



Morphological and phytochemical studies of *Suaeda maritima* (L.) Dumort growing along the coastal belt of Purba Medinipur District, West Bengal, India in search of the prospective variation

Maniklal Pati¹, Asis Kumar Nandi²*

¹Department of Botany, Egra SSB College, Egra, Purba Medinipur, West Bengal – 721 429, India, ²Department of Botany and Forestry, Vidyasagar University, Medinipur, West Bengal – 721 102, India

ABSTRACT

Received: September 18, 2021 Revised: April 25, 2022 Accepted: April 27, 2022 Published: May 28, 2022

*Corresponding Author: Asis Kumar Nandi E-mail: aknindresearch@gmail. com *Suaeda maritima* (L.) Dumort of the family Chenopodiaceae is an annual succulent mangrove herb. This annual salt marsh is quite regularly used by the local people for food and pharmaceutical. This species has been cursorily noted to have variation in some morphological characters. Earlier reports indicated the presence of triterpenoid e.g. alpha amyrin in some species of *Suaeda*. However, no report on the variation in the quantity of it in this species was presented. The present study has furnished an account of subtle variation in morphology of this herb growing on different sites in the area under study. It also shows a difference in the amount of alpha amyrin in the plant individuals of different places, revealed through the HPTLC study. Morphological variations have been noted mostly in respect of the characteristics of the stem and leaf of the species.

KEYWORDS: Suaeda maritima, Morphological diversity, Alpha amyrin, Chromatographic analysis

INTRODUCTION

Mangrove vegetations grow along with the coastal belts of tropical and sub-tropical regions, usually between 25° N and 25° S latitude throughout the world (Tomlinson, 1986). Annual succulent herbs of the species of Suaeda grow naturally in soils having a high concentration of salt of mangroves (Untawale, 1984). Suaeda maritima grows luxuriantly along the coastal belt of Purba Medinipur district of West Bengal in India, right from Hijli-Sarif of Khejuri to Udaypur of Digha (Das, 2015). This range of coast in Purba Medinipur district is lying between 21° 51´27´´N to 21°36´5´´N latitude and 87°29´88´´E to 88°12′40′′E longitude. There is a record of its use as a vegetable and also in curing malady (Trease & Evans, 2002) and such uses are also noticed among the local people of the area under study. Though this herb grows almost continuously along the entire stretch of the region mentioned here, shows subtle variation in gross morphology along the site of its growing. Such diversity might be more due to the variation in the chemical and physical

properties of the soil along the region, rather than the genetic property of the plants for this contiguously and naturally growing herb. Such variation might also have a bearing on the biochemical constitution of them, too. Early literatures recount several different phytochemical compounds like triterpenoid, sterols, alkaloids, acids, glycosides (Krishchenko et al., 1984; Kapadia et al., 1985; Miftakhova et al., 1999), proteins and amino acids (Marie, 1965) to occur in this species. Alpha amyrin, a pentacyclic triterpenoid, a biomolecule of worth, has earlier been reported to occur in this species (Ghosh et al., 1985). α and β amyrins are two structural isomers possessing a wide spectrum of pharmaceutical and biological functions like, antimicrobial, insecticidal (Bandeira et al., 2006; Ekalu et al., 2019), anti-arthritic, anti-inflamatory, anti-nociceptive, anti-depressant, anti-hyperglycemic (Siani et al., 1999; Oliveira et al., 2004a; Oliveira et al., 2005b; Aragao et al., 2006; Aragao et al., 2007; Holanda et al., 2008; Melo et al., 2010; Barros et al., 2011; Melo et al., 2011; Santos et al., 2012; Aragao et al., 2015; Carvalho et al., 2017; Pinto et al., 2017), anti-ulcer, gastroprotective

Copyright: © The authors. This article is open access and licensed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0/) which permits unrestricted, use, distribution and reproduction in any medium, or format for any purpose, even commercially provided the work is properly cited. Attribution — You must give appropriate credit, provide a link to the license, and indicate if changes were made.

(Oliveira *et al.*, 2004b; Oliveira *et al.*, 2005a; Prabhakar *et al.*, 2017). Any variation in the amount of this secondary metabolite occurring between different plant individuals of this species would provide scope for selection of better producer among them, aspect of economic significance. With this matter as an objective, the present study has scrutinized and revealed subtle morphological and biochemical diversities amongst the individuals of the species growing in the coastal area of West Bengal.

MATERIALS AND METHODS

Collection and Identification of Plant Material

Aerial parts of *Suaeda maritime*, collected from the coast of Bay of Bengal in Purba Medinipur district of the state of West Bengal, India, were taken as study material. The plant samples collected from Bankiput has been designated as FSB and that from Sankarpur as FSS in this literature. Plant samples were collected at the end of monsoon from Sankarpur (21°61 '97 ' N, 87°57 '22 ' E) and Bankiput (21°76 '67 ' N, 87°86 '65 ' E) situated 41.5 km apart, along the coastal belt of Purba Medinipur district, at their flowering and fruiting stage. Herbarium was prepared with the collected sample according to Jain and Rao (1977) and Brummit and Powell (1992). The herbarium was identified and authenticated by the Central National Herbarium (CAL) of Botanical Survey of India (voucher specimen number CNH//2015/Tech.II/17/299).

Morphological Studies

Morphological studies were carried out with freshly collected whole plants. The studies included habits of the plant, leaf, stem, root, flower, fruit and seed. Along with gross morphological study with bare eyes, different plant parts like leaf, flower, fruit and seed were studied and measured under a microscope.

Biochemical Studies

For the purpose of biochemical study the aerial parts of the plant were chopped into pieces and dried in shade at room temperature and were pounded to powder in a mechanical grinder. Twenty grams of the dried aerial plant part from each sample was weighed and poured into 150 mL of methanol solvent. The mixture was stirred every 30 min for 3 h and thereafter kept for two days (Ali et al., 2001; Mammen et al., 2010; Reich & Schibli, 2011). The extracts of the plants from two sites of the collection were filtered separately using Whatman No 1 filter paper at 25°C temperature. The extracts obtained, thus, were concentrated to one third of their original volume by placing in a rotary evaporator. The concentrates were transferred into reagent bottles and stored in a refrigerator for HPTLC analysis.0.50 mL of this solution was diluted up to 10.0 mL by methanol to obtain the working standard solution of alpha-amyrin (Stahl, 1969). Standard alpha-amyrin (purity 99.3%), from Sigma-Aldrich Chemie GmbH (Aldrich Division, Steinbeim, Germany) was used as reference. A stock solution of this standard chemical 1000 µg/mL was prepared.

High Performance Liquid Chromatography Analysis

Chromatography was performed on $20 \,\mathrm{cm} \times 10 \,\mathrm{cm}$ TLC aluminum precoated silica gel $60F_{254}$ plate, with $200\,\mu m$ layer thickness (E. Merck, Mumbai, India). Standard and sample solutions were applied to the plates as 8 mm bands, 13 mm apart from each other and the Plate dimension 20 X 10 cm and 10 mm from the bottom edge of the plate, under a continuous spray of inert gas by means of a Camag Linomat V TLC sample applicator with a 100 µL syringe (Hamilton, Bonaduz, Switzerland). After the application, prederivatization was performed by exposing the plate to iodine vapor for 10 minutes. The prederivatized plate was developed vertically ascending in a twin-trough glass chamber (Camag, Switzerland) saturated with a mobile phase comprising petroleum ether: ethyl acetate: acetonitrile (8.2:1.2:0.1 v/v/v) (Stahl, 1969; Ghosh et al., 1985; Kapadia et al., 1985). The optimized chamber saturation time for the mobile phase was 20 minutes at room temperature. The chromatographic run length was 90 mm from the bottom edge of the plate. After development, the plate was air dried and derivatized by dipping the developed plate in anisaldehydesulphuric acid reagent for 2 seconds (Stahl, 1969; Ghosh et al., 1985; Martelanc et al., 2009). The plate was then airdried for complete removal of anisaldehyde-sulphuric acid and heated at 110°C for 3 minutes. Densitometric scanning was then performed at $= 580 \,\mathrm{nm}$ for alpha-amyrin using Camag TLC scanner 4 with winCATS software version 1.4.6. The slit dimension used was $5.00 \times 0.45 \text{ mm}$ (micro) with a scanning speed of 20 mm/sec, throughout the analysis (Stahl, 1969; Martelanc et al., 2007; Kpoviessi et al., 2008). The applied chromatographic conditions permitted a well separation of the triterpenoid -alpha amyrin from the plant extract of two samples without any decomposition of alpha amyrin.

Linearity

Determination of linear dynamic range concentration of alpha amyrin was done by applying 5μ L and 6μ L of TLC plate of working standard containing alpha amyrin. The peak area obtained from densitogram for each applied concentration of alpha amyrin was noted. The calibration curves of the standard were obtained by plotting graphs of the mean peak area of the standard versus the corresponding concentration.

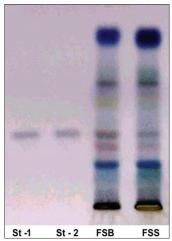
System Suitability

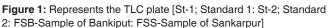
The chromatographic separation was performed by injecting 5μ L and 6 μ L standard solution of alpha amyrin on a TLC plate in two replicates under specified chromatographic conditions (Table-1). The values of percent relative standard deviations of peak area from the chromatogram and retention factor of standards were taken as an indicator of system suitability. The value of the retention factor for standard alpha-amyrin was 0.49. As the values of percent relative standard deviation of peak area were found to be less than 2

and peaks were well-resolved, the method was considered worthy for analysis.

Specificity

The specificity of the HPTLC method was ascertained by comparing visible chromatograms of alpha-amyrin standard with the chromatogram of two samples. The chromatograms were compared by the overlay. A good correlation was observed between chromatograms obtained from standard and samples (Figure 1).





Assay

The developed and validated HPTLC method was used for quantification of alpha-amyrin from the extract of dried whole plant powder of *S. maritima*. $20\,\mu$ L of each extract of plant materials of two zones was applied on the same TLC plate. The plate was developed and scanned under the specified chromatographic conditions.

Estimation of Alpha Amyrin

The amounts of alpha-amyrin present in each sample solution were determined from the calibration curve, by using the peak areas of alpha-amyrin in the sample. The flow rate of 150 nL/s was used.

RESULT AND DISCUSSION

Morphology

The general account of the annual herbaceous S. *maritima*, showed it to be quite bushy in nature with profuse branches (Figure 2A-C) and erect stem; stem reddish purple, glabrous, tender at younger parts and woody in the older region, internodes solid; leaves simple, alternate, sessile, exstipulate, linear, fleshy, semiterete, succulent and entire (Figure 2D). The inflorescence is a terminal spike and spike at leaf axil; flower is minute, ebracteate, bisexual, complete, regular, sessile, hypogynous, whitish green (Figure 2D); perianth 5, polytepalous, short,



Figure 2: A-C S. maritima plant; D-Flowering twig; E-Carpel; F-Fruit; G-Seed

Table 1: HPTLC performance of standard α amyrin

| Track | Sample | Applied volume | Start Rf | Start Height | Max Rf | Max Height | End Height | Area | Area % | Amount (mg/gm) | Sample ID |
|-------|----------|----------------|----------|--------------|--------|------------|------------|--------|--------|----------------|--------------|
| 1 | Standard | 5µl | 0.46 | 9.7 | 0.51 | 153.3 | 0.8 | 4197.9 | 48.55 | 0.05 | Alpha amyrin |
| 2 | Standard | 6μΙ | 0.44 | 11.3 | 0.49 | 177.8 | 6.3 | 5079.6 | 50.38 | 0.06 | Alpha amyrin |

Table 2: Morphological traits of two population of S. maritima

| Regions | FSB | FSS | | |
|--------------------------|----------------|----------------|--|--|
| Morphological characters | | | | |
| Plant height (mm) | 583±10.21 | 862±11.64 | | |
| Stem circumference (mm) | 44±2.37 | 62±2.41 | | |
| Length of internode (mm) | 83±3.89 | 118±5.2 | | |
| Leaf length (mm) | 19±1.58 | 25±2.1 | | |
| Leaf breadth (mm) | 2±0.17 | 3±0.33 | | |
| Leaf thickness (mm) | 0.8±0.07 | 1±0.13 | | |
| Flower length (mm) | 1±0.27 | 1.5 ± 0.36 | | |
| Tepal length (mm) | 0.7±0.05 | 0.9±0.06 | | |
| Fruit length (mm) | 1.3 ± 0.38 | 1.5 ± 0.47 | | |
| Fruit breadth (mm) | $0.9 \pm 0.$ | 1±0.16 | | |

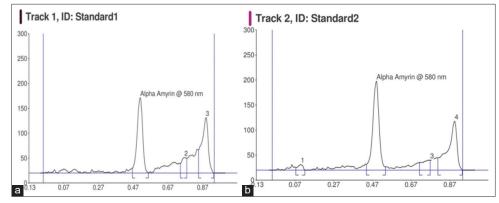


Figure 3: Represent HPTLC chromatogram of standard obtained at = 580 nm

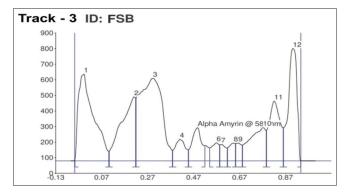


Figure 4: Represents HPTLC chromatogram of sample *S. maritima* at Bankiput obtained at = 580 nm

globose or urceolate; stamens 5, free, filaments short, white, anther rather large, basifixed, bilobed; carpels 3, syncarpous, ovary superior, sessile, wide based, adnate below to perianth, ovoid, rounded apex, style minute, stigma 2, recurved (Figure 2E); fruit small, utricle with persistent perianth (Figure 2F); seeds with coriaceous testa and slender embryo (Figure 2G).

Variation in Morphology

Plants collected from two zones did not show any striking morphological variations though subtle variations plant height, stem circumference, leaf length, breadth, thickness, flower and tepal length and fruit length and breadth were noted (Table 2).

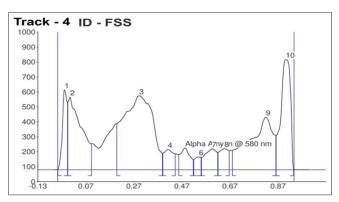


Figure 5: Represents HPTLC chromatogram of sample *S. maritima* at Sankarpur obtained at = 580 nm

Amount of Alpha Amyrin

The confirmation of the method of measuring the amount of alpha amyrin with respect to standard has been displayed in the Table 1 and Figure 3. The amount of amyrin has been found to be greater in the plants of Bankiput (0.12 mg/g i.e. 4.37%) than those of Sankarpur (0.068 mg/g i.e. 3.123%) (Table 3, Figure 4 & 5).

CONCLUSION

The occurrence of subtle morphological changes along with the considerable difference in the amount of alpha amyrin

| Table 3: HPTLC performance o | f $lpha$ amyrin from two p | opulations of <i>S. maritima</i> |
|------------------------------|----------------------------|----------------------------------|
|------------------------------|----------------------------|----------------------------------|

| Track | Sample | Applied volume | Start Rf | Start Height | Max Rf | Max Height | End height | Area | Area % | Alpha amyrin mg/gm |
|-------|--------|----------------|----------|--------------|--------|------------|------------|--------|--------|--------------------|
| 3 | FSB | 20µl | 0.45 | 72.1 | 0.49 | 213.3 | 98.4 | 7752.0 | 4.37 | 0.12 |
| 4 | FSS | 20µl | 0.45 | 100.9 | 0.48 | 146.6 | 70.6 | 5167.6 | 3.13 | 0.068 |

content among the individuals of *Suaeda maritima* growing at two distantly placed sites along the coastal area of West Bengal represents intraspecific diversity of it. Though the sites of their occurrence are contiguous and the individuals of the species are growing interruptedly in patches, the diversity, as witnessed, might be due to the difference in micro-environmental factors rather than the difference in their genetic content.

ACKNOWLEDGEMENT

The authors wish to thank RKM Quality Testing Laboratory, Narendrapur, Kolkata, India, for providing the laboratory facility. The authors are also grateful to the Director of Central National Herbarium, Howrah, India for the identification and authentication of the species and special thanks go to the local people for their kind co-operation during sample collection.

REFERENCES

- Ali, M., Ali, S. N., & Ramachandram, R. (2001). Phytochemical investigation of aerial parts of *Pluchea lanceolata* C. B. Clarke. *Indian Journal of Chemistry*, 40(8), 698–706.
- Aragao, G. F., Carneiro, L. M., & Junior, A. P. (2006). A possible mechanism for anxiolytic and antidepressant effects of alpha-and betaamyrin from *Protium heptaphyllum* (Aubl.) March. *Pharmacology Biochemistry and Behavior*, 85, 827–834. https://doi.org/10.1016/j. pbb.2006.11.019
- Aragao, G. F., Cunha, P. M. C., & Nogueira, B. P. (2007). Analgesic and antiinflammatory activities of the isomeric mixture of alpha- and betaamyrin from *Protium heptaphyllum* (Aubl.) March. *Journal of Herbal Pharmacotherapy*, 7(2), 31-47. https://doi.org/10.1300/j157v07n02_03
- Aragao, G. F., Carneiro, L. M., & Rota-Junior, A. P. (2015). Alterations in brain amino acid metabolism and inhibitory effects on PK are possibly correlated with anticonvulsant effects of the isomeric mixture alpha and beta-amyrin from *Protium heptaphyllum*. *Pharmaceutical Biology*, 53(3), 407–413. https://doi.org/10.3109/13880209.2014.923001
- Bandeira, P. N., Fonseca, A. M., & Costa, S.M. (2006). Anti-bacterialand antioxidant activities of the essential oil of resin of *Protium heptaphyllum*. *Natural Product Communications*, 1(2), 117–200.
- Barros, F. W., Bandeira, P. N., Lima, D. J., Meira, A. S., de Farias, S. S., Albuquerque, M. R., dos Santos, H. S., Lemos, T. L., de Morais, M. O., Costa-Lotufo, L. V., & Pessoa, C. (2011). Amyrin esters induce cell death by apoptosis in HL-60 leukemia cells. *Bioorganic & Medicinal Chemistry*, *19*(3), 1268–1276. https://doi.org/10.1016/j. bmc.2010.12.016
- Brummit, R. K. & Powell, C. E. (1992). *Author of Plant Names*. Kew: Royal Botanical Garden.
- Carvalho, K. M., de Melo, T. S., de Melo, K. M., Quinderé, A. L., de Oliveira, F. T., Viana, A. F., Nunes, P. I., Quetz, J. D., Viana, D. A., da Silva, A. A., Havt, A., Fonseca, S. G., Chaves, M. H., Rao, V. S., & Santos, F. A. (2017). Amyrins from *Protium heptaphyllum* reduce high-fat dietinduced obesity in mice via modulation of enzymatic, hormonal and inflammatory responses. *Planta Medica*, *83*(3-04), 285–291. https:// doi.org/10.1055/s-0042-114222
- Das, D. C. (2015). Study of the tidal vegetation of Purba Medinipur district of West Bengal, India. *International Journal of Bioassays*, 4(05), 3915-3921.
- Ekalu, A., Ayo, R. G., Habila, J. D., & Hamisu, I. (2019). Bioactivities of phaeophytin a, α-amyrin, and lupeol from *Brachystelma togoense* Schltr. *Journal of The Turkish Chemical Society*, 6(3), 411-418. https:// doi.org/10.18596/jotcsa.571770

- Ghosh, A., Misra, S., Dutta, A. K., & Choudhury, A. (1985). Pentacyclic tri-terpenoids and sterols from seven species of mangrove. *Phytochemistyry*, 24(8), 1725 -1727. https://doi.org/10.1016/S0031-9422(00)82541-8
- Holanda, P. S. A., Pinto, L. M. S., & Cunha, G. M. A. (2008). Antiinflammatory effect of alpha, beta-amyrin, a pentacyclic triterpene from *Protium heptaphyllum* in rat model of acute periodontitis. *Inflammopharmacology*, *16*(1), 48–52. https://doi.org/10.1007/ s10787-007-1609-x
- Jain, S. K., & Rao, R. R. (1977). *A Handbook of field and Herbarium Methods.* New Delhi, India: Today and tomorrow's Printers and Publishers.
- Kapadia, Z., Hussian, N., & Badar, Y. (1985). Chemical investigation of Suaeda nudiflora Willd. Karachi Journal of Science, 13(2), 113-18.
- Kpoviessi, D. S. S., Gbaguidi, F., & Gbenou, J. (2008). Validation of a method for the determination of sterols and triterpenes in the aerial part of *Justicia anselliana* (Nees) T. Anders by capillary gas chromatography. *Journal of Pharmaceutical and Biomedical Analysis*, 48(4), 1127–1135. https://doi.org/10.1016/j.jpba.2008.08.036
- Krishchenko, V. P., Rotar, A. I., Zadniprany, I. U. F., Kosorukov, K. L., Protor, U., Anofrina, N. D., & Timiryazevsk, I. Z. V. (1984). Chemical composition and nutritive value of plants from the family Chenopodiaceae of the pasture massif in Libya. *Izvestiia - Timiriazevskaia sel'skokhoziaistvennaia akademiia*, *4*, 38-45.
- Mammen, D., Daniel, M., & Sane, R.T. (2010). Seasonal and geographical variations in chemical constituents of *Leptadenia reticulate*. *International Journal of Pharmaceutical Sciences Review and Research*, 4(2), 111–116.
- Marie, G. (1965). Metabolism of nitrogen in halophytes. Amino acids and free amides in young *Suaeda macrocarps* plants harvested in their natural environment. *Comptes rendus de l'Académie des Sciences*, *261*(14), 2724-6.
- Martelanc, M., Vovk, I., & Simonovska, B. (2007). Determination of three major triterpenoids in epicuticular wax of cabbage (*Brassica oleracea* L.) by high-performance liquid chromatography with UV and mass spectrometric detection. *Journal of Chromatography* A, *1164*(1-2), 145–152. https://doi.org/10.1016/j.chroma.2007.06.062
- Martelanc, M., Vovk, I., & Simonovska, B. (2009). Separation and identification of some common isomeric plant triterpenoids by thinlayer chromatography and high-performance liquid chromatography. *Journal of Chromatography*, 1216(38), 6662–6670. https://doi. org/10.1016/j.chroma.2009.07.038
- Melo, C. M., Carvalho, K. M., & Neves, J. C. (2010). Alpha, beta-amyrin, a natural triterpenoid ameliorates L-arginine-induced acute pancreatitis in rats. *World Journal of Gastroenterology*, *16*(34), 4272–4280. https:// doi.org/10.3748/wjg.v16.i34.4272
- Melo, C. M., Morais, T. C., & Tome, A. R. (2011). Anti-inflammatory effect of α , β -amyrin, a triterpene from *Protium heptaphyllum* on ceruleininduced acute pancreatitis in mice. *Journal of Inflammation Research*, *60*(7), 673–681.
- Miftakhova, A. F., Burashera, G. Sh., & Abilov, Zh. A. (1999). The chemical composition of some Kazakhstan glassworts. *Chemistry of Natural Compounds*, 35(2), 225-226.
- Oliveira, F. A., Lima-Junior, R. C., & Cordeiro, W. M. (2004a). Pentacyclic triterpenoids, alpha, beta-amyrins, suppress the scratching behavior in a mouse model of pruritus. *Pharmacology Biochemistry and Behavior*, 78, 719–725. https://doi.org/10.1016/j.pbb.2004.05.013
- Oliveira, F. A., Vieira-Junior, G. M., & Chaves, M. H. (2004b). Gastroprotective effect of the mixture of alpha- and beta-amyrin from *Protium heptaphyllum*: role of capsaicin-sensitive primary afferent neurons. *Planta Medica*, 70, 780–782. https://doi.org/10.1055/s-2004-827212
- Oliveira, F. A., Chaves, M. H., & Almeida, F.R. (2005a). Protective effect of alpha- and beta-amyrin, a triterpene mixture from *Protium heptaphyllum* (Aubl.) March. trunk wood resin, against acetaminophen-induced liver injury in mice. *Journal of Ethnopharmacology*, *98*, 103–108. https:// doi.org/10.1016/j.jep.2005.01.036
- Oliveira, F. A., Costa, C. L., & Chaves, M. H. (2005b). Attenuation of capsaicin-induced acute and visceral nociceptive pain by alpha-and

beta-amyrin, a triterpene mixture isolated from *Protium heptaphyllum* resin in mice. Life Sciences, 77, 2942–2952. https://doi.org/10.1016/j. lfs.2005.05.031

- Pinto, S. A. H., Pinto, L. M. S., Cunha, G. M. A., Chaves, M. H., Santos, F. A., & Rao, V. S. (2005). Anti-inflammatory effect of α, β-amyrin, a pentacyclic triterpene from *Protium heptaphyllum* in rat model of acute periodontitis. *Inflammopharmacology*, *15*, 1–5. https://doi. org/10.1007/s10787-007-1609-x
- Prabhakar, P, Reeta, K. H., & Maulik, S. K. (2017). α-Amyrin attenuates high fructose diet-induced metabolic syndrome in rats. *Applied Physiology*, *Nutrition, and Metabolism*, 42, 23-32. https://doi.org/10.1139/apnm-2016-0088
- Reich, E., & Schibli, A. (2011). *High-Performance Thin-Layer Chromatography* for the Analysis of Medicinal Plants. (Ist Ed.). Thieme Medical.
- Santos, F. A., Frota, J. T. & Arruda, B. R. (2012). Anti-hyperglycemic and hypolipidemic effects of α, β-amyrin, a triterpenoid mixture from *Protium heptaphyllum* in mice. *Lipids in Health and Disease*, 11, 98.

https://doi.org/10.1186/1476-511X-11-98

- Siani, A. C., Ramos, M. F., Menezes-de-Lima, O., Jr, Ribeiro-dos-Santos, R., Fernadez-Ferreira, E., Soares, R. O., Rosas, E. C., Susunaga, G. S., Guimarães, A. C., Zoghbi, M. G., & Henriques, M. G. (1999). Evaluation of anti-inflammatory-related activity of essential oils from the leaves and resin of species of Protium. *Journal of Ethnopharmacology*, *66*(1), 57–69. https://doi.org/10.1016/s0378-8741(98)00148-2
- Stahl, E. (1969). *Thin Layer Chromatography Handbook*. (2nd Ed.). Springer-Verlage: Berlin; Heidelberg: New York.
- Tomlinson, P. B. (1986). *The Botany of Mangroves*. Cambridge, UK: Cambridge University Press.
- Trease, G. E., & Evans, W. C. (2002). *Pharmacognosy.* (15thEd.). University of Nottingham, Nottingham, UK.
- Untawale, A. G. (1984). Present status of mangroves along the west coast of India. Paper presented at the Asian Symposium of Mangrove Environment- Resources & Management, Jakarta.