colorant plants in traditional food culture. The data were processed by basic mathematic method in microsoft excel.

## **Results and Discussions**

# Colorant plants could be used in traditional food culture in Vietnam

Our survey results show that there are forty nine species belonging to 30 families giving pigments for food coloring described in Table 1. Indigenous tribes use botanies for dyeing in traditional food and national festivals. Among forty nine, twelve species are consumed for red color, thirteen species for yellow, eleven species for black and other colors. *Peristrophe bivalvis* has diverse varieties for different colors.

# Colorant plants used mostly in Vietnam traditional food culture

In the present study, in comparison with previous study in Vietnam (Dam *et al.*, 2016), we found and added nine

SI. No	Family name	Latin name	Vietnamese name	Part containing	Colors	Food
1	Acanthaceae	Dicliptera chinensis (L.) Juss	Diễn	Leaves	Red	Rice
		<i>Peristrophe bivalvis</i> (L.) Merr. (several cultivars)	Cẩm	Leaves	Red, Purple, yellow	Rice
		Strobilanthes cusia (Nees) Kuntze	Chàm mèo	Leaves	Black	Rice
2	Altingiaceae	Liquidambar formosana Hance	Sau sau	Leaves	Black	Rice
3	Amaranthaceae	Amaranthus caudatus	Dền tía	Leaves	Red	Drink
		Iresineherbstii Hook. F. ex Lindl.	Nhung hoa	Leaves	Red	Rice
	Anacardiaceae	Rhus chinensis Mill.	Muối	Stem (bark)	Black	Rice
i	Asteraceae	Artemisia indica Willd	Ngải cứu	leaves	green	Cake
		Gnaphalium affine D. Don	Rau khúc	Whole plant	Green	Cake
5	Basellaceae	Basella rubra L.	Mồng tơi	Fruit	Purple	Rice
7	Bignoniaceae	Oroxylum indicum (L.) Kurz	Núc nác	Stem-ash	Black	Cake
3	Bixaceae	Bixa orellana L.	Điều nhuộm	Seed aril	Orange, dark- yellow	Rice
9	Buddlejaceae	Buddleja macrostachya Wall. Ex Benth.	Búp lệ chùm to	Flower	Yellow	Rice
	,	Buddleja officinalis Maxim.	Mật mông hoa	Flower	Yellow	Rice
		Buddleja paniculata Wall.	Búp lệ chùm tụ tán	Flower	Yellow	Rice
0	Burseraceae	Canarium tramdenum Dai & Yakovl	Trám đen	Fruit	Black	Rice
1	Caesalpiniaceae	Caesalpinia sappan L.	Tô mộc	Wood	Red	Rice
	•	Saraca dives Pierre	Vàng anh	flower	yellow	Rice
2	Cucurbitaceae	Momordica cochinchinensis (Lour.) Spreng	Gấc	Seed aril	Red	Rice
		Luffa cylindria (L.) M.J.Roem	Mướp	Leaves	green	Rice
3	Dracaenaceae	Dracaena cochinchinensis (Lour.) S.C.Chen	Bồng bồng	Stem	Red	Drink
4	Elaeagnaceae	Elaeagnus latifolia L.	Nhót	Leaves	Black	Rice
15	Fabaceae	Clitoria ternatea L.	Đậu biếc	Flower	Blue	Rice
		Dalbergia volubilis Roxb.	Trắc leo	Ash	Black	Rice, cake
		Milletia sp	Cát sâm	Stem	Red	Drink
		Vigna cylindrical (L.) Skeels	Đậu đen	Seed	Black	Rice
		Spatholobus suberectus Dunn	Kê huyết đằng	Stem, leaves	Red	Drink rice
6	Iridaceae	Eleutherin bulbosa (Mill.) Urban	Sâm đại hành	Corn	Red	Drink
7	Magoliaceae	Micheliame diocris Dandy	Giổi xanh	Leaves	Green	Rice
8	Malvaceae	<i>Hibiscus sabdariffa</i> L.	Bụp giấm	Fruit	Red	Drink
9	Marantaceae	Phrynium imbricatum Gagnep	Lá dong	Leaves	Green	Rice cake
20	Menispermaceae	Fibraureatin ctoria Lour.	Hoàng đằng	Stem	Yellow	Wine
21	Moraceae	Morus alba L.	Dâu tằm	Fruit	purple	Drink
2	Myrtaceae	Rhodomyrtus tomentosa (Ait.) Hassk	Sim	Fruit	Purple	Drink
23	Pandanaceae	Pandanus amarylliforlius	Dứa thơm	Leaves	Green	Rice, cake
24	Poaceae	Oryza sativa L. var. glutinosa Blanco	Lúa nếp	Wooden ask seed	Black, mordant	Rice, cake
		<i>Thysanolaena maxima</i> (Roxb). Kuntze	Chít	Leaves	yellow	Rice, cake
25	Polygonaceae	Reynoutria japonica Hout.	Cốt khí	Tuber	Yellow	Stick: rice
26	Verbenaceae	<i>Gmelina arborea</i> Roxb. Ex Sm.	Lõi thọ	Flower	Yellow	Rice
27	Rubiaceae	Luculia gratissima (Wall.) Sweet	Gạc nai	Wood	Yellow	Drink
		Gardenia augusta (L.) Merr. Paederia Lanuginosa Wall.	Dành dành Mơ tam thể	Fruit, flower Leaves	Yellow Black	Rice Rice,
0	Smilassass	Smilar alabra	Khúc khắc	Stam	Pad	cake
28	Smilacaceae	Smilax glabra Rochmaria rivag (L.) Coudich		Stem	Red	Drink
29 30	Urticaceae Zingiberaceae	Boehmeria nivea (L.) Gaudich Alpinia gagnepainii (Gagnep.) K. Schum	Lá gai Riềng	Leaves Leaves	Black Green	Cake Rice
		Alpinia officinarum Hance	Riềng	Leaves	Green	Rice
		Curcuma longa L.	Nghệ vàng	Rhizomes	Yellow	Rice
		<i>Curcuma zedoaria</i> (Christm) Rosc.	Nghệ đen	Rhizome	Yellow	Rice
		Zingiber officinale Roscoe	Gừng		//	Rice

Table 1. Checklist of natural colorant plants for food in Vietnam

plants: Strobilanthes cusia (Family: Acanthaceae) Iresineherbstii Hook. F. ex Lindl. (Family: Amaranthaceae), Saraca dives Pierre (Family: Caesalpiniaceae), Luffa cylindria (L.) M.J.Roem (Family: Cucurbitaceae), Elaeagnus latifolia L. (Family: Elaeagnaceae) Spatholobus suberectus (Family: Fabaceae) Gardenia augusta (L.) Merr. (Family: Rubiaceae), *Smilax glabra* (Family: Smilacaceae) *Boehmeria nivea* (L.) Gaudich (Family: Urticaceae).

Also according to survey results, we recorded that among possible dye plants, there are seven colorant plants (*Peristrophe bivalvis* L., *Liquidambar formosana* Hance, *Basella rubra* L., *Gardenia jasminoides*, *Momordica cochinchinensis* Spreng, *Turmeric* (*Curcuma longa*), *Pandanus amarylliforlius*), which are consumed commonly in Northern Vietnam (Fig. 1).

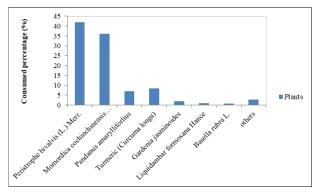


Fig. 1. Percentage of colorant plants consumed in food culture

## Peristrophe bivalvis (L.) Merr.

Peristrophe bivalvis (L.) Merr. is known and used widely in food processing as ingredient to look more attractively. Each ethnic tribe and region has a kind of specific traditional food processed with P. bivalvis, which make characteristic culture values. Based on the various colors and morphological characteristics of extracts, P. bivalvis possesses four colorant types: purples, purple-megenta, yellow, and red (Chi, 1999). P. bivalvis has many advantage features applied in biology and pharmacy for blood problems as assisted therapy of high blood pressure, high levels of lipids; inhibiting development of fungi and bacteria (Yang et al., 2002; Cheng et al., 2004; Verma et al., 2008; Wiart et al., 2004). Other works also showed prominent bacterial inhibitory capacity of ethanolic extracts (Tanavade et al., 2012a, b). Peristrophine is important constituent isolated from the colour water-extract (CWE) of its leaves. Trinh et al. (2014) also investigated the acute oral toxicity of the CWE in mice and recorded LD<sub>50</sub> of the CWE is 9100 ± 290 mg/kg of body weight. In addition, in vitro cytotoxic test revealed peristrophine to have HepG2 carcinoma cells growth inhibitory capacity at IC<sub>50</sub> of 3.90  $\mu$ g/ mL. The antioxidant effect of color extract from this plant also was determined by Nguyen et al. (2016).

### Momordica cochinchinensis Spreng

Gac (Momordica cochinchinensis Spreng.), a tropical fruit, classified in the Cucurbitaceae is used both as a food and a traditional medicine in many nations as China, Viettnam and Thailand (Hoang, 2007) and East and Southeast Asia countries (Iwamoto *et al.*, 1985). The fruit is with a thick yellow mesocarp and red soft and sticky aril (Tran *et al.*, 2008). The red aril surrounding the seeds of mature fruit is used to make a dish named as "Xoi Gac" in Vietnamese, a type of famously glutinous rice eaten at traditional holiday and anniversary days. In addition, the premature green fruit is consumed as a vegetable in Thailand (Kubola and Siriamornpun, 2011) and India (Joseph and Bharathi, 2008). The red aril is rich in lycopene and  $\beta$ -carotene content and

they are seen as a great natural red colorant. The highest content of lycopene in fresh aril in absolutely ripe fruit was reported as 3.728 mg/g while  $\beta$ -carotene was 0.379 mg/g (Nhung et al., 2010). Lycopene level is at higher five times compared to the tomatoes (Rao and Agarwal, 1999; Lenucci et al., 2009) and the  $\beta$ -carotene level is higher eight times compared to the carrots (Cefola et al., 2012). Two these constituents are reported to have high antioxidant effects (Aoki et al., 2002; Kubola et al., 2011). Recently, there are several reports of using potential of the Gac aril as a natural material to made food dye. The red aril powder could be used to enhance a yellow red color in many recipes such as glutinous rice, yogurt, cheese sauce and fettuccine pasta (Hoang, 2007). The content and retention of  $\beta$ -carotene and lycopene in Gac aril was changed by various temperatures. They significantly reduced in temperature condition at too high. The temperature at 60°C is the best to make natural food colorant (Tanongkankit et al., 2014).

#### Turmeric (Curcuma longa)

Turmeric (Curcuma longa) rhizome, an important tropical spice, belongs to ginger family Zingiberaceae (Aggarwal et al., 2005). The yellow savoury powder produced from its zhizomes have long been utilized in traditional medicince (Joshi et al., 2009). The attention has increased considerably over the years because of its benefits in the food colorant processing manufacture (Aggarwal et al., 2003). Although it is used popularly as a yellow flavour spicy in household, using to process sweet dishes is restricted because of its specific taste. Eating turmeric daily diet can help to protect and enhance human health (Nishiyama et al., 2005). Curcumin is the main coloring principal component extracted from its zhizomes and used as replacements for artificial dyes in the food processing field (Govindarajan, 1980). Yellow of turmeric is formed by three major colorant including curcumin, bis demethoxy curcumin and demethoxy curcumin (Khurana and Ho, 1988). Antioxidant capacity of them was reported by Toda et al. (1985). Yellow curcumin is seen as a food preservative agent and has great worth in pharmaceutical manufacture (Sampathu et al., 2000; Liang et al., 2007). Its anti-microbial evidences were also developed by Egan et al. (2004).

#### Pandanus amarylliforlius

Pandanus amaryllifolius, a tropical plant, is widely cropped and used as a natural green dye in Vietnam and other nations because its leaves contain high chlorophylls. Its leaves can be used to repel insect such as American cockroaches (Periplaneta americana L.) and also to produce dye in the food industry. Pandan consists of the following aromatic compounds, namely, 2-acetyl-1-pyrroline which was found in Basmati and Jasmine rice (Jiang, 1990; Laksanalamai and Ilangantileke, 1993). This plant can be a potential source of natural green pigment (Al-Alwani et al., 2017). Antioxidant effects were confirmed by the presence of quercetin (Miean and Mohamed, 2001), and other chemical components such as (Ferruzzi et al., 2002; Lee et al., 2004; Nor et al., 2008). It can be thought that Pandanus amaryllifolius is a worth source to make green pigments applied in food processing because of its values (Wissgott and Bortlik, 1996).

#### Gardenia jasminoides

The fruit of gardenia (*Gardenia jasminoides* Ellis.) was commonly used not only as a natural yellow dye but also one of the popular folk medicines since the detection of its biological property a few decades ago. Xiao et al. (2017) reported over 60 chemical compounds isolated from this plant. It has been used for the treatment of inflammation, fever, jaundice, headache, edema, hepatic disorders and hypertension (Tseng et al., 1995), and the extracts of gardenia fruit are in yellow or red (Toshiro and Shigeru, 2000), and have been consumed as food colorants in noodle and confectionery products in oriental countries (Yamada et al., 1996). In Japan, some 320 tons of gardenia yellow are consumed (Thantsin, 2011), and requirement has been increasing. Both water and ethanol extracts from its fruits had been found to exert antioxidant capacity. The water extract had a higher antioxidant compared to the ethanol extract (Pham et al., 2000). G. jasminoides - ethanolic extracts showed a protective effect against potential gastric disease due to the presence of ursolic acid and genipin which induce the death of AGS and SUN638 gastric cancer cells (Lee et al., 2009).

Antihyperlipidemia: the water extracts of *G. jasminoides* contain crocin displaying antihyperlipidemic effect. Crocin and crocetin significantly decreased the triglyceride and total cholesterol, but there is a significant increase in the high density lipoprotein-cholesterol levels in tested mice (Lee *et al.*, 2005).

Retinal demaging inhibition: Crocetin (100 mg/kg, p.o.) reduced retinal damage due to their antioxidant potential and by down-regulating caspase-3 and -9 activities after retinal damage. Moreover, crocetin showed improvement capacity of the quality of sleep in healthy adult man (Yamauchi *et al.*, 2011); Ishizuka *et al.*, 2013) and crocetin at dose of 50 mg/kg for 2 hours after resuscitation significantly relieved renal dysfunction induced by hemorrhage shock and resuscitation (Wang *et al.*, 2012).

## Liquidambar formosana Hance

Leaves of Liquidambar formosana Hance (LH) has been consumed as a rice dye for anniversary food by a small indigenous tribe in Guangxi province China (Wang et al., 2009). The LH leaves can be boiled and crunched for use in the soaking of sticky rice to make black and shiny rice with delicious taste (Chen, 1998). Zhang et al. (2017) reported 22 chemical constituents including phenolic acids (gallic acid, digallic acid, 3,5-dihydroxy-methoxybenzoic acid, ethyl gallate, gallic acid derivative, ethyl gallate-O-gallic acid, ethyl gallate-O-gallic acid isomer); flavanols (Catechin and Catechin derivative), flavonols, tannins and lignin isolated from its leaf. Ouyang et al. (2016) had reported 68 chemical constituents in the LH. The different solvent extracts from leaves showed outstanding antioxidant being higher positive control (Butylated hydroxytoluene) (Wang et al., 2010). As a kind of natural pigment resources, the LH has great potential in foods and cosmetics industry. Its leaves, a natural antioxidant source, are used as both a pigment and an important source in medicine and health care. All parts of this plant could be used in traditional medicine. The leaves have a clear aroma when rubbed, weinxin and a slightly bitter flavor (Huang et al., 2000) and are used as heat antidotes, to arrest diarrhea, and to treat dysentery, treat pain, heatstroke, postpartum wind, and pediatric tetanus neonatorium. Other studies also reported that its leaves possess many pharmacological functions as the treatment of diabetes and blood pressure (Zhong et al., 2010). Antibacterial effects were evaluated by Zhong et al. (2007) and Zheng et al. (2005). Leaves extract showed the growth inhibitory capacity of human leukemia K562 cells (Xei *et al.*, 2015) as well as enhance non-specific immune and cellular immune functions in normal mice (Zhong *et al.*, 2012). Extracts of the LH displayed significantly cytotoxic on human colon adenocinoma cells HT-29 (Zhang *et al.*, 2015). Some compounds from this plant also showed significant cytotoxicity against tumor cells (Dat *et al.*, 2004; Yang *et al.*, 2011). The extracts of LH leaves are commonly used as antibacterial sources in traditional folk medicine in China. L. formosana leaves showed strong effects against *Staphylococcus aureus, Staphylococcus, Shigella flexneri* (Zhong *et al.*, 2007). Anti-inflammatory activity was also reported (Hua *et al.*, 2014; Yang *et al.*, 2013).

# Basella rubra L.

Basella rubra L. belonging to Basellaceae has the potential benefits of pigment rich fruit extracts in food culture (Kumar et al., 2015a). Moreover, the pink colored tender twines and leaves of B. rubra are known as a leafy vegetable (Kumar et al., 2015b). Its ripened fruits possess plenteous betalains that can be used as food grade natural colorants. Fruit extracts contain phenolic acids (coumaric acid, generic acid, ferulic aicd and sinapic acid); flavonoids (myricetin, quercetin, luteolin, apigenin and kaemferol) (Kumar et al., 2015c). Kumar and Giridhar, 2016 have reported 348.2 mg/100 g of betalains with 296.6 betacyanins and 51.6 mg/100 g of betaxanthins from the fruit extracts (Kumar and Giridhar, 2016). Betalains are the water soluble natural red and yellow colored indole derived pigments found in the vacuoles of plant cells and have great potential as a natural dye in food processing industry and in pharmacology as antioxidant, anti-inflammatory, and detoxifying agents (Kanner et al., 2001).

# Conclusion

In traditional food culture of Vietnam, though 49 species can be consumed as natural pigments, 7 species were used mostly. Bioactive and pharmacological studies on their extracts and main bioactive components were reported. Based on real-life interviews and literature references and field survey, these data are firm evidence for further clinical application and also recommend producing in food processing and pharmaceutical industry.

# References

- Aggarwal, B. B., Kumar, A., and Bharti, A.C. (2003). Anticancer potential of curcumin: Preclinical and clinical studies. *Anticancer Research*, 23, 363-398.
- Aggarwal, B.B., Kumar, A., Aggarwal, M.S., and Shishodia, S. (2005). Curcumin derived from turmeric (*Curcuma longa*): a spice for all seasons. *Phytopharmaceuticals in Cancer Chemoprevention (Edited by H. Press)*. Pp. 349-387. Boca Raton: CRC Press.
- Al-Alwani, M.A.M., Mohamad, A.B., Kadhum, A.A.H., Ludin, N.A., Safie, N.E., Razali, M.Z., Ismail, M. and Sopian, K. (2017). Natural dye extracted from *Pandannus amaryllifolius* leaves as sensitizer in fabrication of dye-sensitized solar cells. *International Journal of Electrochemical science*, 12, 747-761.
- Aoki, H., Kieu, M.T.N. Kuze, N., Tomisaka, K., and Chuyen, V.N. (2002). Corotenoid pigments in Gac fruit

(Momordica cochinchinenensis Spreng). Bioscience, Biotechnology and Biochemistry, 66(11), 2479–2482.

- Cefola, M., Pace, B., Renna, M., Santamaria, P., Signore, A., and Serio, F. (2012). Compositional analysis ans antioxidant profile of yellow, orange and purple polignano carrots. *Italian Journal of Food Science*, 24, 284–291.
- Chen, Z.H. (1998). Study on extraction of melanin from the leaves of *Liquidambar formosana* Hance. *Journal of Chemical Industry of Forest Products*, 1, 20-21
- Cheng, Z., Lu, J., and Liu, J. (2004). Effects of *Peritrophe roxburghiana* on blood pressure in renal hypertensive and hyperlipidemic rats. *Zhong Yao Cai*, 27(12), 927-930.
- Chi, V.V. (1999). (A dictionary of Vietnamese Medicinal Plant), Tu dien Cay thuoc Vietnam. Medicine publication, Vietnam, pp.191. (In Vietnamese).
- Dam, N.A.L., Ninh, B.K., Sumimura, Y. (2016). Ethnobotany of Colorant Plants in Ethnic Communities in Northern Vietnam. *Anthropology*, 4(1), 158.
- Dat, N.T., Lee, I.S., Cai, X.F., Shen, G., and Kim, Y.H. (2004). Oleanane triterpenoids with inhibitory activity against NFAT transcription factor from *Liquidamber formosana*. *Biological and Pharmaceutical Bulletin*, 27(3), 426-428.
- Egan, M.E., Pearson, M., Weiner, S.A., Rajendran, V., Rubin, D., and Glockner, P.J. (2004). Curcumin, a major constituent of turmeric, corrects cystic fibrosis defects. *Science*, 304, 600-602.
- Ferruzzi, M.G., Böhm, V., Courtney, P.D., and Schwartz, S.J. (2002). Antioxidant and antimutagenic activity of dietary chlorophyll derivatives determined by radical scavenging and bacterial reverse mutagenesis assays. *Journal of Food Science*, 67(7), 2589–2595.
- Govindarajan, V.S. (1980). Turmeric-chemistry, technology, and quality. *Critical Reviews in Food Science and Nutrition*, 12,199-301.
- Hua, K.F., Yang, T.J., Chiu, H.W., and Ho, C.L. (2014). Essential oil from leaves of *Liquidambar formosana* ameliorates inflammatory response in lipopolysaccharide-activated mouse macrophages. *Natural Product Communications*, 9(6), 869-872.
- Huang, T.K., Ding, Z.Z., and Zhao, S.X. (2000). Xian Dai Ben Cao Gang Mu.1<sup>st</sup> Vol. China Press of Traditional Chinese Medicine, Beijing.
- Hoang, T.T. (2007). Producing carotenoid-rich powder from Gac fruit. College of Health and Science: Centre for Plant and Food Science. University of Western Sydney.
- Ishizuka, F., Shimazawa, M, Umigai, N., Ogishima, H., Nakamura, S., Tsuruma, K., and Hara, H. (2013). Crocetin, a carotenoid derivative, inhibits retinal ischemic damage in mice. *European Journal of Pharmacology*, 703, 1-10.
- Iwamoto, M., Okabe, H., Yamauchi, T., Tanaka, M., Rokutani, Y., Hara, S., Mihashi, K., and Higuchi, R. (1985). Studies on the constituents of *Momordica cochinchinensis* Spreng. I. Isolation and characterization of the seed saponins, momordica saponins I and

II. Chemical and Pharmaceutical Bulletin, 33, 1–7.

- Joshi, P., Jain, S., and Sharma, V. (2009). Turmeric (*Curcuma longa*) a natural source of edible yellow colour. *International Journal of Food Science and Technology*, 44, 2402–2406.
- Jiang, J. (1990). Volatile composition of pandan leaves (*Pandanus amaryllifolius*). In Flavor chemistry of ethnic foods. Shahidi, F. and Ho, C.T. Eds. pp. 105-109, Kluwer Academic, New York.
- Joseph, J.K., and Bharathi, L.K. (2008). Sweet gourd (Momordica cochinchinensis (Lour) Spreng.). Underutilized Underexploited Horticultural Crops, 4,185–191.
- Kapoor, V.P. (2006). Food Colours: Concern regarding their safety and toxicity. Environment Newsletter of ISEB India. *International Society of Environment Botanists*, 12.
- Kumar, J.K., and Sinha, A.K. (2004). Resurgence of natural colourants: A holistic view. *Natural Product Letters*, 18, 59-84.
- Khurana, A., and Ho, C.T. (1988). High performance liquid chromatographic analysis of curcuminoids and their photooxidative decomposition compounds in *Curcuma longa* L. *Journal of Liquid Chromatography*, 11, 2295-2304.
- Kumar, S.S. and Giridhar, P. (2016). Stabilization of bioactive betalain pigment from fruits of *Basella rubra*L. through maltodextrin encapsulation. *Madridge Journal of Food Technology*, 1(1), 66-70.
- Kanner, J., Harel, S. and Granit, R. (2001). Betalains- a new class of dietary cationized antioxidants. *Journal of Agricultural and Food Chemistry*, 49(11), 5178-5185.
- Kubola, J., and Siriamornpun, S. (2011). Phytochemicals and antioxidant activity of different fruit fractions (peel, pulp, aril and seed) of Thai gac (*Momordica cochinchinensis* Spreng.). *Food Chemistry*, 127, 1138– 1145.
- Kumar, S.S., Manoj, P. and Giridhar, P. (2015a). Nutrition facts and functional attributes of foliage of *Basella* spp. *LWT-Food Science and Technology*, 64(1), 468-474.
- Kumar, S.S., Manoj, P. and Giridhar, P. (2015b). A method for red-violet pigments extraction from fruits of Malabar spinach (*Basella rubra*) with enhanced antioxidant potential under fermentation. *Journal of Food Science and Technology*, 52(5), 3037-3043.
- Kumar, S.S., Manoj, P., Giridhar, P., Shrivastava, R. and Bharadwaj, M. (2015b). Fruit extracts of *Besella rubra* that are rich in bioactives and betalains exhibit antioxidant activity and cytotoxicity against human cervical carcinoma cells. *Journal of Functional Foods*, 15, 509-515.
- Laksanalamai, V., and Ilangantileke, S. (1993). Comparison of aroma compound (2-acetyl-1-pyrroline) in leaves from pandan (*Pandanus amaryllifolius*) and Thai fragrant rice (Khao Dawk Mali-105). *Cereal Chemistry*, 70, 381–384.
- Lee, B.L., Su, J., and Ong, C.N. (2004). Monomeric C18 chromatographic method for the liquid chromatographic

determination of lipophilic antioxidants in plants. *Journal Chromatography*, 1048: 263–267.

- Lee, I.A., Lee, J.H., Baek, N.I., and Kim, D.H. (2005). Antihyperlipidemic effect of crocin isolated from the fructus of *Gardenia jasminoides* and its metabolite crocetin. *Biological and Pharmaceutical Bulletin*, 28, 2106-2110.
- Lee, J.H., Lee, D.U., and Jeong, C.C. (2009). *Gardenia jasminodies* Ellis ethanol extract and its constituents reduce the risks of gastritis and reverse gastric lesions in rats. *Food Chemical Toxicology*, 47, 1127-1131.
- Liang, J. L., Meng, Y. Z., and Lei, C. G. (2007). Study on antiseptic effects of curcumin. *China Food Additives*. 2, 73-79.
- Lenucci, M. S., Caccioppola, A., Durante, M., Serrone, L., de Caroli, M., Piro, G., and Dalessandro, G. (2009). Carotenoid content during tomato (Solanum lycopersicum L.) fruit ripening in traditional and high-pigment cultivars. Italian Journal of Food Science, 21, 461–472.
- Miean, K.H., and Mohamed, S. (2001). Flavonoid (myricetin, quercetin, kaempferol, luteolin and apigenin) content of edible tropical plants. *Journal of Agricultural and Food Chemistry*, 49, 3106–3112.
- Nhung, D.T.T., Bung, P.N., Ha, N.T., and Phong, T.K. (2010). Changes in lycopene and beta carotene contents in aril and oil of gac fruit during storage. *Food Chemistry*, 121, 326–331.
- Nguyen, V.Q., Do, T.K., Luan, T.D., Truong, N.M., Nobukazu, N., and Tran, D.X. (2016). The Potential use of a fooddyeing plant *Peristrophe bivalvis* (L.) Merr. in Northern Vietnam. *International Journal of Pharmacology*, *Phytochemistry and Ethnomedicine*, 4, 14-26.
- Nor, F.M., Mohamed, S., Idris, N.A., and Ismail, R. (2008). Antioxidative properties of *Pandanus amaryllifolius* leaf extracts in accelerated oxidation and deep frying studies. *Food Chemistry*, 110, 319–327.
- Nishiyama, T., Mae, T, Kishida, H, Tsukagawa, M., Mimaki, Y., Kuroda, M., Sashida, Y., Takahashi, K., Kawada, T., Nakagawa, K., and Kitahara, M. (2005). Curcuminoids and Sesquiterpenoids in Turmeric (*Curcuma longa* L.) Suppress an Increase in Blood Glucose Level in Type 2 Diabetic KK-Ay Mice. Journal of Agricultural and Food Chemistry, 53(4), 959-63.
- Ouyang, X.L., Yi, S., Lu, H.Y., Wu, S.M., and Zhao, H.Q. (2016). *Liquidambar formosana* Hance: A mini-review of chemical constituents and Pharmacology. *European Journal of Medicinal Plants*, 17(1), 1-11.
- Pham, T.Q., Cormier, F., Farnworth, E., Tong, V.H., and Van Calsteren, M.R. (2000). Antioxidant properties of crocin from *Gardenia jasminoides* Ellis and study of the reactions of crocin with linoleic acid and crocin with oxygen. *Journal of and Food Chemistry*, 48, 1455-1461.
- Rao, A.V., and Agarwal, S. (1999). Role of lycopene as antioxidant carotenoid in the prevention of chronic diseases: a review. *Nutrition Research*, 19, 305–323.

Sampathu, S.R., Lakshminarayanan, S., Sowbhagya, H.B.,

Krishnamurthy, N., and Asha, M.R. (2000). Use of curcumin as a natural yellow colourant in ice cream. National Seminar on Natural Colouring Agents, February 2000, Lucknow, India.

- Tanongkankit, Y., Sutthaphan, T., Kaewmanas, J., Poonnoy, P., and Narkprasom, K. (2014). Evolutions of βcarotene and lycopene in a natural food colorant from Gac (Momordica cochinchinensis Spreng) arils during drying. International Conference on Nutrition and Food Sciences, 71(12), 56-60.
- Tanavade, S.S., Naikwade, N.S., and Chougule, D.D. (2012a). Antimicrobial activity of ethanolic extracts of leaves and stems of *Persistrophe bivalvis* Merrill. *International Journal of Biomedical Research*, 2, 106-108.
- Tanavade, S.S., Naikwade, N.S., and Chougule. (2012b). In vitro anticancer activity of ethanolic and aqueous extracts of *Peristrophe bivalvis* Merrill. *Research Journal of Pharmarcy and Technology*, 5(10), 1324-1327.
- Thantsin , K. (2011). Natural Colorant from Gardenia jasminoides Ellis (Cape Jasmine). Universities Research Journal, 4(1), 65-74.
- Toda, S., Miyase, T., Arichi, H., and Takino, Y. (1985). Natural anti- oxidants III. Antioxidative components isolated from rhizome of *Curcuma longa* L. *Chemical and Pharmaceutical Bulletin*, 33 (4), 1725-1728.
- Tran, T.H., Nguyen, M.H., Zabaras, D., and Vu, L.T.T. (2008). Process development of Gac powder by using different enzymes and drying techniques. *Journal of Food Engineering*, 85 (3), 359–365.
- Trinh, T.T., Nguyen, T.T.T.H., Le, T.H.N., Pham, T.N., Delfino, D.V., and Sung, T.V. (2014). Isolation, characterization and biological evaluation of a phenoxazine, a natural dyestuff isolated from leaves of *Peristrophe bivalvis*. *Natural Product Research*, 27(8), 771-774.
- Tseng, T.H., Chu, C.Y., Huang, J.M., Shiow, S.J. and Wang, C.J. (1995). Crocetin protects against demage in rat primary hepatocytes. *Cancer Letter*, 97, 61-67.
- Toshiro, W. and Shigeru, T. (2000). Analysis of natural food pigments by capillary electrophoresis. *Journal of Chromatography* A, 880, 311-322.
- Verma, R.K., Chaurasia, L., and Katiyar, S. (2008). Potential antifungal plants for controlling building fungi. *Indian Journal of Natural Products and Resources*, 7(4), 374-387.
- Wang, G., Tian, D., Yan, W, Zhu, F., and Li, S. (2009). Impact of litter addition an exclusion on soil respiration in a Liquidambar formosana forest and a nearby Cinnamomum camphora forest of central southern China. Acta Ecologica Sinia, 29(3), 1607-1615.
- Wang, K., Pan, Y., Wang, H., Zhang, Y., Lei, Q., Zhu, Z., Li, H., and Liang, M. (2010). Antioxidant activities of *Liquidambar formosana* Hance leaf extracts. *Medicinal Chemistry Research*, 19, 166-176.
- Wang, Y., Yan, J., Xi, L., Qian, Z., Wang, Z., and Yang, L. (2012). Protective effect of crocetin on hemorrhagic shock-induced acute renal failure in rats. *Shock*, 38(1), 63-67.

- Wiart, C., Mogana, S., Khalifah, S., Mahan, M., Ismail, S., Buckle, M., Narayana, A.K., and Sulaiman, M. (2004). Antimicrobial screening of plants used for traditional medicine in the state of Parak, Peninsular Malaysia. *Fitoterapia*, 75(1), 68-73.
- Wissgott, U., and Bortlik, K. (1996). Prospects for new food colorants. *Trends in Food Science and Technology*, 7, 289-302.
- Xiao, W., Li, S., Wang, S., and Ho, C.T. (2017). Chemistry and bioactivity of *Gardenia jasminoides*. Journal of Food and Drug Analysis, 25, 43-61.
- Xie, Q.J, Xu, X.Y., Zheng, J.R., and Zhong, Y.T. (2015). Study on effect of extract from the leaves of *Liquidambar formosana* Hance on K562 cells. *Chinese Materia Medica*, 7, 1493-1495.
- Yang, W., Gu, F., Lu, J. and Yang, M. (2002). Effect of the extract from *Peristrophe roxburghiana* on hemorheology in rats. *Zhong Yao Cai*. 25(10), 727-728.
- Yang, N.Y., Chen, J.H., Zhou, G.S., Tang, Y.P., Duan, J.A., Tian, L.J., and Liu. X.H. (2011). Pentacyclic triterpense from the resin of *Liquidambar formosana*. *Fitoterapia*, 82(6), 927-931.
- Yang, Z.J., Zhang, Y.H., Feng, J., Yao, X.Q., and Jin, X.H. (2013). Pharmacological activity and chemical composition of *Liquidambar formosana* leaves volatile oil. *Science and Technology of Food Industry*, 34(4), 76-79.
- Yamada, S., Oshima, H., Saito, I., and Hayakawa, J. (1996). Adoption of crocetin as an indicator compound for detection of gardenia yellow in food products (Analysis of natural coloring matters in food V). *Journal of the Food Hygienic Society of Japan*, 37, 372-377.

- Zhang, J., Chou, G., Liu, Z., and Koh, G.Y. (2015). In vitro cytotoxicity and antioxidation of a whole fruit extract of *Liquidambar formosana* exerted by different constituents. *European Journal of Medicinal Plants*, 6(1), 34.
- Zhang, L., Zhu, M.F., Tu, Z.C., Zhao, Y., Wang, H., Li, G.J., Wang, H., and Sha, X.M. (2017). α-Glucosidase inhibition, anty-glycation and antioxidant activities of *Liquidambar formosana* Hance leaf, and identification of phytochemical profile. *South African Journal of Botany*, 113, 239-247.
- Zheng, Y., Liu, N.F., Xiao, W.H., and Zhang, Q. (2005). Study on preventing effect of extract from the leaves of *Liquidambar formosana* Hance on pepper black spot. *Journal of Jiangxi Agricultural University*, 27, 96-96
- Zhong, Y.T., Wang, L., Wang, X.L., and Sun, X.T. (2010). Determination of trace elements in the leaves of *Liquidambar formosana* Hance by flame atomic absorption spectrometry. *LiShiZhen Medicine and Materia Medica Research*, 21, 1457-1458.
- Zhong, Y.T., Wang, X.L., Sun, X.T., Xu, J., Wang, L. and Zhang, W.P. (2012). The sudy of ex-tract from leaves of *Liquidambar formosana* Hance on immue-regulatory effects in mice. *Pharmacology and Clinics of Chi-nese Metaria Medica*, 28(1), 124-126.
- Zhong, Y.T., Wang, X.L. and Ma, L.L. (2007). Study on antibacterial activity of the leaves of *Liquidambar* formosana Hance. *LiShiZhen Medicine and Materia Medica Research*, 18, 1693-1694.
- Yamauchi, M., Tsuruma, K., Imai, S., Nakanishi, T., Umigai N., Shimazawa, M., and Hara, H. (2011). Crocetin prevents retinal degeneration inculded by oxidative and endoplasmic reticulum stresses via inhibition of caspase activity. *European Journal of Pharmacology*, 650, 110-119.