

# **REGULAR ARTICLE**

# Phytoextraction of lead through *Helianthus annuus* and its effects on the growth of *Cajanus cajan*

# Vijay Kumar Sinhal<sup>1</sup>, Vipin Yadav<sup>1</sup> and Adarsh pandey<sup>2</sup>

1 Department of Botany, Maharaja Agrasen Mahavidyalaya Bareilly, UP

2 Department of Botany, S.S.(P.G.)College,Shajahanpur, UP

# **KEYWORDS**

A. galanga, Extractive values, Ash values

## CORRESPONDENCE

Vijay Kumar Sinhal, Department of Botany, Maharaja Agrasen Mahavidyalaya Bareilly, UP

E-mail:sinhal@rediffmail.com

EDITOR

Adarsh Pandey

CB Volume 2, Year 2011, Pages 01-07

#### ABSTRACT

To evaluate the effects of Pb (Lead Nitrate) on the growth of *Cajanus cajan* and it's remediation, the two varieties of *Cajanus cajan* (Var. UPAS-120 & Var. ICPL-151) were sown in two different set of fields. Both the set of field were irrigated with different doses of Pb (19, 20, & 21 mg lit<sup>-1</sup>). In the second set of experiment the same varieties of *Cajanus cajan* were intercropped with *Helianthus annuus* to remediate the Pb from soil. A concentration dependent decrease was noticed in all the growth parameters (i.e. plant height, fresh & dry Weight of plants, chlorophyll content, and protein content) and biochemical parameters (enzymes like Nitrate reductase, Glutamine oxo glutarate amino transferase and Glutamate dehydrogenase, activities) studied but the proline content was increased with increase in concentration of Pb. The maximum reduction (% of control) was noticed under the influence of higher dose of Pb (21 mg lit<sup>-1</sup>) in both the varieties of *Cajanus cajan*. Variety UPAS-120 was found to be more susceptible to Pb toxicity than variety ICPL – 151.

In second set of experiment (where *Cajanus cajan* was intercropped with *Helianthus annuus*) a significant recovery of the toxic effects of Pb was noticed in all the parameters studied. Atomic absorption spectrophotometer study suggested that the accumulation of Pb through *Helianthus annuus* grown in Pb treated soil was higher than non-treated soil. The results of the present investigation reveals that *Helianthus annuus* can be used as phytoremediator to reclaim the soil contaminated with Pb.

# Introduction

The pollution of the environment with toxic metals has become a world wide problem, affecting crop yields, soil biomass and fertility, contributing to bioaccumulation in the food chain. Consequently, the use of plants contaminated with high levels of heavy metals for food, might pose a serious risk to human and animal health (Wang et al. 2003). Pb is hazardous at higher concentration and inhibits growth of plants (Bergmann 1992, Ali et al. 1999, Soares et al. 2001, Bekiaroglou & Karatagli 2002, Khudsar et al. 2004). The elevated level of heavy metals reduces the activities of enzymes like catalase, superoxide dismutage, guaiacol peroxidase, protease, nitrogenase, nitrate reductase etc. (Antipchuk et al. 2000, Panda and Khan 2003, Chhetri et al. 2004, Singh et al. 2005, Sinhal V.K. 2005, Sinhal et al. 2006). Photosynthesis and transpiration are also reduced by high concentrations of Pb ( Van Assche et al. 1979) The observed reduction in growth is also a consequence of Pb interference with nutrient uptake (Chanay 1993, Kaya et al. 2000) and certain enzyme activities (Quariti et al. 1997).

Phytoremediation is a group of technologies that use plants to reduce, remove, degrade or immobilize environmental toxins, primarily those of anthropogenic origin, with the aim of restoring area sites to a condition usable for private or public applications. Phytoremediation can be ciassified into six different remediation methods: Phytoextraction, Phytostabilization, Phytodegradation, Phytovolatilization, Rhizofilterations, & Enhanced Rhizosphere Biodegradation. Among these methods, phytoextraction is a remediation method that uses plants to remove heavy metals from soil (Suresh & Ravishankar 2004, Solhi *et al.* 2005). Thus present investigation has been under taken to assess the phytotoxicity of Pb on the growth of C*ajanus cajan* and *Helianthus annuus* has been chosen as a phytoremediator plant to remediate Pb from treated soil.

#### Materials and Methods

The seeds of the two varieties i.e. var. UPAS-120 & var. ICPL - 151 of Cajanus cajan and Helianthus annuus (var. Akash-333) were obtained from seed centre, Haldi, Pantnagar, India. The field was divided into two parts; one was for the evaluation of the effects of Pb on the growth of Cajanus cajan and second for phytoremediation experiment. The first part of the field was consists of 8 small plots, each having an area of 9 m<sup>2</sup> (3mx3m), four plots for each variety. Plots were arranged as follows Control (irrigated with tap water), treatments A, B & C ( irrigated with 19,20,& 21 mg lit '1 concentration of Pb separately). The second part of the field was consist of 6 small plots, each having an area of 9m<sup>2</sup>, 3 plots for each variety of Cajanus cajan which were intercropped with Helianthus annuus i.e., these plots were arranged as D, E & F (having Cajanus cajan + Helianthus annuus treated with 19,20 & 21 mg lit<sup>-1</sup> concentration of Pb respectively). Prior to sowing, seeds were presoaked in distilled water for 8 hours to help in their easy from germination. The seeds were sown in June when the mean monthly temperature ranged 26°C (minimum) to 41°C (maximum) and were irrigated with different doses of Pb at 10, 15, 30 & 45 days of sowing.

To determine the effects of different doses of Pb (19, 20 & 21 mg lit<sup>1</sup>) on plant height, five plants were randomly selected from each treatment. The plants height was observed in 25 & 45 days old plants and was measured in centimeters. The fresh

weight was measured in grams. For dry weight, plants were dried in oven at 60°C for 48 hours and measured in gms. Chlorophyll content was measured by the method of Arnon (1949), Protein content by the method of Lowery *et al.* (1951), NR activity by the method of Srivastava (1974), GOGAT activity by the method of Meers *et al.* (1970), GDH activity by the method of Wolf & Wiliams (1973) and Proline content was determined by the method of Bates *et al.* (1973). The amount of Pb accumulated (mg gm<sup>-1</sup> dry wt.) by *Helianthus annuus* was determined by using Atomic Absorption Spectrophotometer (GBC Avanta S AAS, Australia) after samples were digested with concentrated HNO<sub>3</sub> + HCIO<sub>4</sub>.

One way ANOVA was carried out to compare the means of different treatments at 5% level of significance.

#### Results

The effects of different concentrations of Pb (19, 20 & 21 mg lit<sup>1</sup>) were noticed at 25 & 45 days of sowing on all the parameters studied. All the doses of Pb were found to have inhibitory effects on plant height (in cm.) but the maximum reduction was noticed in variety UPAS – 120 (32.30%) at highest

does of Pb (21 mg lit<sup>-1</sup>) in 45 days old plants. The maximum retardation i.e. 40.84% in fresh weight (in gms) and 43.15% in dry weight (in gms) was noticed in 45 days old plants in variety UPAS-120 with the highest does (21 mg lit<sup>-1</sup>) of Pb. A concentration dependent decrease was also noticed in case of chlorophyll content (mg g<sup>-1</sup> fresh weight), and the maximum reduction was noticed (43.28%) with higher does of Pb (21 mg lit 1) in variety UPAS-120. All the concentrations of Pb significantly decreased the Nitrate reductase (in NO2<sup>-</sup>hr<sup>-1</sup>g<sup>-1</sup> fresh weight) and Glutamine oxoglutarate amino transferase (µM NADPH oxi min-<sup>1</sup>g<sup>-1</sup> fresh weight) activity, though, the highest reduction of 39.14% and 51.27% respectively were observed with the highest dose of Pb (21 mg lit<sup>-1</sup>) in variety UPAS-120. Similarly Glutamate dehydrogenase activity (µM NADPH oxi min<sup>-1</sup>g<sup>-1</sup> fresh weight) was also reduced to 42.64% under the influence of highest dose of Pb (21 mg lit<sup>-1</sup>). All the concentrations of Pb also showed inhibitory effect on protein content (in mg  $g^{-1}$  dry weight) but the maximum reduction i.e., 47.30% was noticed with the higher dose of Pb (21 mg lit<sup>-1</sup>) in variety UPAS-120 in 45 days old plants. (Fig. 1&2).





- A = *Cajanus cajan* treated with 19 mg.lit<sup>-1</sup> Pb Nitrate
- $B = Cajanus \ cajan \ treated \ with \ 20 \ mg.lit^{-1} \ Pb \ Nitrate$
- $C = Cajanus \ cajan \ treated \ with \ 21 \ mg.lit^{\cdot 1}$  Pb Nitrate e
- D = Cajanus cajan + Helianthus annuus treated with 19 mg.lit<sup>-1</sup> Pb Nitrate
- E = Cajanus cajan + Helianthus annuus treated with 20 mg.lit<sup>1</sup> Pb Nitrate
- F = Cajanus cajan + Helianthus annuus treated with 21 mg.lit<sup>-1</sup> Pb Nitrate

Results with reverse trend were noticed in case of proline content ( $\mu$  moles g<sup>-1</sup> fresh weight) i.e., a concentration dependent increase was noticed in both the varieties in 25 and 45 days old plants. The maximum proline content (.189  $\mu$  moles g<sup>-1</sup> fr. wt.) was noticed with higher does of Pb (21 mg lit<sup>-1</sup>) in variety UPAS-120 in 45 days old plants. (Fig.-6)

The recovery of phytotoxic effects of different doses of Pb (19, 20 & 21 mg lit<sup>1</sup>) on *Cajanus cajan* was achieved through phytoremediation with *Helianthus annuus*. In case of variety UPAS-120, the maximum recovery (% to reduction) through

Helianthus annuus was noticed in plant height (53.23%), fresh weight of plants (73.39%), dry weight of plants (71.98%), chlorophyll content (68.88%), NR activity (73.08%), GOGAT activity (67.56%), GDH activity (72.42%) and protein content (77.15%) while in case of variety ICPL-151, the maximum recovery (% to reduction) was noticed in plant height (75.46%), fresh weight of plants (74.13%), dry weight of plants (75.41%), chlorophyll content (72.08%), NR activity (77.36%), GOGAT activity (75.10%), GDH activity (80.11%) and protein content (79.80%). (Fig. -3 & 4)



At 25 Days





# Fig.2: Effects of Pb on the growth of Cajanus cajan (ICPL-151) and its phytoremediation through Helianthus annuus

- A = Cajanus cajan treated with 19 mg.lit<sup>-1</sup> Pb Nitrate
- $\mathbf{B}$  = Cajanus cajan treated with 20 mg.lit  $^{_{1}}$  Pb Nitrate
- C = Cajanus cajan treated with 21 mg.lit<sup>-1</sup> Pb Nitrate
- D = Cajanus cajan + Helianthus annuus treated with 19 mg.lit<sup>-1</sup> Pb Nitrate
- $\mathbf{E}$  =  $Cajanus\,cajan$  +  $Helianthus\,annuus\, \mathrm{treated}$  with 20 mg.lit  $^1$  Pb Nitrate
- $\mathbf{F}$  = Cajanus cajan + Helianthus annuus treated with 21 mg.lit $^{_1}$  Pb Nitrate







Fig.3: Effects of Pb on the growth of *Cajanus cajan* (UPAS-120) showing maximum reduction (% to control) and maximum recovery (% to reduction)



At 25 Days



Fig.4: Effects of Pb on the growth of *Cajanus cajan* (ICPL-151) showing maximum reduction (% to control) and maximum recovery (% to reduction)





#### Fig.5: Comparison of amounts of Pb accumulated by different parts of Helianthus annuus (after 75 days of intercropping with Cajanus cajan)

Significant differences were obtained between concentrations of Pb in different parts of *Helianthus annuus* grown in contaminated soil as compared to its concentration in non-contaminated soil. The highest concentration of Pb was noticed in leaves (393.12 mg kg<sup>-1</sup> DW) as compared to stem (240.23 mg kg<sup>-1</sup> DW) and roots (80.15 mg kg<sup>-1</sup> DW) under the influence of highest dose of Pb (21 mg lit<sup>-1</sup>). (Fig.-5)

## Discussion

In the present investigation Pb was found to be phytotoxic in nature at all the concentrations (19, 20, & 21 mg lit

<sup>1</sup>) and significantly reduced the plant height of *Cajanus cajan*. Although Pb is essential for plant growth, its excess is inhibitive as was reported for a number of leguminous plants (Khudsar 1999), but at higher concentration it's interference with certain essential metabolic events of plant suggested that the inhibition of plant height under the influence of heavy metals is probably due to results of interference of these heavy metals with the respiratory activity and mobilization of reserves (Tripathi and Mohanty 1980, Alia *et al.* 1995, Jain *et al.* 2000, Khudsar 2004).



Fig. 6: Effects of Pb on proline content ( $\mu$  moles g<sup>1</sup> fresh weight of leaves) of leaves in *Cajanus cajan* 

Fresh and dry weight of plant was reduced significantly at every concentration of Pb (19, 20 & 21 mg lit<sup>-1</sup>) whereas the maximum reduction with the higher dose of Pb (21 mg lit<sup>-1</sup>) was noticed in variety UPAS – 120 as compared to variety ICPL-151. Decrease in fresh weight of plants was might be due heavy loss of moisture. The decrease in dry weight might be due to reduction in photosynthesis and chlorophyll 'a' synthesis as suggested by Okhi (1978), Joshi *et al* (1999), Sinhal (2005), Sinhal *et al* (2006).

A concentration dependent decrease was also noticed in chlorophyll content. Prasad & Prasad (1987) suggested that heavy metal stress inhibits chlorophyll biosynthesis by interacting with sulphydryl (-SH) group of the two enzymes viz., δ-Amino laevulinic acid (ALA) dehydratase and Protochlorophyllide reductase, involved in chlorophyll biosynthesis.

The reduction of nitrate to nitrite by the enzyme nitrate reductase (N.R.E.C. 1.6.6.1) is a rate limiting step in overall nitrate assimilation and also controls the rate of protein synthesis in plants (Srivastava, 1980). The NR activity also reduced significantly under the influence of Pb treatments. The reduction in NR activity with Pb treatments may be due to it's strong affinity for -N and -S ligands (Woolhouse, 1983) and therefore they can bind to the active component of the enzyme such as histidine and sulphydryl groups (-SH) and can eventually inhibit NR activity (Van Assche & Clijsters 1990). The cause of inhibition of NR activity by heavy metals may be multifacial e.g. due to reduced supply of NADPH, disorganization of chloroplasts, less NO<sub>3</sub> supply to the site of synthesis (Singh *et al.*, 1998). The Pb concentrations were also found to have inhibitory effects on NR activity by Khudsar *et al.* (2004).

Concentration dependent decrease was also found in GOGAT activity. GOGAT is the main enzyme of GS-GOGAT pathway of ammonium assimilation in which  $\rm NH_{4^+}$  and glutamate form glutamine which converts into glutamic acid in the presence of enzyme GOGAT. The reduction in the assimilation of ammonium ions by heavy metals in cucumber plant (Burzynski and Buczek, 1998) may be due to intereference of metal ions with the active site of the ammonium assimilating enzymes.

www.currentbotany.org ISSN: 2220-4822

Ammonia & 2-oxyoglutaric acid forms glutamic acid in the presence of enzyme GDH. The treatments of Pb significantly reduced GDH enzyme in both the varieties of Cajanus cajan. According to Miller et al. (1973) GDH enzyme contain free-SH (Sulphydryl) group at the active centre and it is well known fact that heavy metals show strong affinity for -SH group, therefore, Pb treatments (19, 20 & 21 mg lit<sup>-1</sup>) adversely affected GDH activity. According to Tomar et al. (2000), the protein content under heavy metal stress may be affected due to (i) Enhanced protein hydrolysis resulting in decreased concentration of soluble proteins (Melinchuk et al., 1982). (ii) Catalytic activity of heavy metals (Bhattacharya and Choudhari, 1997) and (iii) Protein Synthesis becoming reduced under all stress conditions. According to Chhetri et al. (2004) when heavy metal toxicity crosses the threshold, the protein level decreases and this might be due to the breakdown of protein synthesis mechanism at toxic concentration level of heavy metals or due to reduced incorporation of free amino acids into protein. Concentration of protein was significantly reduced in Artemisia annua leaves by Pb treatments (Khudsar et al., 2004).

Reverse trend of results was noticed in case of proline content viz., a concentration dependent increase was noticed in proline content. The higher proline content was observed with the higher dose of Pb (21 mg lit<sup>-1</sup>) in both the varieties of Cajanus cajan. The proline content accumulated in number of plants when they are subjected to salt or metal stress (Muthukumarsamy 1997). Proline has been shown to play an important role in ameliorating environmental stress in plants, including heavy metals stress (Surasak et al. 2002, Sharma & Dietz 2006). However, there has been much disagreement regarding the mechanism by which proline reduces heavy metal stress. Free proline has been proposed to act as an osmoprotectant (Delauney and Verma 1993, Taylor 1996), a protein stabilizer (Shan & Dubey, 1998), a metal chelator (Fargo & Muller, 1979), an inhibitor of lipid peroxidation (Mehta and Gaur, 1999) a hydroxyl radical scavenger (Smirnoff and Cumber, 1989). It is evident that there is no clear consensus regarding the mechanism by which proline reduces heavy metal stress.

The phytoremediation experiments, in which Helianthus annuus was intercropped with Cajanus cajan, were found to have a significant recovery of the reduction induced by different Pb treatments. Atomic Absorption spectrophotometer studies of Helianthus annuus reveals that a remarkable amount of Pb was accumulated by different parts of the Helianthus annuus but the highest accumulation was noticed in roots as compared to shoot and leaves. The accumulation of Pb through Helianthus annuus was also noticed by Herrero et al. (2003). Lesage et al. (2005), Solhi et al. (2005), Susan et al. (2006). Thus Helianthus annuus act as a powerful phytoremediator and hyper accumulator for Pb and makes the soil less toxic. Gratao et al. (2005) focused their attention on the physiological mechanism by which the metal is taken up, transported and sequestrated but they could know little about the genetic basis of hyper accumulation when compared with the genetic basis of metal tolerance (Pollard et al., 2002). Oven et al. (2002) pointed out that hyper accumulation and detoxification includes chelating to organic acids or proteins (Martens *et al* .1996).

The overall view of the present investigation is that higher dose of Pb (21 mg lit<sup>-1</sup>) exert higher toxic effects on plant growth and metabolism of *Cajanus cajan* which is further confirmed by presence of highest level of proline under the influence of higher dose of Pb (21 mg lit<sup>-1</sup>). Proline is an effective indicator of heavy metal stress. Further the recovery of the reduction in growth and biochemical parameters, induced by Pb, could be achieved by the process of phytoremediation (through *Helianthus annuus*). Thus *Helianthus annuus* can be used as phytoremediator plant to remediate the soil contaminated with Pb.

# References

- Ali, G., Srivastava, P.S. Iqbal, M.: Morphogenic and biochemical responses of *Bacopa monniera* cultures to Pb toxicity. - Plant Sci. **139**: 1-7, 1999.
- Alia, K. V. Prasad, S.K. Pradha Saradhi, P.: Effect of Pb on free radicals and proline in *Brassica juncea* and *Cajanus cajan.* -Phytochemistry. **39**: 45-47, 1995.
- Antipchuk, A.F. Rangelova, V.N., Tantsurenko, F.V.: Effects of heavy metals and reclaimerson formation and functioning of Legume – rhizobial symbiosis. – Mikrobiologi Chnii – Zhurnal. 62(6): 44-50, 2000.
- Arjunan, G., Samboorna Raman, S.: Pb deficiency in *Magnifera* indica and *Psidium guajora*. - Hindu. 117:165, 1994.
- Arnon, D.I.: Copper enzymes in isolated chloroplasts polyphenol oxidase in *Beta vulgaris*. – Plant Physiol. **24**: 1-15, 1949.
- Bakiaroglou, P. Karatagli, S.: Effect of lead & Pb on Mentha spicata. Journal of Agronomy and Crop Science. 188(3): 201-205, 2002.
- Bates, L.S. Waldran, R.P., Teare, L.D.: Rapid determination of free proline for water stress studies. - Plant & Soil. 39: 205-207, 1973.
- Bergmann, W.L: Nutritional disorders of plants. Gustav Fisher Verag, Jena – Stuffgart – New York. 1992.
- Bhattacharya, M., Choudhuri, M. A.: Effect of lead and cadmium on the biochemical changes in the leaves of terrestrial (*Vigna*) and aquatic (*Hydrilla*) plants under solution culture.
  Indian J. Plant Physiol. **32**. 99-103, 1997.
- Burzynski, M., Buczek, J.: Uptake and assimilation of ammonium ions by cucumber seedlings from solutions with different pH and addition of heavy metals. - Acta- Societati Botonicorum Poloniae. 67(2):197-200, 1998.
- Chaney, R.L.: Pb phytotoxicity In: Robson, A.D. (ed.): Pb in soil and plants. - Kluwer Academic Publishers, Dordrecht Pp. 135-150, 1993.
- Chhetri, D.R.m Supratin, Modak Ahmed Safiruddin.: Physiological and Biochemical response of two rice bean (*Vigna umbellata*) cultivars to heavy metal stress. - Environ & Ecology. **22**(1): 27-33, 2004.
- Delauney, A.J., Verma, D.P.S.: Proline biosynthesis and osmoregulation in plants. - Plant J. **4**:215-223, 1993.
- Farago, M.E., Muller, W.A. : Plants which accumulate metals. Part IV. A possible copper-proline complex from the roots of Armeria maritima. - Inorg. Chim Acta. 32, L93-94, 1979.

- Gratao, P. L., Prasad, M. N. V. Cardoso, P. F. Leda, P. J., Azevedo, R. A.: Phytoremediation: green technology for the clean up of toxic metals in the environment. - Brazilian Journal of Plant Physiology. 27 (1), 2005.
- Herrero, E. M., Lopex, Gonzalvez, A. Ruiz, M. A., Lucas, Garica, J. A., Barbas, C.: Uptake and distribution of Zinc, cadmium, lead and copper in *Brassica napus* Var. Oleifera and *Helianthus annuus* grown in contaminated soils. - Int. J. Phytoremediation. 5(2) 153-167, 2003.
- Jain, Radha, Srivastava, S., Madan, V.K.: Influence of Chromium on growth and cell division of sugar cane. - Indian J. Plant Physiology 5(3): 228-231, 2000.
- Jones, B., Wolf, B., Mills, H. A. Plant analysis Hand book: a practical sampling, preparation, analysis and interpretation guide. - Micro-Macro International, Athens GA, 1991.
- Joshi, V.N. Rathore, S.S. Arora, S.K.: Effect of Chromium on growth and development of cowpea (*Vigna unguicalata* L.). -India J. Environ. Prot. **19**, 745-749, 1999.
- Kaya, C. Higgs, D., Burton, A: Plant growth, Phosphorus nutrition and acid phosphatase enzyme activity in three tomato cultivars grown hydroponically on different Zinc concentrations. J. Plant Nutr. 23: 569-579, 2000.
- Khudsar, T.: Studies on the effects of some heavy metal pollutants on *Cajanus cajan* (Linn.) Huth. – Ph.D. Thesis Jamia Hamdard, New Delhi 1999.
- Khudsar, T., Mahmooduzzafar, Iqbal M. Siaram, R.K.: Pb induced changes in morpho-physiological and biochemical parameters in *Artemisia annua*. - Biologia Plantarum. 48(2): 255-260, 2004.
- Lesage, E., Meers, E., Vervacke, P., Lamsal, S., Hopgood, M, Tract, F. M., Verloo, M. G.: Enhanced phytoextraction: II. Effect of EDTA and citric acid on heavy metal uptake by *Helianthus annuus* from a calcareous soil. - Int. Journal Phytoremediation. 7(2): 143-152, 2005.
- Lowry, O.H. Roserborough, N.J., Farr, A.L., Randell, R.J.: Protein measurement with Folin-Phenol reagent. – J. Biol. Chem. 193: 265-275, 1951.
- Martens, S. N., Stephens, B. W., Grusak, M. A.: Identification of an iron translocator / putative signal molecule in the phloem of higher plants. - Plant Physiol. **S111**, 302 (Abstract), 1996.
- Meers, J.L. Tempest, D.W. Brown, C.M.: Glutamine (amide) 2oxyoglutarate amino transferase, oxido reductase (NADP) and enzyme involved in the synthesis of glutamate by some bacteria. - J. Gen. Microbial. 64: 187-194, 1970.
- Mehta, S.K., Gaur, J.P.: Heavy metal induced praline accumulation and its role in ameliorating metal toxicity in *Chlorella vulgaris*. - New Phytol. **143**: 253-259, 1999.
- Melinchuk, Y. U. P., Lish, Ko. A. K., Kalinin, F. L.: Cadmium effect on free amino acid content in germs of pea seeds at early germination stages. - Fiziologiya biokhimiya Kulturngh Rastenii. 14: 383-385, 1982.
- Miller, R.J., Bittell, J.E. and Kaeppe, D. E.: The effect of cadmium on electron and energy transfer reaction in corn mitochondria. Physiologia Plantarum. 28:166-171, 1973.
- Mishra, S.N. Sing, D.B., Chaudhary, Anju.: Nitrate and ammonium effect on Indian mustard seedling growth under salinity. J. Exp. Biol. 32: 916-918, 1996.
- Muthukumarsami, M., Karikalan, L., Rajan, s. n., Panneerselvam, R.: Effect of Nacl stress on protein, praline contents and protease adenosine triphosphatase activities in chickpea plants. Geobios (Jodhpur) 24(2-3): 119-123, 1997.
- Okhi, K: Pb Concentration in soybean as related to growth, photosynthesis and carbonic anhydrase activity. – Physiol Plant. **18**: 79-82, 1978
- Oven, M., Grill, E., Golan, Goldhirsh, A., Kutchan, T.M., Meinhart, H.Z.: Increase of free cysteine and citric acid in plant Cells exposed to cobalt ions. - Phytochemistry. 60: 467-474, 2002.
- Panda, S.K., Khan, M.H.: Antioxidant efficiency in rice (*Oryza sativa* L.) leaves under heavy metal toxicity. J. Plant Biology. **30(1)**: 23-29, 2003.
- Pollard, J. A., Powell, K.D., Harper, F. A., Smith, J.A.C.: The genetic basis of metal hyper accumulation in plants. - CRC Crit Rev. Plant Sci. 21 539-566, 2002.

- Prasad, D.D.K., Prasad, A.R.K.: Effect of Lead and Mercury on chlorophyll synthesis in mungbean plants. – Phytochem. 26: 881-883, 1987.
- Quariti, O. Gouia, Gharbai, M.H.: Responses of bean and tomato plants to cadmium growth, mineral nutrition and nitrate reduction. Plant Physiol Biochem. 35: 347-354, 1997.
- Shah, K., Dubey, R.S.: Effect of cadmium on proline accumulation and ribonuclease activity in rice seedlings: Role of Proline as possible enzyme protectant. - Biol. Plant. 40: 121-130, 1998.
- Singh, V.P., Srivastava, A., Singh, U.P.: Biochemical & Genetic effects of heavy metals plants. Plant Diversity, Microbial interaction and Environmental Biology, Edited: Dr. S.N. Chaturvedi and K.P. Singh: 215-225, 2005.
- Singh, R.P., Dabas, S., Choudhary, A., and Maheshwari, R.: Effect of lead on nitrate reductase activity and alleviation of lead toxicity by inorganic salts and 6-benzylaminopurine. -Biologia Plantarum. 40(3): 399-404.1998.
- Sinhal, V.K.: Phytotoxic, Cytogenetic and Biochemical effects of Pb<sup>2+</sup> & Pb<sup>2+</sup> in Vigna mungo (L). Hepper. - Ph.D. thesis, M.J.P. Rohilkhand University Bareilly, India, 2005.
- Sinhal, V.K., Srivastava, A.K., Singh, V.P.: Effects of Pb<sup>2+</sup> & Pb<sup>2+</sup> on nitrogen metabolism in *Vigna mungo* (L). Hepper. -Journal of Indian Botanical Society (In Press), 2006.
- Smirnoff, N., Cumbes, O.J.: Hydroxyl radical scavenging activity of compatible solutes. - Phytochemistery. 28: 1057-1060, 1989.
- Soares, C.R.F.S., Grazziotti, P.H., Siquaira, J.O. Carvalho, J.H. De.: Pb toxicity on growth and nutrition of *Eucalyptus maculata* and *Eucalyptus urophylla*. - Pesquisa Agropecuaria Brasileira. **36**(2): 339-348, 2001.
- Solhi, M., Hossain, Shareatmadari., Ha, Jabbasi. M.A.: Lead and Pb extraction potential of two common crop plants *Helianthus annuus* and *Brassica napus*. 'Water, Air & Soil Pollution. **167** (1-4): 59-71, 2005.
- Srivastava, H.S.: In vivo activity of nitrate reductase in maize plants. India J. Biochem and Biophys. **11**: 230-232, 1980.

- Stewert, G.R., Lee, J.A.: The Role of proline accumulation in halophytes. Planta. **120**: 279-280, 1974.
- Surasak, S., Samuel, B., Desh, P.S.V., Richard, T. Sayre. : Molecular Mechanisms of praline mediated tolerance to toxic heavy metal in transgenic Microalgal. - The plant cell. 14: 2837-2847, 2002.
- Suresh, B., Ravishankar, G.A.: Phytoremediation A novel and promising approach for environmental clean-up. - Crit. Rev. Biotechnol. 24: 97-124, 2004.
- Susan, Tandy. Rainer, Schulin.., Bernd, Nowack.: The influence of EDDS on the uptake of heavy metals in hydroponically grown sunflowers. - Chemosphere. 62 (9) 1454-1463, 2006.
- Taylor, C.B.: Proline and water deficit: UPS, downs, ins and outs. - Plant Cell. 8 KK 1- 1224.
- Tirpathy, B.C., Mohanty, P.: Pb inhibited electron transport of photosynthesis in isolated barley chloroplast. *Plant Physiol.* 66: 1174-1178, 1980.
- Tomar, Manju., Kaur, Indradeep., Neelu., Bhatnagar, A.K.: Effect of enhanced lead in soil on growth and development of *Vigna radiata* (L.) Wilezek. - Indian J. Plant Physiol. 5(1): 13-18, 2000.
- Van, Assche, F, Clijstors, H. Marcella, R.: Photosynthesis in *Phaseolus vulgaris* (L.) as influenced by Supra-optimal Pb nutrition – In: Marcelle, R. Ciijsters, It. Van Poucke, M.: Photosynthesis and Plant Development **Pp**. 175-184. Dr. W. Junk, The Hauge, 1979.
- Wang, Q. R., Cui, Y.s., Liu, X.M., Dong, Y.T. Christie, P.: Soil contamination and uptake of heavy metals at polluted sites in China. - J. Environ. Sci. Heal A 38: 823-8338, 2003.
- Woolhouse, H.W.: Toxicity and tolerance in the response of plants to metals. Pages 245-300 in O.L. Nobel, L.B. Osmond and H. Ziegler, editors. Physiological plant ecology III: Encyclopedia of plant physiology, new series, volume 12C. Springer-verlag, New York, U.S.A, 1983.
- Wolf, P.L., Williams, D.: Practical Clinical enzymology; Techniques and interpretation. - Wiley- Interscience Publication.NewYork,1973.