

Efficacy of Azadirachtin Treated Nets on Adults of *Aedes aegypti* and *Culex quinquefasciatus* (Diptera: Culicidae)

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Abstract

Mosquitoes are vectors of a variety of diseases like malaria, dengue, chikungunya and filariasis causing several thousand deaths throughout the tropical and subtropical regions of the world. Vector control measures involving the use of insecticide treated net have become popular and has been recommended by world health organization for management of mosquitoes. However the effectiveness of the treated net is facing a threat due to the problem of emergence of insecticide resistance among mosquitoes. Alternatives to the conventional insecticides are being explored with emphasis on mammalian safety and prevention of rapid resistance development in insects. Botanical derivatives have proven to be an effective option for the same purpose. One such chemical is Azadirachtin. It is extracted from neem tree and is known to have potent toxicity against several insects. In the present study we tested the efficacy of Azadirachtin treated nets as a potential mosquito adulticide. Mosquito nets were treated with varying concentration of Azadirachtin in order to determine the optimum dose of the chemical required for a mortality of adult mosquitoes. Two species of mosquitoes which are abundant in Central Gujarat region were tested viz. *Aedes aegypti* and *Culex quinquefasciatus*. Both the insects were found to be susceptible to the Azadirachtin however LC₅₀ and LC₉₉ values for *Ae.aegypti* were lower suggesting its higher susceptibility to Azadirachtin compared to *Cx.quinquefasciatus*. The present study shows that Azadirachtin is effective in managing the population of adult mosquitoes and can be used as an alternative for conventional insecticides used in the insecticide nets.

1. Introduction

Malaria and other vector-borne diseases such as dengue, filariasis¹, and Japanese encephalitis² are major contributors to the communicable disease burden in the South-East Asia Region. The risk of different communities to vector-borne diseases varies according to their socioeconomic status (e.g. tribes, urban slum dwellers), physiological conditions (e.g. pregnant women, young children), geographical location (e.g. those living in areas with poor access to interventions, and in cross-border areas), occupational exposures (e.g. migrant workers, miners), natural hazards (e.g. disease exacerbated by earthquakes, floods, cyclones etc) and socio-political disturbances (e.g. refugees).

Two generations ago hopes were high that malaria could be eradicated. But from 1980s till date, malarial vectors have gained resistance to several classes of insecticides and pathogen has developed resistance to antimalarial drugs. Malaria has re-emerged in several Central Asian and Eastern European countries and in South-East Asia³. Pregnant women are high susceptibility to malaria and other diseases⁴⁻⁶. Mother and foetus are faced with adverse consequences which may even

lead to the death of foetus⁷⁻¹⁰ and in certain cases leading to maternal anemia¹¹ and of low birth weight⁶.

Measures to fight malaria epidemic are two pronged, one by regulating the vector population and other by antimalarial drugs. Among the two, the former approach is cheaper and most efficient in protecting large population throughout the world. Several vector control measures like larvicidal control measures, indoor residual spraying have been undertaken, which though effective for a short time has its drawbacks. The spraying of pesticides especially pyrethroids and organochlorines expose the human population to the risk of pesticide toxicity and its residual effects. These vector control measures are costly when a large area has to be brought under vector control program.

The alternative method which is much more efficient, cheaper and long lasting is the use of insecticide treated nets (ITNs)³. ITNs when used by all the population of a community can bring down vector borne diseases by as much as 50%¹⁻². ITNs treated with conventional insecticides like

pyrethroids have started becoming less effective due to the development of insecticide resistance among mosquitoes. In a few people using ITNs treated with pyrethroids has led to dermal irritation. Hence there is a need for a much more ecofriendly alternative which could solve the problem of insecticide resistance and at the same time is safer to humans.

One such alternative is botanical Azadiracthin extracted from Neem tree. Azadiracthin has been recognized as a potential insecticide having several properties like antifeedant activity, insect growth regulator, and steriliant activity¹²⁻¹⁴. In the present study we analyzed the efficacy of mosquito nets treated with Azadiracthin on the survival of vector population of *Culex quinquefasciatus* and *Aedes aegypti*. We also calculated the optimised dose required for the total management of adults of *Cx. quinquefasciatus* and *Ae. aegypti*.

2. Materials and Methods

Mosquito larvae were collected from urban areas of the city from overhead tanks, ponds, stagnant water bodies and brought to Insectary. The larvae were isolated based on their morphological characteristics and were reared till adulthood. Larvae were maintained in plastic trays having tap water and a diet comprising of glucose biscuit and brewer's yeast was provided. Water in beaker was changed every day. The larval culture was maintained at a temperature of $29 \pm 2^\circ \text{C}$, relative humidity: 60% and 14:10h L: D photoperiod. The pupae developing from the larvae were separated from the larval stock and transferred to 500 ml glass beakers. Beakers containing pupae were kept in wire gauze rearing cages (45cm (L) \times 45cm (B) \times 45cm (H)); the adults emerging from pupae were then kept in the rearing cage. The newly emerged adults were provided with honey solution and blood meal on alternative days using rat. Plastic container having tap water was kept in adult rearing cages for oviposition. Eggs were harvested at regular intervals and transferred to glass beaker having water for hatching.

Bioassay Protocol: Polyester multifilament net having mesh size of 156 holes/inch, light pink in colour was used in the present study. The net (50cm x 50cm) was treated with the Azadiracthin and placed vertically in the center of Bioassay cage (45 x 45 x 45cms). A set of 9 holes (each 1 cm diameter) were made horizontally in the centre of the net. 15 sugar fed mosquitoes one day old were released in the cage in every test. The net was placed in such a way that the mosquitoes released from one side can pass to the other side only through holes by touching the net. This ensured the maximum likelihood of mosquitoes coming in

contact with the net while moving from one end of the cage to other. Observations regarding mortality of the mosquitoes were taken after 12 hours after the release of mosquitoes. The test was repeated 20 times and appropriate control tests having water soaked polyester net were performed.

The insecticide used in the study was procured from Nicoorgo Manures Pvt ltd. The chemical is neem based product having Azadiracthin as active ingredient. Four concentrations of 50,100,200 and 300 ppm respectively were prepared from the stock solution. Polyester net was soaked in the insecticide solution overnight and then was allowed to dry in shade. This was done in accordance to WHOPES (2002) regulation regarding the use of insecticide treated net.

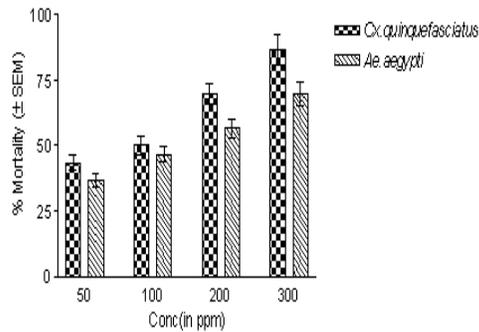
Data Analysis: Percentage mortality was corrected relative to the controls using Abbott's formula¹⁵. The data was subjected to probit analysis in accordance to Finney¹⁶ to determine the optimum concentration of LC₅₀ and LC₉₉ for *Cx. quinquefasciatus* and *Ae. aegypti* using statistical software SPSS (ver 7.65).

3. Results and Discussion

The results of the study show that *Ae. aegypti* were susceptible at lower concentration of azadiracthin whereas *Cx. quinquefasciatus* mosquitoes showed susceptibility at comparatively higher doses. LC₅₀ and LC₉₉ values calculated using Probit table gave us the diagnostic concentration at which 50% and 99% of test populations are killed (Table 1). The data after 12 hour post exposure to Azadiracthin shows that *Ae. aegypti* (LC₅₀ – 46.606 ppm; LC₉₉ – 323.815 ppm) is more susceptible than *Cx. quinquefasciatus* (LC₅₀ – 77.774 ppm; LC₉₉ – 462.363 ppm) (Figure I).

The global strategy adopted by WHO in managing vector borne diseases using integrated vector control, lays emphasis on selective vector control methods. Among these methods, insecticide treated nets have been very effective comparable to indoor residual spray (IRS) and are also cheaper¹⁷. ITNs have been recommended for malaria preventions by WHO¹⁸⁻¹⁹. Presently only pyrethroid treated polyester nets have been used for community vector control programs. These net have been popular because they produce knockdown and lead to mortality among mosquitoes²⁰.

Figure 1: Percent Mortality (\pm SEM) of *Cx. quinquefasciatus* and *Ae. aegypti* adults exposed to different doses of Azadirachtin treated net



In recent years mosquitoes have started gaining resistance to pyrethroids²¹⁻²² due to their intensive use as household insecticides. This has led to decreased effectiveness of ITNs as a vector control tool²³. Washing of ITN has led to decreased bioefficacy of these nets²⁴ and permethrin treated nets have been known to cause dermal irritation in

a few people. Alternatives to pyrethroids are botanicals most of which are relatively safer to human and other non target organisms and at the same time show selective toxicity towards insect pests.

In this study the effectiveness of Azadirachtin shows variation in LC₅₀ and LC₉₉ values in both the species of mosquitoes (Table 1) varies in different species of insects²⁵. This emphasizes the need for a proper insecticide resistance monitoring program which should look into the resistance levels among mosquitoes at least on a yearly basis. This would help in the dose optimization for widespread use in the community health programs. The present study shows that Azadirachtin treated nets can be effective in the control of *Ae. aegypti* and *Cx. quinquefasciatus*. Higher diagnostic dose for *Culex* mosquitoes gives us indication about the development of cross resistance as a result of exposure to conventional insecticides used earlier in the study area.

Table 1: Probit regression response of *Cx. quinquefasciatus* and *Ae. aegypti* to Azadirachtin treated net

Species	LC ₅₀		LC ₉₉	
	LC ₅₀ (in ppm)	95% CI	LC ₉₉ (in ppm)	95% CI
<i>Cx. quinquefasciatus</i>	77.463	-77.774 to 135.229	462.363	325.892 to 1106.450
<i>Ae. aegypti</i>	46.606	-100.298 to 94.700	323.815	232.979 to 726.196

In the study shows that nets treated with Azadirachtin can form an alternative for pyrethroids which can be effective in reducing the problem of insecticide resistance among mosquitoes. They can act as a cheaper and effective alternative in community vector control programs. Further studies needs to be undertaken regarding the efficacy of these net after washing and their efficacy against other species of mosquitoes also needs to be explored. However, the present studies are encouraging and suggest that the use of ITN treated with Azadirachtin can be a safe, cheap and effective alternative in community vector control programs.

References

1. Reyburn H., Ashford R., Mohsen M., Hewitt S., Rowland M., A randomized controlled trial of insecticide-treated bednets and chaddars or top sheets, and residual spraying of interior rooms for the prevention of cutaneous

- leishmaniasis in Kabul, Afghanistan Transactions of Royal Society of Tropical Medicine and Hygiene, 94, 361-366 (2000)
2. Dapeng L., Renguo Y., Jinduo S., Hongru H., Wang Z., The effect of DDT spraying and bed nets impregnated with pyrethroid insecticide on the incidence of Japanese encephalitis virus infection. Transactions of Royal Society of Tropical Medicine and Hygiene, 88: 629-631(1994)
3. WHO., Roll Back Malaria Department and UNICEF. Pp 293. www.ops-oms.org/English/ad/dpc/cd/mal-world-rpt-2005.htm, (2005)
4. Steketee RW., Nahlen BL., Parise ME., Menendez C., The burden of malaria in pregnancy in malaria-endemic areas. American Journal of Tropical Medicine and Hygiene, 64: 28-35 (2001)
5. Lindsay S., Ansell J., Selman C., Cox V., Hamilton K., Effect of pregnancy on exposure

- to malaria mosquitoes. *Lancet*, 355: 1972 (2000)
6. Brabin BJ., An analysis of malaria in pregnancy in Africa. *Bulletin of World Health Organization*, 61: 1005–1016 (1983)
 7. Nosten F., Rogerson SJ., Beeson JG., McGready R., Mutabingwa TK., Malaria in pregnancy and the endemicity spectrum: What can we learn? *Trends in Parasitology*, 20: 425–432 (2004)
 8. Van Geertruyden JP., Thomas F., Erhart A., D'Alessandro U., The Contribution of malaria in pregnancy to perinatal mortality. *American Journal of Tropical Medicine and Hygiene*, 71: 35–40 (2004)
 9. Guyatt HL., Snow RW., Malaria in pregnancy as an indirect cause of infant mortality in sub-Saharan Africa. *Transactions of Royal Society of Tropical Medicine and Hygiene*, 95: 569-576 (2001a)
 10. Marchant T., Schellenberg JA., Nathan R., Abdulla S., Mukasa O., Anaemia in pregnancy and infant mortality in Tanzania. *Tropical Medicine and International Health*, 9: 262–266 (2004)
 11. Guyatt HL., Snow RW., The epidemiology and burden of *Plasmodium falciparum* related anemia among pregnant women in sub-Saharan Africa. *American Journal of Tropical Medicine and Hygiene*, 64:36-44(2001b)
 12. Aerts RJ., Mordue AJ., Luntz A., Feeding deterrence and toxicity of neem triterpenoids. *Journal of Chemical Ecology*, 23: 2117- 2133, (1997)
 13. Dorn A., Effects of azadirachtin on reproduction and egg development of the heteropteran *Oncopeltus fasciatus* Dallas. *Journal of Applied Entomology*, 102: 313-319 (1986)
 14. Blaney WM., Sommonds MSJ., Ley WV., Anderson JC., Toogood PL., Antifeedant effects of azadirachtin and structurally related compounds on lepidopterous larvae. *Entomologia Experimentalis et applicata*, 55: 149-160 (1990)
 15. Abbott WS., A method for computing the effectiveness of insecticides. *Journal of Economic Entomology*, 18: 265-267, (1925)
 16. Finney DJ., *Probit Analysis*, third ed. Cambridge University Press, New York, (1975)
 17. Curtis CF., Mnzava AEP., Comparison of house spraying and insecticide-treated nets for malaria control. *Bulletin of World Health Organization*, 78: 1389-1400 (2000)
 18. Lengler C., Cattani J., Savigny D., *Net Gain: A new method for preventing malaria deaths*. International Development Research Centre, Ottawa, World Health Organisation, Geneva, (1996)
 19. Ziam M., Aitio A., Nakashima N., Safety of Pyrethroid-treated nets. *Medical and Veterinary Entomology*, 14: 1-5 (2000)
 20. Elliot M., The Pyrethroids: early discovery, recent advance and the future. *Pesticide Science*, 27: 337-351 (1989)
 21. Hargreaves K., Koekemoer LL., Brooke BD., Hunt RH., Mthembu J., Coetzee M., *Anopheles funestus* resistant to pyrethroid insecticides in South Africa. *Medical and Veterinary Entomology*, 14: 1- 9 (2000)
 22. Hemingway J., Lindsay SW., Small GJ., Jawara M., Collins FH., Insecticide susceptibility status in individual species of *Anopheles gambiae* complex (Diptera: Culicidae) in an area of the Gambia where pyrethroid impregnated bednets are used extensively for malaria control. *Bulletin of Entomological Research*, 85: 229-234 (1995)
 23. Etang J., Chandre F., Guillet P., Manga L., Reduced bio-efficacy of permethrin EC impregnated bednets against *Anopheles gambiae* strain with oxidase-based pyrethriod tolerance. *Malaria Journal*, 3: 46 (2004)
 24. Rafinejad J., Vatandoost H., Nikpoor F., Abai MR., Shaeghi M., Duchon S., Rafi F., Effect of washing on the bioefficacy of insecticide treated nets (ITNs) and long-lasting insecticidal nets (LLINs) against malaria vector *Anopheles stephensi* by three bioassay methods. *Journal of Vector Borne Diseases*, 45: 143-150 (2008)
 25. Perry AS., Yamamoto I., Ishaaya I., Perry R., *Insecticides in Agriculture and Environment - Retrospects and Prospects*. Narosa Publishing House, NewDelhi, pp 208-220 (1998)