



ISSN: 2455-9377

# Effect of heavy metals on phenylpropanoid biosynthesis in *Euonymus alatus*

Ramaraj Sathasivam<sup>1†</sup>, Haeng-Hoon Kim<sup>2†</sup>, Bao Van Nguyen<sup>3</sup>, Jiwon Yoon<sup>1</sup>, Byung Bae Park<sup>4</sup>, Jae Kwang Kim<sup>5\*</sup>, Sang Un Park<sup>1,3\*</sup>

<sup>1</sup>Department of Crop Science, Chungnam National University, 99 Daehak-ro, Yuseong-Gu, Daejeon 34134, Republic of Korea, <sup>2</sup>Department of Agricultural Life Science, Suncheon National University, Suncheon 57922, Republic of Korea, <sup>3</sup>Department of Smart Agriculture Systems, Chungnam National University, 99 Daehak-ro, Yuseong-gu, Daejeon 34134, Republic of Korea, <sup>4</sup>Department of Environment and Forest Resources, Chungnam National University, 99 Daehak-ro, Yuseong-gu, Daejeon 34134, Republic of Korea, <sup>5</sup>Division of Life Sciences and Convergence Research Center for Insect Vectors, College of Life Sciences and Bioengineering, Incheon National University, Yeonsu-gu, Incheon 22012, Republic of Korea  
<sup>†</sup>Contributed equally to this work

## ABSTRACT

The productivity of the phenylpropanoid biosynthesis pathway in plants varies depending on the type of stress. In this work, we looked into how different phenylpropanoid chemicals accumulated in *Euonymus alatus* following exposure to different concentrations of CuCl<sub>2</sub> (0.1, 0.5, and 1 mM), HgCl<sub>2</sub> (0.1, 0.5, and 1 mM), and NiSO<sub>4</sub> (10, 50, and 100 mM). We analyzed some of the individual phenolic chemicals by high-performance liquid chromatography (HPLC). In nearly all cases, rutin showed the largest concentration among the phenylpropanoid chemicals, followed by epicatechin, sinapic acid, *p*-coumaric acid, *trans*-cinnamic acid, ferulic acid, and caffeic acid. However, due to the change in the concentration of the heavy metals, the amount of phenylpropanoid changed. The highest accumulation of phenylpropanoid was documented in 0.1 mM CuCl<sub>2</sub>, whereas it was reduced in 1 mM HgCl<sub>2</sub> exposed plants. These findings unequivocally demonstrate that the phenylpropanoid metabolic pathway took part in the heavy metal tolerance process, which shielded *E. alatus* from the oxidative damage brought on by heavy metals. Thus, under a variety of environmental stress situations, this species with a high tolerance to heavy metals may survive.

**KEYWORDS:** *Euonymus alatus*, Heavy metals, Phenolic compounds, CuCl<sub>2</sub>, HgCl<sub>2</sub>, NiSO<sub>4</sub>

**Received:** July 14, 2023  
**Revised:** November 26, 2023  
**Accepted:** November 29, 2023  
**Published:** December 08, 2023

### \*Corresponding authors:

Jae Kwang Kim  
E-mail: kjkpj@inu.ac.kr  
Sang Un Park  
E-mail: supark@cnu.ac.kr

## INTRODUCTION

*Euonymus alatus*, commonly identified as winged spindle or burning bush, is a deciduous tree species of flowering plant belonging to the Celastraceae family. It is disseminated mainly in China, Japan, and Korea. Because of its eye-catching fall color and bright pink or orange fruit, this plant is a favorite ornamental in gardens and parks (Qin *et al.*, 2011; Ning *et al.*, 2022). In traditional Chinese and Korean medicine, corky-winged stems are used. This plant can treat cancer, hyperglycemia, and complications from diabetes (Fan *et al.*, 2020; He *et al.*, 2022). The following secondary metabolites i.e., flavonoids, terpenoids, steroids, lignans, cardenolides, phenolic acids, and alkaloids have been documented from this plant in different experiments (Zhai *et al.*, 2016).

A class of long-lasting, non-biodegradable inorganic chemical substances known as heavy metals causes harm

to both people and plants as well as animals. They are extremely harmful to plant tissue at greater doses (Lombardi & Sebastiani, 2005), which has a substantial impact on the plastoquinone, carotenoid level, electron transport system, and chloroplast (Kisa *et al.*, 2016). Large tracts of land are contaminated with heavy metals, particularly by inorganic pollutants brought on by urbanization, industrial waste, and agriculture (Demirezen & Aksoy, 2006; Kisa *et al.*, 2016). It is becoming more and more crucial to develop heavy metal-tolerant plants, which can detoxify heavy metals, in order to solve this issue. Because it is inexpensive and simple to operate and maintain, plant-based phytoremediation of heavy metals is one of the most successful methods (Kivaisi, 2001; Madera-Parra *et al.*, 2015). Most plants accumulate secondary metabolites as a result of abiotic stressors (Zhao *et al.*, 2005). Among such metabolites, the most significant and often studied metabolic pathway is the phenylpropanoid pathway (Biala & Jasinski, 2018).

Copyright: © The authors. This article is open access and licensed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0/>) which permits unrestricted, use, distribution and reproduction in any medium, or format for any purpose, even commercially provided the work is properly cited. Attribution — You must give appropriate credit, provide a link to the license, and indicate if changes were made.

Phenylpropanoids compounds have anti-allergenic, anti-bacterial, anti-inflammatory, antioxidant, and anti-viral activities, which are beneficial to the health of humans (Cevallos-Casals & Cisneros-Zevallos, 2010; Korkina *et al.*, 2011; Panda *et al.*, 2011). Furthermore, these substances have anti-diabetic and anti-cancer properties, aiding in the prevention of cardiovascular illnesses (Yang *et al.*, 2001; Yao *et al.*, 2004). Additionally, they are essential mediators for the interactions between plants and other creatures and play a critical role in responding to stressful circumstances like fluctuations in light intensity and mineral deficits (Clemens & Weber, 2016). Kisa *et al.* (2016) studied how heavy metals affect the metabolic profile of phenolic compounds in plants. Nevertheless, it is unknown how heavy metals affect *E. alatus*'s phenylpropanoid profile and how the phenylpropanoid pathway contributes to the species' ability to survive in adverse environments. It is therefore likely that the phenylpropanoid accumulation is responsible for this tolerance capacity. Gaining insight into the process that allows heavy metals to be tolerated might improve the effectiveness of reducing heavy metals effect.

## MATERIALS AND METHODS

### Plant Materials

The studied plating material of *Euonymus alatus* (one-year-old) grown from a cutting was procured from Xplant (Seoul, Korea) and arranged into individual pots 11 cm × 11 cm in size. For five months, the one-year-old *E. alatus* trees were raised in a greenhouse at Chungnam National University's experimental farm in Daejeon, South Korea. After reaching a height of 70 cm, the trees were treated with different concentrations of heavy metals.

### Heavy Metal Stress Treatments

Different concentrations of CuCl<sub>2</sub> (0.1, 0.5, and 1 mM), HgCl<sub>2</sub> (0.1, 0.5, and 1 mM), and NiSO<sub>4</sub> (10, 50, and 100 mM) were tested, in order to know the impact of Cu, Hg, and Ni toxicity on *E. alatus*. The standard stock solution was used to create all the working concentrations, whereas all of the heavy metals were acquired from a commercial source (Sigma, St. Louis, MO, USA). Three duplicates were carried out for each experimental dosage.

### Analysis of Phenylpropanoid Content and High-Performance Liquid Chromatography (HPLC)

The leaf samples of *E. alatus* were harvested weekly following heavy metal treatments. HPLC was used to determine the phenylpropanoid content of the samples after they were freeze-dried for 72 hours. A small modification was made to the previously described methodology of Kim *et al.* (2020) for the analysis of the soluble phenylpropanoid compounds. Using a mortar and pestle, the stored samples were taken and ground into a fine powder. Each sample was weighed out at 100 mg and put into 3 mL of 80% aqueous MeOH for HPLC analysis. After one minute of vortexing, the mixtures were immediately

subjected to a vigorous 60-minute sonication at 35 °C. After centrifuging sonicated samples at 10,000 × g for 15 minutes, the supernatants were gathered, filtered, and sterilized using 0.45 μm filters in preparation for HPLC analysis. Reversed-phase chromatography was used to perform HPLC separation on an Agilent 1260 Infinity Quaternary LC (Agilent Technologies, Inc., Germany) using a C18 column (250 × 4.6 mm, 5 μm, RStech, Daejeon, South Korea). The mobile phase, gradient programs, HPLC conditions, identification, and quantification of phenylpropanoid compounds were similar to the protocol described by Kim *et al.* (2022).

### Statistical Analysis

The results were analyzed using IBM SPSS Statistics 24 software. Duncan Multiple Range Test with one-way ANOVA at the 5% significance level was used for the analyses. For each treatment, all experiments were done in triplicate.

## RESULTS

### Effect of CuCl<sub>2</sub> on Phenylpropanoid Contents

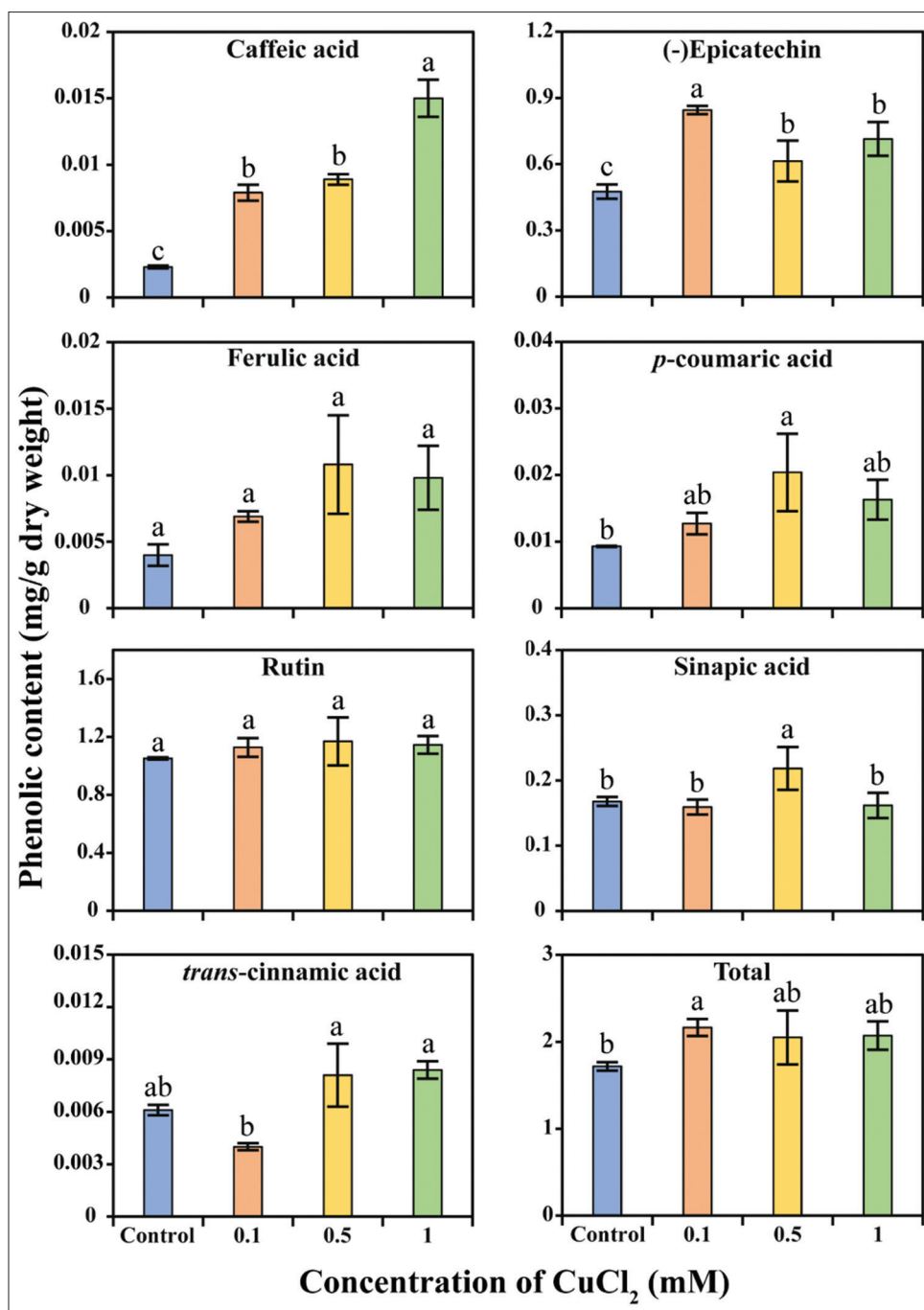
Depending on the concentration of CuCl<sub>2</sub>, *E. alatus* exhibited a wide variety of responses related to phenylpropanoid biosynthesis. The total phenylpropanoid contents reached 2.16, 2.05, and 2.07 mg/g dry weight at exposure to 0.1, 0.5, and 1 mM CuCl<sub>2</sub>, respectively. At all the concentrations, the highest phenylpropanoid content (mg/g dry weight) was obtained for rutin followed by epicatechin, sinapic acid, *p*-coumaric acid, ferulic acid, *trans*-cinnamic acid, and caffeic acid. However, when compared to the control, *trans*-cinnamic acid showed a decreased accumulation at 0.1 mM CuCl<sub>2</sub>. Most of the phenylpropanoid contents were higher in the plant exposed to 0.5 mM CuCl<sub>2</sub>, except for caffeic acid and epicatechin (Figure 1).

### Effect of HgCl<sub>2</sub> on Phenylpropanoid Contents

There was a minor impact of HgCl<sub>2</sub> on the phenylpropanoids' accumulation. The total phenylpropanoid production (mg/g dry weight) was slightly higher at control followed by 0.5, 0.1, and 1.0 mM of HgCl<sub>2</sub>. The phenylpropanoid level decreased at higher doses. In contrast, most of the phenylpropanoid contents showed slight or little accumulation in the plant exposed to HgCl<sub>2</sub>. From the overall dose, the highest phenylpropanoid contents (mg/g dry weight) were observed for rutin followed by epicatechin, sinapic acid, *trans*-cinnamic acid, *p*-coumaric acid, ferulic acid and caffeic acid (Figure 2).

### Effect of NiSO<sub>4</sub> on Phenylpropanoid Contents

The plant exposed to different concentrations of NiSO<sub>4</sub> showed that overall phenylpropanoid concentration rose significantly. The plants subjected to 100 mM NiSO<sub>4</sub> exhibited greater accumulations than those exposed to 10 mM and 50 mM. The total phenylpropanoid accumulation increased with increasing the concentration of heavy metals. Rutin had the



**Figure 1:** Effect of different concentrations of  $\text{CuCl}_2$  on phenylpropanoid compounds in *Euonymus alatus*

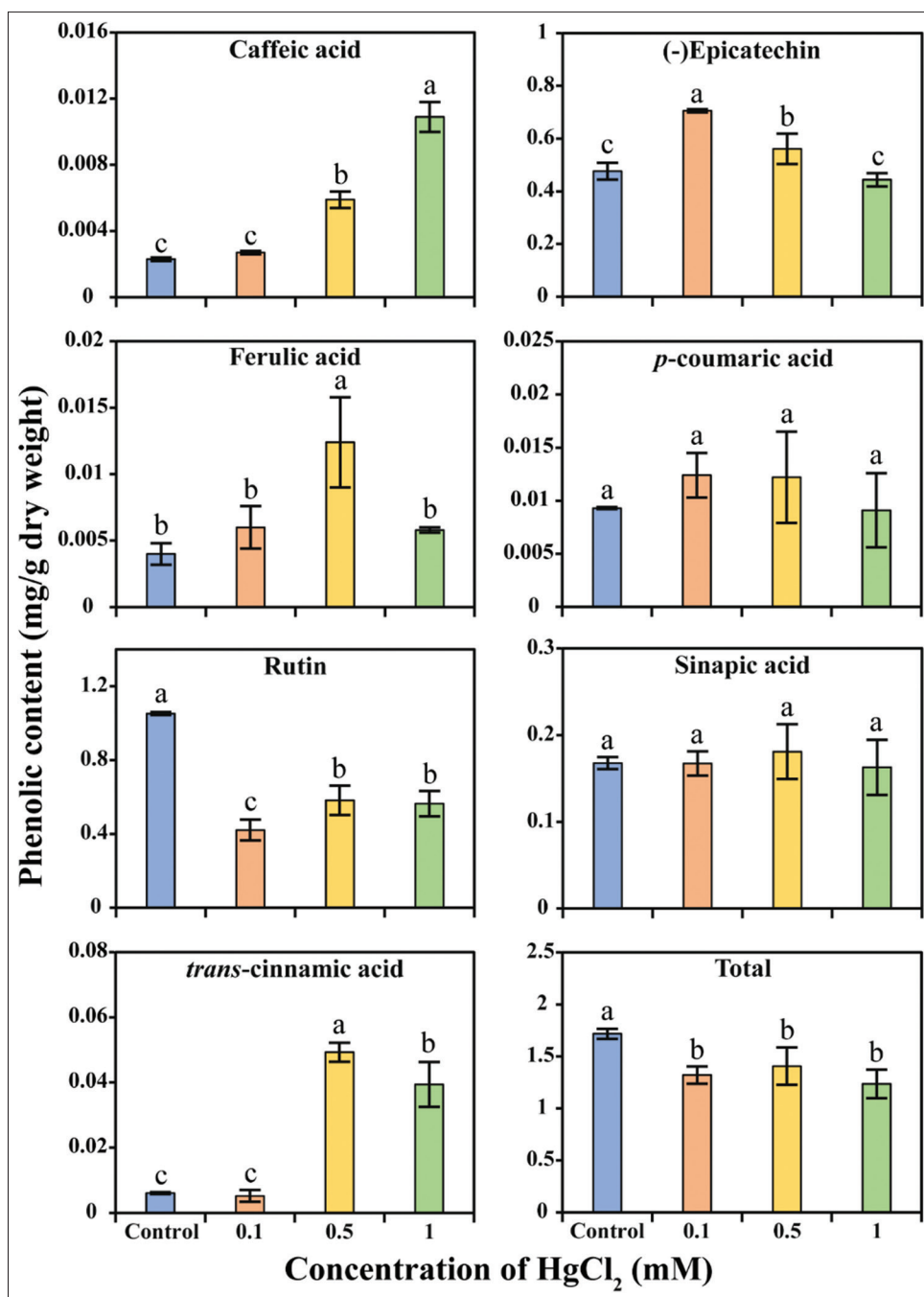
highest concentration (mg/g dry weight), whereas epicatechin, sinapic acid, p-coumaric acid, trans-cinnamic acid, ferulic acid, and caffeic acid showed close concentrations as that of the rutin. On the other hand, the levels of trans-cinnamic acid and caffeic acid were considerably lower than in the control (Figure 3).

## DISCUSSION

The way that plants react to stress may be influenced by phenylpropanoid chemicals. The productivity of the phenylpropanoid biosynthesis pathway in plants varies

depending on the type of stress. In this work, we examined the accumulation of several chemicals involved in the phenylpropanoid pathway following exposure to different heavy metals. Using varying doses of  $\text{CuCl}_2$ ,  $\text{HgCl}_2$ , and  $\text{NiSO}_4$ , the impact of Cu, Hg, and Ni toxicity on the phenylpropanoid biosynthesis of *E. alatus* was evaluated in this work.

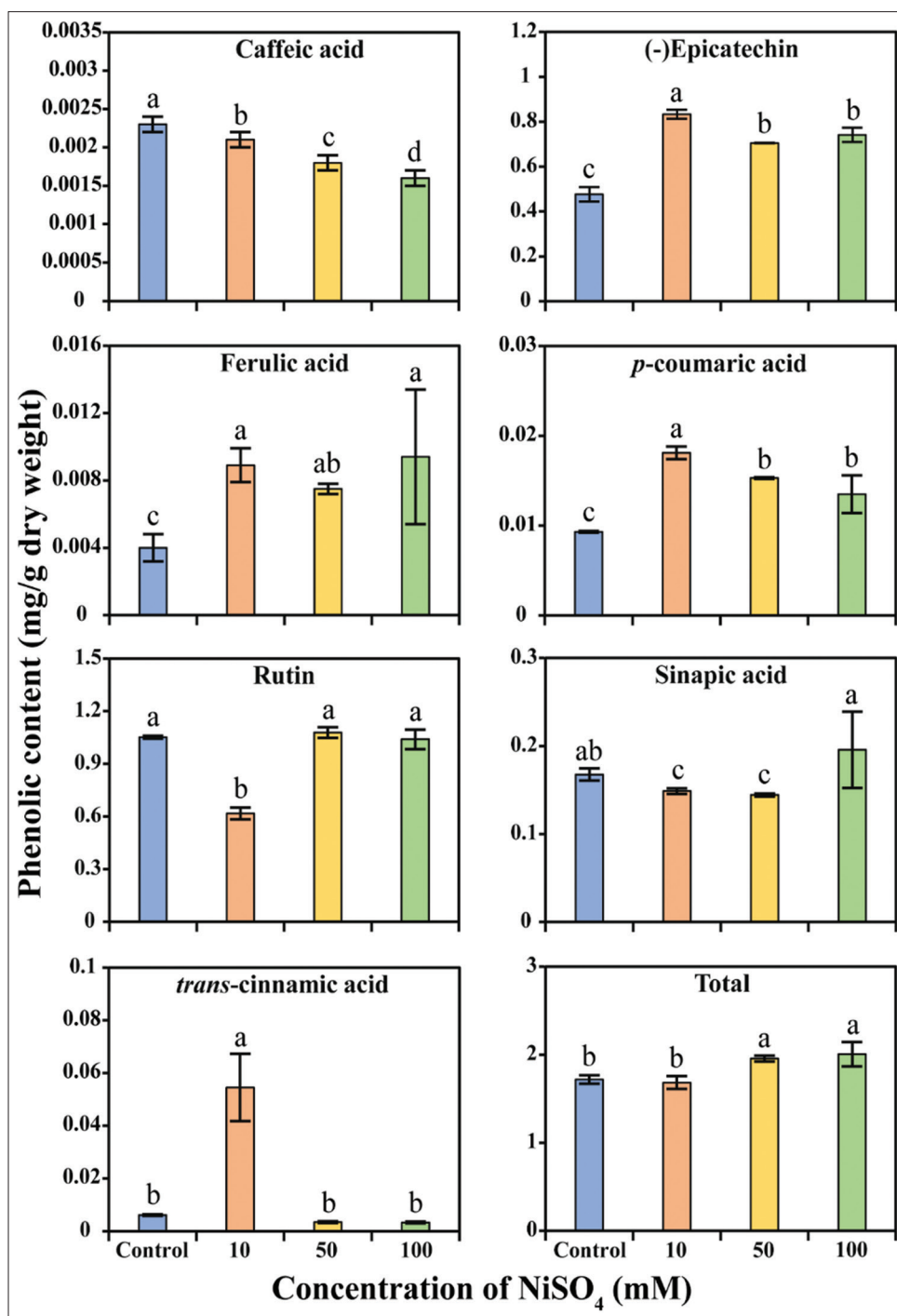
According to our findings, the amount of phenylpropanoid compounds varied depending on the heavy metal employed. In this study, we examined some of the specific phenolic compounds. Among the phenylpropanoid compounds, the



**Figure 2:** Effect of different concentrations of  $\text{HgCl}_2$  on phenylpropanoid compounds in *Euonymus alatus*

highest accumulation was noticed for rutin followed by epicatechin, sinapic acid, p-coumaric acid, trans-cinnamic acid, ferulic acid, and caffeic acid in all the exposed heavy metals. However, their concentration varied due to the variation of heavy metals and their concentration. This result was consistent with the previous study that the exposure of *Robinia pseudoacacia* to various heavy metals, the result showed that the concentration of the phenylpropanoid varied depending on the concentration of the heavy metals (Kim *et al.*, 2020). According to earlier research, the phenylpropanoid molecules mentioned above have potent antioxidant properties (Mehra *et al.*, 2013; Enogieru *et al.*, 2018; Grzesik *et al.*, 2018; Shen

*et al.*, 2019; Xu *et al.*, 2019), and exposure of plants to heavy metals boosted their accumulation. This indicates a relationship between the concentration of heavy metals in the plant's organs and the accumulation of different phenylpropanoid chemicals (Kisa *et al.*, 2016). It was noticed that comparatively the use of different concentrations of  $\text{CuCl}_2$  showed the maximum accumulation of total phenylpropanoid biosynthesis than  $\text{HgCl}_2$  and  $\text{NiSO}_4$ . A similar result was obtained in the previous study that *R. pseudoacacia* was exposed to various heavy metals, the  $\text{CuCl}_2$  exposure showed the highest accumulation of phenylpropanoid compounds (Kim *et al.*, 2020). In this study, the maximum amount of phenylpropanoid was found



**Figure 3:** Effect of different concentrations of NiSO<sub>4</sub> on phenylpropanoid biosynthesis in *Euonymus alatus*

in 0.1 mM CuCl<sub>2</sub>, however, when *E. alatus* was exposed to 1 mM HgCl<sub>2</sub>, the amount of phenylpropanoid decreased. In fact, with the use of HgCl<sub>2</sub>, the accumulation of phenylpropanoid decreased compared to control. This finding suggests that a decline in important enzyme activity associated with the phenylpropanoid biosynthesis pathway may be the cause of the drop in phenylpropanoid concentrations (Chung *et al.*, 2006; Kisa *et al.*, 2016). This suggested that a significant amount of different phenylpropanoid compounds accumulated in *E. alatus* to shield them against a variety of heavy metal stressors.

## CONCLUSION

The present investigation revealed that the plant's total phenylpropanoid compound levels were progressively elevated in *E. alatus* upon exposure to heavy metals. In all the heavy metal exposure the highest individual phenylpropanoid content was achieved in rutin and epicatechin, which leads to the highest accumulation of total phenolic content. In light of this, the synthesis of phenylpropanoid chemicals may help to lessen the formation of ROS generated by stress brought on by heavy

metals and let *E. alatus* thrive in heavily polluted environments. This partially elucidates why *E. alatus* is tolerant to various heavy metals, making it a useful species for the bioremediation of heavy metals from the environment. This helps to explain that *E. alatus* is a valuable species for the bioremediation of heavy metals from the environment due to its tolerance to different heavy metals.

## ACKNOWLEDGMENTS

This study was carried out with the support of ‘R&D Program for Forest Science Technology (Project No. 2021379B10-2123-BD02)’ provided by Korea Forest Service (Korea Forestry Promotion Institute).

## AUTHOR’S CONTRIBUTION

J.K.K. and S.U.P. designed the experiments. R.S., H.-H.K., B.V.N., J.Y. and B.B.P. performed the experiments and analyzed the data. R.S. wrote the manuscript. J.K.K. and S.U.P. revised the manuscript. All authors read and approved the final manuscript.

## REFERENCES

- Biala, W., & Jasinski, M. (2018). The phenylpropanoid case - It is transport that matters. *Frontiers in Plant Science*, *9*, 1610. <https://doi.org/10.3389/fpls.2018.01610>
- Cevallos-Casals, B. A., & Cisneros-Zevallos, L. (2010). Impact of germination on phenolic content and antioxidant activity of 13 edible seed species. *Food Chemistry*, *119*(4), 1485-1490. <https://doi.org/10.1016/j.foodchem.2009.09.030>
- Chung, I. M., Kim, J. J., Lim, J. D., Yu, C. Y., Kim, S. H., & Hahn, S. J. (2006). Comparison of resveratrol, SOD activity, phenolic compounds and free amino acids in *Rehmannia glutinosa* under temperature and water stress. *Environmental and Experimental Botany*, *56*(1), 44-53. <https://doi.org/10.1016/j.enxpb.2005.01.001>
- Clemens, S., & Weber, M. (2016). The essential role of coumarin secretion for Fe acquisition from alkaline soil. *Plant Signaling & Behavior*, *11*(2), e1114197. <https://doi.org/10.1080/15592324.2015.1114197>
- Demirezen, D., & Aksoy, A. (2006). Heavy metal levels in vegetables in Turkey are within safe limits for Cu, Zn, Ni and exceeded for Cd and Pb. *Journal of Food Quality*, *29*(3), 252-265. <https://doi.org/10.1111/j.1745-4557.2006.00072.x>
- Enogieru, A. B., Haylett, W., Hiss, D. C., Bardien, S., & Ekpo, O. E. (2018). Rutin as a potent antioxidant: Implications for neurodegenerative disorders. *Oxidative Medicine and Cell Longevity*, *2018*, 6241017. <https://doi.org/10.1155/2018/6241017>
- Fan, L., Zhang, C., Ai, L., Wang, L., Li, L., Fan, W., Li, R., He, L., Wu, C., & Huang, Y. (2020). Traditional uses, botany, phytochemistry, pharmacology, separation and analysis technologies of *Euonymus alatus* (Thunb.) Siebold: a comprehensive review. *Journal of Ethnopharmacology*, *259*, 112942. <https://doi.org/10.1016/j.jep.2020.112942>
- Gzesik, M., Naparło, K., Bartosz, G., & Sadowska-Bartos, I. (2018). Antioxidant properties of catechins: Comparison with other antioxidants. *Food Chemistry*, *241*, 480-492. <https://doi.org/10.1016/j.foodchem.2017.08.117>
- He, S.-Y., Qiu, X.-M., Wang, Y.-Q., Su, Z.-Q., Zhang, B.-Y., Wen, Z., Yang, Y.-F., Xing, B.-F., Hong, M., & Liao, R. (2022). Intervention effect of *Potentilla discolor*-*Euonymus alatus* on intestinal flora of type 2 diabetes mellitus rats. *European Review for Medical and Pharmacological Science*, *26*(24), 9062-9071. [https://doi.org/10.26355/eurrev\\_202212\\_30655](https://doi.org/10.26355/eurrev_202212_30655)
- Kim, N. S., Sathasivam, R., Chun, S. W., Youn, W. B., Park, S. U., & Park, B. B. (2020). Biosynthesis of phenylpropanoids and their protective effect against heavy metals in nitrogen-fixing black locust (*Robinia pseudoacacia*). *Tropical Journal of Pharmaceutical Research*, *19*(5), 1065-1072. <https://doi.org/10.4314/tjpr.v19i5.23>
- Kisa, D., Elmastas, M., Ozturk, L., & Kayir, O. (2016). Responses of the phenolic compounds of *Zea mays* under heavy metal stress. *Applied Biological Chemistry*, *59*, 813-820. <https://doi.org/10.1007/s13765-016-0229-9>
- Kivaisi, A. K. (2001) The potential for constructed wetlands for wastewater treatment and reuse in developing countries: a review. *Ecological Engineering*, *16*(4), 545-560. [https://doi.org/10.1016/S0925-8574\(00\)00113-0](https://doi.org/10.1016/S0925-8574(00)00113-0)
- Korkina, L., Kostyuk, V., De Luca, C. & Pastore, S. (2011). Plant phenylpropanoids as emerging anti-inflammatory agents. *Mini-Reviews in Medicinal Chemistry*, *11*(10), 823-835. <https://doi.org/10.2174/138955711796575489>
- Lombardi, L., & Sebastiani, L. (2005). Copper toxicity in *Prunus cerasifera*: growth and antioxidant enzymes responses of *in vitro* grown plants. *Plant Science*, *168*(3), 797-802. <https://doi.org/10.1016/j.plantsci.2004.10.012>
- Madera-Parra, C. A., Pena-Salamanca, E. J., Pena, M. R., Rousseau, D. P. L., & Lens, P. N. L. (2015). Phytoremediation of Landfill Leachate with *Colocasia esculenta*, *Gynerum sagittatum* and *Heliconia psittacorum* in Constructed Wetlands. *International Journal of Phytoremediation*, *17*(1), 16-24. <https://doi.org/10.1080/15226514.2013.828014>
- Mehra, P., Garg, M., Koul, A., & Bansal, D. D. (2013). Effect of (+)-catechin hydrate on oxidative stress induced by high sucrose and high fat diet in male Wistar rats. *Indian Journal of Experimental Biology*, *51*(10), 823-827.
- Ning, K., Zhou, T., Wang, Y., El-Kassaby, Y. A., & Ma, Y. (2022). The complete chloroplast genome of *Euonymus alatus* (Celastraceae). *Mitochondrial DNA Part B Resources*, *7*(4), 707-708. <https://doi.org/10.1080/23802359.2022.2067494>
- Panda, P., Appalashetti, M., & Judeh, Z. M. A. (2011). Phenylpropanoid sucrose esters: Plant-derived natural products as potential leads for new therapeutics. *Current Medicinal Chemistry*, *18*(21), 3234-3251. <https://doi.org/10.2174/092986711796391589>
- Qin, X. Y., Yan, S. X., & Wei, F. Y. (2011). Geographical distribution of plants of Celastraceae in China. *Journal of Northeast Forestry University*, *39*(1), 120-123.
- Shen, Y. B., Song, X., Li, L., Sun, J., Jaiswal, Y., Huang, J. Q., Liu, C., Yang, W. J., Williams, L., Zhang, H., & Guan, Y. (2019). Protective effects of *p*-coumaric acid against oxidant and hyperlipidemia-an *in vitro* and *in vivo* evaluation. *Biomedicine and Pharmacotherapy*, *111*, 579-587. <https://doi.org/10.1016/j.biopha.2018.12.074>
- Xu, D., Hu, M.-J., Wang, Y.-Q., & Cui, Y.-L. (2019). Antioxidant activities of quercetin and its complexes for medicinal application. *Molecules*, *24*(6), 1123. <https://doi.org/10.3390/molecules24061123>
- Yang, C. S., Landau, J. M., Huang, M.-T., & Newmark, H. L. (2001). Inhibition of carcinogenesis by dietary polyphenolic compounds. *Annual Review of Nutrition*, *21*, 381-406. <https://doi.org/10.1146/annurev.nutr.21.1.381>
- Yao, L. H., Jiang, Y. M., Shi, J., Tomas-Barberan, F. A., Datta, N., Singanusong, R., & Chen, S. S. (2004). Flavonoids in food and their health benefits. *Plant Foods for Human Nutrition*, *59*, 113-122. <https://doi.org/10.1007/s11130-004-0049-7>
- Zhai, X., Lenon, G. B., Xue, C. C. L., & Li, C.-G. (2016). *Euonymus alatus*: A review on its phytochemistry and antidiabetic activity. *Evidence Based Complementary and Alternative Medicine*, *2016*, 9425714. <https://doi.org/10.1155/2016/9425714>
- Zhao, J., Davis, L. C., & Verpoorte, R. (2005). Elicitor signal transduction leading to production of plant secondary metabolites. *Biotechnology Advances*, *23*(4), 283-333. <https://doi.org/10.1016/j.biotechadv.2005.01.003>