Effect of biofertilizers and organic supplements on the growth of black pepper rooted cuttings (*Piper nigrum* L.)

T S Aswathy*, M K Dhanya, J Jintu, T Sathyan, T T Preethy & M Murugan

Cardamom Research Station, Kerala Agricultural University, Pampadumpara-685 553, Idukki, Kerala.

*E-mail: aswathyrethna91@gmail.com

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Abstract

An experiment was conducted at the Cardamom Research Station, Kerala Agricultural University, Pampadumpara (Kerala) with an objective to study the effect of different biofertilizers (Phosphorus solubilizing bacteria, *Azospirillum* and Plant Growth Promoting Rhizobacteria Mix I) and organic supplements (fish extract and humic acid) on the growth of black pepper rooted cuttings. The results of the experiment indicated that application of Phosphorus solubilizing bacteria (5 g) along with *Azospirillum* (5 g), humic acid (0.2%) and fish extract (0.5%) was the best combination for the production of black pepper rooted cuttings with improved vegetative characters (plant height, number of leaves, number of roots, length of roots and leaf area) compared to their individual inoculation. This innovative information can be effectively utilized and advocated for the commercial production of black pepper rooted cuttings with lusty growth.

Keywords: black pepper, bio fertilizers, organic supplements, roots and shoot growth, nutrient management

The availability of quality planting material is one of the major issues that black pepper growers face in India. The conventional propagation methods have several limitations due to low success rate, poor rooting, spread of soil borne pathogens and poor survival rate of transplanted rooted cuttings (Rini et al. 2016). In order to overcome these problems, a technological intervention is needed to boost both the production and supply of quality planting materials. In the context of promoting organic cultivation, the use of biofertilizers for quality seedlings/cuttings production secures much importance because they are eco-friendly, low cost, capable of improving crop yields and quality sustainably. Biofertilizers improve growth rate of plants and soil health as they act as plant strengtheners, phytostimulators, plant health improvers, and have the potential to fix nitrogen (Babalola 2014). The fertility of the soil is also restored by biofertilizers so that plants were better protected from getting any diseases (Amna 2010). Fish emulsions are used as a source of nitrogen during the early or vegetative stage of development to boost plant growth and also this has been documented to promote seedling growth in many crops (Murray & Anderson 2004). Humic substances
are organic materials that improve soil structure, fertility and nutrient uptake (Trevisan et al. 2010). Hence, the present study was designed and conducted at the Cardamom Research Station, Pampadumpara, Kerala Agricultural University during 2017 to evaluate the efficacy of promising bio-fertilizers and select organic supplements on the production of quality black pepper rooted cuttings.

The most popular local black pepper variety ‘Karimunda’ was chosen in the evaluation. This study comprised eight treatments including untreated control, sole and combined application of following bioagents and organic supplements.

<table>
<thead>
<tr>
<th>Treatments with dose</th>
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</thead>
<tbody>
<tr>
<td>T&lt;sub&gt;1&lt;/sub&gt;</td>
</tr>
<tr>
<td>T&lt;sub&gt;2&lt;/sub&gt;</td>
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<td>T&lt;sub&gt;3&lt;/sub&gt;</td>
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<td>T&lt;sub&gt;4&lt;/sub&gt;</td>
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<td>T&lt;sub&gt;5&lt;/sub&gt;</td>
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<td>T&lt;sub&gt;7&lt;/sub&gt;</td>
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<td>T&lt;sub&gt;8&lt;/sub&gt;</td>
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</tbody>
</table>

Local Karimunda cuttings were planted in polythene bags size of 20 × 15 cm filled with potting mixture composed of garden soil, sand, and farm yard manure in 1:1:1 proportion. The potting mixture had an initial nutrient status of organic carbon (1.98%), N (0.704 mg/100 g) available P (4.01 mg/100 g) and K (43 mg/100 g) with a pH of 5.3. The treatments were superimposed and evaluated <i>in vivo</i> under greenhouse condition. Commercial formulations of phosphorus solubilizers (5 × 10<sup>7</sup> cfu), Azospirillum (5 × 10<sup>7</sup> cfu) and PGPR Mix I (Consortium of Azospirillum lipoferum, Azotobacter chroococcum, Bacillus megaterium and Bacillus sporothermodurans each with 5 × 10<sup>7</sup> cfu) were obtained from the Department of Agricultural Microbiology, College of Agriculture, Vellayani. The humic acid containing 12% potassium humate was used in this study. Fish extract was prepared as per the standard procedure. Equal quantity of fish and jaggery was taken, sliced, mixed and kept in closed container with periodical stirring up to 30 days, after that the solution was strained through a muslin cloth and stored.

The experiment was conducted using Completely Randomized Design (CRD) with four replications. Five plants were kept in each replication. In addition to this five plants/treatment were also maintained for destructive sampling to study the root characters. Talc based formulation of PSB, Azospirillum, PGPR Mix I, humic acid and fish extract individually and in combination were applied to the root zone of one month old black pepper cuttings grown in polythene bags. The treatments were given twice at fortnightly interval (when the plants were 30 and 45 days old).

Observations on the height, number of leaves, leaf area and root characters were recorded 60 days after second treatment application. The experimental soil (potting media) was analyzed for physicochemical properties like pH, organic carbon and available N, P and K content. The leaf chlorophyll content was estimated as per the protocol of Sadasivam & Manickam (1992). The data collected from the experiment were processed statistically with appropriate statistical tool for the interpretation of the results.

There was no significant difference with respect to morphological characters before treatment application. But good improvement in vegetative characters was noted after the application of biofertilizers and organic supplements. Black pepper rooted cuttings responded well to combined inoculation of
biofertilizers and organic supplements compared to individual inoculation and control (Table 1). Application of PSB along with *Azospirillum*, humic acid and fish extract (T<sub>6</sub>) resulted in increased plant height (98.66 cm), numbers of leaves (12.16), number of roots (13.50), root length (28 cm) and highest leaf area (63.18 cm<sup>2</sup>) 60 days after second treatment application compared to all the other treatments. Similar increase of growth was recorded in black pepper for combined inoculation of biofertilizers such as *Azospirillum*, Phosphobacteria and VAM (Kandiannan et al. 2000; Bopaiah & Khader 1989). In addition to this, fish emulsions have been reported to increase the nitrogen accessibility (Weinert et al. 2014). According to Chen & Aviad (1990) the application of humic substances increase root length and produce more secondary roots. As a result, the plants were capable of absorbing more available nutrients from the soil which in turn resulted better establishment and subsequent growth and development. Thus, the combined inoculation of PSB and *Azospirillum* along with humic acid and fish extract performed best through improving the morphological characters of the cuttings compared to their individual application.

Total chlorophyll content was maximum in plants treated with PSB along with *Azospirillum*, humic acid and fish extract (0.64 mg/100 g) followed by the combined application of PGPR Mix I along with *Azospirillum*, humic acid and fish extract (0.60 mg/100 g). The least quantity of chlorophyll was observed in control plants (0.42 mg/100 g). This can be attributed to the increased uptake of nutrients leading to enhanced chlorophyll content. Pereira et al. (2015) reported that maize plants when inoculated with *Azospirillum* under different dosages of nitrogen enhanced the chlorophyll content. Our best treatment also includes *Azospirillum* as one component. This contributes to the increased chlorophyll content of the treated plants.

Nutrient status of the potting mixture 60 days after second treatment application (Table 2) showed that, readily available nutrient content of potting mixture was improved through the application of biofertilizers and organic supplements compared to control. Among the treatments, PSB along with *Azospirillum* humic acid and fish extract treated soil showed higher levels of organic carbon (3.17%), available phosphorus (17.83 mg/100 g) and potassium (84.66 mg/100 g) whereas, available nitrogen was highest for combined application of PGPR Mix I along with *Azospirillum*, humic acid and fish extract (1.05 mg/100 g). Increased availability of nutrients in the potting mixture was attributed through combined application of biofertilizers and organics. Bio inoculants have had influence in increasing the organic carbon content of turmeric (Sumathi et al. 2011). Biofertilizers like *Azospirillum* and PSB were highly beneficial to plants through augmentation of nitrogen and phosphorus content in soil, thus making these two essential nutrients available to the plant and also produce phytohormones like auxins (Singh et al. 2011; Rocheli et al. 2015). In our experiment also treatments consisting of PSB along with *Azospirillum*, humic acid and fish extract (T<sub>6</sub>) resulted in high available organic carbon, P & K compared to control which is responsible for increased growth parameters observed. The increase in P availability could be attributed to the application of PSB which produces organic acids that act as a chelating agent and thereby, releases P into the soil solution and making it more available resulted in improved root growth. These findings are in agreement with the findings of Naik & Hari babu (2007) and Sharma et al. (2009) in guava.

It is evident that, the combined application of PSB (5 g) along with *Azospirillum* (5 g), 0.2% humic acid (100 mL plant<sup>-1</sup>) and 0.5% fish extract (100 mL plant<sup>-1</sup>) followed by PGPR Mix I (5 g) along with 0.2% humic acid (100 mL plant<sup>-1</sup>) and 0.5% fish extract (100 mL plant<sup>-1</sup>) produced healthy black pepper rooted cuttings with good morphological characters than those of sole application of any of these biofertilizers and organics. Therefore, this technology could be effectively advocated to produce black pepper root cuttings with lusty growth in nurseries.
Table 1. Effect of biofertilizers and organic supplements on growth of black pepper rooted cuttings after second treatment application

<table>
<thead>
<tr>
<th>Treatments</th>
<th>Plant height (cm)</th>
<th>Number of leaves</th>
<th>Number of roots</th>
<th>Length of roots</th>
<th>Leaf area (cm)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Phosphorus solubilizing bacteria (PSB)- 10 g plant(^{-1})</td>
<td>76.50</td>
<td>10.33</td>
<td>12.83</td>
<td>21.00</td>
<td>48.45</td>
</tr>
<tr>
<td><em>Azospirillum</em> - 10 g plant(^{-1})</td>
<td>75.50</td>
<td>9.00</td>
<td>9.16</td>
<td>15.86</td>
<td>43.29</td>
</tr>
<tr>
<td>PGPR Mix I- 10 g plant(^{-1})</td>
<td>73.83</td>
<td>10.00</td>
<td>9.83</td>
<td>17.16</td>
<td>46.02</td>
</tr>
<tr>
<td>Humic acid @2%</td>
<td>67.33</td>
<td>8.50</td>
<td>9.20</td>
<td>15.46</td>
<td>45.60</td>
</tr>
<tr>
<td>Fish amino acid @0.5%</td>
<td>70.66</td>
<td>8.83</td>
<td>6.26</td>
<td>11.83</td>
<td>36.98</td>
</tr>
<tr>
<td>Phosphorus solubilizing bacteria (PSB) 5 g plant(^{-1}) + <em>Azospirillum</em> 5 g plant(^{-1}) + Humic acid @2% + Fish amino acid @0.5%</td>
<td>98.66</td>
<td>12.16</td>
<td>13.50</td>
<td>28.00</td>
<td>63.18</td>
</tr>
<tr>
<td>PGPR Mix I 5 g plant(^{-1}) + Humic acid @2% + Fish amino acid @0.5%</td>
<td>79.50</td>
<td>10.66</td>
<td>10.16</td>
<td>36.00</td>
<td>48.91</td>
</tr>
<tr>
<td>Control</td>
<td>49.33</td>
<td>6.50</td>
<td>5.13</td>
<td>6.33</td>
<td>23.52</td>
</tr>
<tr>
<td>CD (P&lt;0.05)</td>
<td>18.779</td>
<td>2.418</td>
<td>0.491</td>
<td>1.805</td>
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<tr>
<td>CV</td>
<td>14.352</td>
<td>14.409</td>
<td>3.027</td>
<td>5.425</td>
<td>0.320</td>
</tr>
</tbody>
</table>

Table 3. Effect of biofertilizers and organic supplements on nutrient status of potting mixture 60 days after second treatment application

<table>
<thead>
<tr>
<th>Treatments</th>
<th>pH</th>
<th>Organic carbon %</th>
<th>Available N (mg/100 g)</th>
<th>Available P (mg/100 g)</th>
<th>Available K (mg/100 g)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Phosphorus solubilizing bacteria (PSB)- 10 g plant(^{-1})</td>
<td>6.7</td>
<td>2.64</td>
<td>0.906</td>
<td>11.41</td>
<td>73.16</td>
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<tr>
<td><em>Azospirillum</em> - 10 g plant(^{-1})</td>
<td>6.5</td>
<td>2.63</td>
<td>0.900</td>
<td>13.63</td>
<td>74.83</td>
</tr>
<tr>
<td>PGPR Mix I- 10 g plant(^{-1})</td>
<td>7.1</td>
<td>2.65</td>
<td>0.913</td>
<td>14.29</td>
<td>75.83</td>
</tr>
<tr>
<td>Humic acid @2%</td>
<td>6.3</td>
<td>2.63</td>
<td>0.903</td>
<td>11.35</td>
<td>68.83</td>
</tr>
<tr>
<td>Fish amino acid @0.5%</td>
<td>6.8</td>
<td>2.64</td>
<td>0.903</td>
<td>7.02</td>
<td>66.83</td>
</tr>
<tr>
<td>Phosphorus solubilizing bacteria (PSB) 5 g plant(^{-1}) + <em>Azospirillum</em> 5 g plant(^{-1}) + Humic acid @2% + Fish amino acid @0.5%</td>
<td>7.38</td>
<td>3.17</td>
<td>0.970</td>
<td>17.83</td>
<td>84.66</td>
</tr>
<tr>
<td>PGPR Mix I 5 g plant(^{-1}) + Humic acid @2% + Fish amino acid @0.5%</td>
<td>7.25</td>
<td>2.92</td>
<td>1.053</td>
<td>14.75</td>
<td>80.33</td>
</tr>
<tr>
<td>Control</td>
<td>5.5</td>
<td>2.01</td>
<td>0.706</td>
<td>3.56</td>
<td>46.00</td>
</tr>
<tr>
<td>CD (P&lt;0.05)</td>
<td>0.444</td>
<td>0.061</td>
<td>0.024</td>
<td>0.206</td>
<td>5.356</td>
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<tr>
<td>CV</td>
<td>3.915</td>
<td>1.411</td>
<td>1.550</td>
<td>0.275</td>
<td>4.336</td>
</tr>
</tbody>
</table>
References


Effect of biocontrol agents on production of rooted back pepper cutting by serpentine method

R S Bhai*, K P Subila, S J Eapen, A Reshma, R Pervez, A Ishwara Bhat & V Srinivasan

ICAR-Indian Institute of Spices Research, Marikkunnu PO, Kozhikode-673 012, Kerala.
*E-mail: rsbhai@rediffmail.com

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Abstract

Availability of disease free quality planting material is a major limiting factor in black pepper cultivation. In order to meet the increasing demand and also to create awareness on good agricultural practices for healthy disease free planting material production to farmers, a nursery experiment was started with improved varieties of black pepper by adopting a non-chemical bio-intensive management strategy. Here solarization of potting mixture was the main concern followed my amending the solarized potting mixture with potential bioagents. The experiment was designed in a two factor CRD with four improved varieties and five treatments. Each treatment contains a combination of two bioagents with antifungal and nematicidal properties respectively. The common recommended fungicide Metalaxyl-Mancozeb (0.125%) and nematicide carbsosulfan (0.1%) was used as control. The treatments were incorporated individually into solarized potting mixture and planted with improved varieties used viz., IISR Girimunda, IISR Malabar Excel, IISR Shakti and IISR Thevam, The plants in each treatment were kept for multiplication by serpentine method with proper irrigation and phytosanitation. The results of plant growth and establishment in different treatments, showed that fortification of solarized potting mixture with Trichoderma harzianum + Pochonia chlamydosporia combination or combination of Streptomyces strains (Act 2+9) are significantly superior (35.46% and 21% respectively) for the production of healthy rooted planting material. IISR Malabar Excel and IISR Thevam produced the maximum number of plants from a single node cutting in treatment with T. harzianum + P. chlamydosporia (T1) (59 nos. and 51 nos. respectively) followed by IISR Malabar Excel with Act 2+9 and Act 5+9 (45 nos. each). So an average of 6-7 plants/month/cutting was produced in the potential treatment while it was only 3-4 plants in control. The advantage of the method is that, after solarization and fortification with respective bioagents, there is no need for further application of any fungicides, insecticides or any other nutrient spray as usually done. Thus the method of soil solarization followed by fortification of either with T. harzianum+ P. chlamydosporia or combination of Streptomyces strains viz., Ketasatospora setae (Act 2) and S. tauricus (Act9) is found suitable for the production of healthy quality planting material of high yielding varieties to meet the increasing demand of planting material with a C:B ratio of 1:2.
The demand for healthy planting material of high yielding black pepper varieties is on the increase and the country needs large quantity of quality planting material to meet the increasing demand. The present study was aimed to establish a model nursery to produce disease free healthy planting material of improved varieties of black pepper by creating awareness among farmers with good agricultural practices and by adopting non-chemical bio-intensive management strategies.

The experiment was conducted in collaboration with a farmer at Omasserry in Thamarassery Panchayat (Kozhikode district). Initially a nursery shed was constructed (24 m × 20 m) and roofed with white polythene sheet of 100 microns. Single node rooted plants of four released varieties of black pepper viz., IISR Girimunda, IISR Malabar Excel, IISR Shakti and IISR Thevam (indexed for viruses and raised under insect proof conditions at ICAR-Indian Institute of Spices Research, Chelavoor) was used as the source material.

Nursery mixture was prepared by mixing soil, sand and FYM in 1:2:1 and sterilized by solarization. Briefly, the nursery mixture prepared was made into small beds of size 3x1m in a place where there is direct exposure to sunlight. The bed was watered thoroughly and covered with polythene sheet of 100 microns and sealed air tight and kept for solarization on 5th November till 25th December 2015.

Biocontrol agents such as *Trichoderma harzianum*, *Pochonia chlamydospora*, and 3 promising *Streptomyces* sp. viz., *Streptomyces tauricus* (strain Act 9), *Streptomyces* sp. (strain Act 5) and *Ketosatospora setae* (strain Act 2) (Bhai et al. 2016) in different combinations were used as growth promoters as well as bioagents for incorporating into the nursery mixture.

The experiment was designed in a two factor CRD with four improved varieties of IISR and five treatments. The five treatments were T1- *T. harzianum* + *P. chlamydospora*, T2- *Streptomyces* strains 2+9, T3- *Streptomyces* strains 5+9, T4- Metalaxyl-Mz+ Carbsosulfan and T5-control without any amendments. The individual treatments were incorporated with the solarized nursery mixture separately and filled in polythene bags (15 cm × 10 cm) @250 g bag⁻¹. *T. harzianum* and *P. chlamydospora* were made in liquid form with water and added @100 mL (cfu 10⁸ mL⁻¹) each to 100 kg potting mixture. *Streptomyces* spp. grown as broth culture in Nutrient broth and 100 mL (cfu 10⁶ mL⁻¹) mixed with 1 kg vermicompost and grown for 5 days (cfu10⁸ mL⁻¹) and applied @1 kg 100 kg⁻¹ potting mixture. The treatment imposed poly bags were arranged inside the nursery and planted with single node virus free (indexed) rooted cuttings as mentioned above and were allowed to grow by serpentine method.

When the number of rooted nodes in the serpentine reached around 10, the rooted middle cuttings were cut and separated leaving three plants at the tip and nucleus plant at the end and were kept for establishment to a 3-4 leaf stage in the same nursery. Five plants each were taken from each treatment and observed for biometric growth parameters. The biometric observations were recorded on height of the plant, fresh and dry weight of the plant, number of roots, root length and root biomass. The soil was analysed for the presence of targeted pathogens like *Phytophthora capsici* and nematodes (*Radopholus similis* and *Meloidogyne incognita*), pH and dehydrogenase activity (DHA).

The data were analyzed by using PROC ANOVA procedure of SAS 9.3. Least square means statements were used for mean separation.

After nine months of growth by serpentine method, the variety IISR Malabar Excel and IISR Thevam produced the maximum number of plants in T1 (*T. harzianum* + *P. chlamydospora*) (59 nos. and 51 nos. respectively) followed by T2 (Act 2+9) in case of Malabar Excel (45nos) and T3 (Act 5+9) (45 nos) in case of Thevam, from a single node cutting. An average of 6-7 plants/month/cutting was produced from these varieties with the treatment T1, while it was only 4 plants/month/cutting in control. In case of IISR Girimunda, the performance was almost
the same with all the three bioagent combinations when compared to Metalaxyl-Mz + Carbsosulfan and control. However, IISR Shakti showed comparatively lesser performance with bioagent combinations. The results of the study clearly showed the response of varieties to bioagents. In all cases, the number of plants produced with Metalaxyl-Mz + Carbsosulfan (T4) was comparatively lesser when compared to control (Table 1). The root system was also healthy in all treatments except for control where the root was not profusely grown. No disease of any kind was observed in any of the plants. Though there is no difference between varieties in fresh weight of the plant, the dry weight (Fig. 1) is significantly superior in treatment with *Streptomyces* strains (Act 2+9) and is at par with *T. harzianum* and *P. chlamydosporia*. Not much difference was observed in the number of leaves between treatments but, there is difference in the height of the plants (Fig. 2) where bioconsortia showed increased height when compared to Metalaxyl-Mz + Carbsosulfan and control. Difference was observed in number of roots, root length, and root fresh and dry biomass (Tables 2 to 5). Since the nursery mixture was solarised and irrigation was limited to once in two days, there was no incidence of soil borne infections caused by *Phytophthora capsici*, *Sclerotium rolfsii* or nematodes (the common diseases otherwise observed in nurseries), in any of the treatments including control (Table 6). The pH of the soil in untreated plants ranged from 4.55-5.66. It is interesting to note that the pH of the soil is raised to neutral in treatments with *T. harzianum* + *P. chlamydosporia* where it ranged from 6.86-7.63. In the case of *Streptomyces* combinations the pH ranged from 4.85 to 6.85 (Table 6) and the dehydrogenase activity which reflects the total oxidative activity of soil microflora, (Liang *et al.* 2014) was unaffected by the incorporation of bioagents like *Trichoderma*, *Pochonia* or *Streptomyces* sp. (Table 6). So without the addition of external nutrients, the micro flora enriched solarized mixture supported the growth of plants as well as prevented the incidence of infection caused by nematodes, *Phytophthora* or any other soil borne pathogens of black pepper. This may be due to the increased microbial activity either through the production of IAA or other growth promoting traits including siderophore production which is observed in the case *Streptomyces* strains (Suseela Bhai *et al.* 2016).

Thankamani *et al.* (2005) reported the effect of *Pseudomonas fluorescens* (IISR-6) and *T. harzianum* (P-26) on the growth of black pepper rooted cuttings in the nursery. However, no

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**Table 1. No. of cuttings produced in nine months from a single plants**

<table>
<thead>
<tr>
<th>Treatments</th>
<th>IISR Girimunda</th>
<th>IISR Malabar Excel</th>
<th>IISR Shakthi</th>
<th>IISR Thevam</th>
<th>Sub plot mean</th>
<th>Increase over control (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>T1 <em>T. harzianum</em> + <em>P. chlamydosporia</em></td>
<td>36.00</td>
<td>59.60</td>
<td>23.00</td>
<td>51.00</td>
<td>42.40</td>
<td>35.46</td>
</tr>
<tr>
<td>T2 Act 2+9</td>
<td>36.00</td>
<td>45.60</td>
<td>32.00</td>
<td>38.00</td>
<td>37.90</td>
<td>21.08</td>
</tr>
<tr>
<td>T3 Act 5+9</td>
<td>34.00</td>
<td>34.00</td>
<td>32.60</td>
<td>45.80</td>
<td>36.60</td>
<td>15.00</td>
</tr>
<tr>
<td>T4 <em>Metalaxyl-Mancozeb</em> + Carbsosulfan</td>
<td>25.40</td>
<td>26.40</td>
<td>34.00</td>
<td>32.60</td>
<td>29.60</td>
<td>-5.43</td>
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<tr>
<td>T5 Control</td>
<td>23.60</td>
<td>29.40</td>
<td>41.60</td>
<td>30.60</td>
<td>31.30</td>
<td>-</td>
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<tr>
<td>Main plot mean</td>
<td>31.00</td>
<td>39.00</td>
<td>32.64</td>
<td>39.60</td>
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<td>LSD (P&lt;0.05)</td>
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<tr>
<td>VT</td>
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<td>V × T</td>
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</table>
Fig. 1. Dry wt (g)/plant

Fig. 2. Height of the plants
### Table 2. Influence of treatments on number of roots

<table>
<thead>
<tr>
<th>Treatments</th>
<th>IISR Girimunda</th>
<th>IISR Malabar Excel</th>
<th>IISR Shakthi</th>
<th>IISR Thevam</th>
<th>Sub plot mean</th>
</tr>
</thead>
<tbody>
<tr>
<td>T1- <em>T. harzianum</em> + <em>P. chlamydosporia</em></td>
<td>7.67</td>
<td>8.67</td>
<td>4.00</td>
<td>12.67</td>
<td>8.25</td>
</tr>
<tr>
<td>T2-Act 2+9</td>
<td>6.00</td>
<td>11.33</td>
<td>8.33</td>
<td>10.00</td>
<td>8.92</td>
</tr>
<tr>
<td>T3-Act 5+9</td>
<td>6.67</td>
<td>8.00</td>
<td>9.33</td>
<td>8.33</td>
<td>8.08</td>
</tr>
<tr>
<td>T4-Metalaxyl-Mancozeb + Carbsosulfan</td>
<td>6.33</td>
<td>7.33</td>
<td>5.00</td>
<td>8.00</td>
<td>6.67</td>
</tr>
<tr>
<td>T5-Control</td>
<td>8.67</td>
<td>6.33</td>
<td>5.33</td>
<td>7.33</td>
<td>6.92</td>
</tr>
<tr>
<td>Main plot mean</td>
<td>7.07</td>
<td>8.33</td>
<td>6.4</td>
<td>9.27</td>
<td></td>
</tr>
<tr>
<td>LSD (P&lt;0.05)</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>V</td>
<td>2.02</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>T</td>
<td>NS</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>V × T</td>
<td>NS</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

### Table 3. Influence of treatments on root dry biomass (g)

<table>
<thead>
<tr>
<th>Treatments</th>
<th>IISR Girimunda</th>
<th>IISR Malabar Excel</th>
<th>IISR Shakthi</th>
<th>IISR Thevam</th>
<th>Sub plot mean</th>
</tr>
</thead>
<tbody>
<tr>
<td>T1- <em>T. harzianum</em> + <em>P. chlamydosporia</em></td>
<td>0.99</td>
<td>0.62</td>
<td>0.52</td>
<td>0.97</td>
<td>0.78</td>
</tr>
<tr>
<td>T2-Act 2+9</td>
<td>0.93</td>
<td>0.98</td>
<td>0.57</td>
<td>0.54</td>
<td>0.76</td>
</tr>
<tr>
<td>T3-Act 5+9</td>
<td>0.51</td>
<td>0.50</td>
<td>0.3</td>
<td>0.40</td>
<td>0.54</td>
</tr>
<tr>
<td>T4-Metalaxyl-Mancozeb + Carbsosulfan</td>
<td>0.47</td>
<td>0.54</td>
<td>0.37</td>
<td>0.51</td>
<td>0.47</td>
</tr>
<tr>
<td>T5-Control</td>
<td>0.44</td>
<td>0.41</td>
<td>0.41</td>
<td>0.56</td>
<td>0.46</td>
</tr>
<tr>
<td>Main plot mean</td>
<td>0.67</td>
<td>0.61</td>
<td>0.52</td>
<td>0.6</td>
<td></td>
</tr>
<tr>
<td>LSD (P&lt;0.05)</td>
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<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>V</td>
<td>0.05</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>T</td>
<td>0.14</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>V × T</td>
<td>0.29</td>
<td></td>
<td></td>
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<td></td>
</tr>
</tbody>
</table>

### Table 4. Influence of treatments on root length (cm)

<table>
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<tr>
<th>Treatments</th>
<th>IISR Girimunda</th>
<th>IISR Malabar Excel</th>
<th>IISR Shakthi</th>
<th>IISR Thevam</th>
<th>Sub plot mean</th>
</tr>
</thead>
<tbody>
<tr>
<td>T1- <em>T. harzianum</em> + <em>P. chlamydosporia</em></td>
<td>31.67</td>
<td>36.33</td>
<td>19.00</td>
<td>22.33</td>
<td>27.33</td>
</tr>
<tr>
<td>T2-Act 2+9</td>
<td>29.00</td>
<td>32.67</td>
<td>21.00</td>
<td>24.33</td>
<td>26.75</td>
</tr>
<tr>
<td>T3-Act 5+9</td>
<td>26.67</td>
<td>26.67</td>
<td>24.67</td>
<td>26.00</td>
<td>26.00</td>
</tr>
<tr>
<td>T4-Metalaxyl-Mancozeb + Carbsosulfan</td>
<td>21.67</td>
<td>29.67</td>
<td>18.00</td>
<td>28.00</td>
<td>24.33</td>
</tr>
<tr>
<td>T5-Control</td>
<td>18.00</td>
<td>26.33</td>
<td>28.33</td>
<td>24.00</td>
<td>24.17</td>
</tr>
<tr>
<td>Main plot mean</td>
<td>25.4</td>
<td>30.33</td>
<td>22.20</td>
<td>24.93</td>
<td></td>
</tr>
<tr>
<td>LSD (P&lt;0.05)</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>V</td>
<td>5.29</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>T</td>
<td>NS</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>V × T</td>
<td>NS</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>
Table 5. Influence of treatments on root fresh biomass (g)

<table>
<thead>
<tr>
<th>Treatments</th>
<th>IISR Girimunda</th>
<th>IISR Malabar Excel</th>
<th>IISR Shakthi</th>
<th>IISR Thevam</th>
<th>Sub plot mean</th>
</tr>
</thead>
<tbody>
<tr>
<td>T₁ - T. harzianum + P. chlamydosporia</td>
<td>5.37</td>
<td>5.83</td>
<td>3.53</td>
<td>6.70</td>
<td>5.36</td>
</tr>
<tr>
<td>T₂ - Act 2+9</td>
<td>4.37</td>
<td>7.07</td>
<td>3.83</td>
<td>4.23</td>
<td>4.88</td>
</tr>
<tr>
<td>T₃ - Act 5+9</td>
<td>2.87</td>
<td>4.37</td>
<td>5.63</td>
<td>3.37</td>
<td>4.06</td>
</tr>
<tr>
<td>T₄ - Metalaxyl-Mancozeb + Carbsosulfan</td>
<td>2.47</td>
<td>4.07</td>
<td>2.93</td>
<td>4.00</td>
<td>3.37</td>
</tr>
<tr>
<td>T₅ - Control</td>
<td>2.97</td>
<td>3.47</td>
<td>3.30</td>
<td>3.97</td>
<td>3.43</td>
</tr>
<tr>
<td>Main plot mean</td>
<td>3.61</td>
<td>4.96</td>
<td>3.85</td>
<td>4.45</td>
<td></td>
</tr>
<tr>
<td>LSD (P&lt;0.05)</td>
<td>V</td>
<td>0.54</td>
<td>T</td>
<td>1.32</td>
<td>V × T</td>
</tr>
</tbody>
</table>

Table 6. Effect of treatments on pH and Dehydrogenase activity

<table>
<thead>
<tr>
<th>Treatments</th>
<th>IISR Girimunda</th>
<th>IISR Malabar Excel</th>
<th>IISR Shakthi</th>
<th>IISR Thevam</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>pH</td>
<td>DHA</td>
<td>pH</td>
<td>DHA</td>
</tr>
<tr>
<td>T₁ - T. ha + P. chal</td>
<td>7.38</td>
<td>1.21</td>
<td>6.86</td>
<td>2.72</td>
</tr>
<tr>
<td>T₂ - Act 2+9</td>
<td>5.56</td>
<td>3.57</td>
<td>4.85</td>
<td>2.74</td>
</tr>
<tr>
<td>T₃ - Act 5+9</td>
<td>5.05</td>
<td>3.17</td>
<td>6.68</td>
<td>3.13</td>
</tr>
<tr>
<td>T₄ - Met-Mz + Carbsosulfan</td>
<td>6.46</td>
<td>2.52</td>
<td>5.44</td>
<td>1.98</td>
</tr>
<tr>
<td>T₅ - Control</td>
<td>5.64</td>
<td>0.94</td>
<td>4.55</td>
<td>0.96</td>
</tr>
<tr>
<td>CD (P&lt;0.05)</td>
<td>0.10</td>
<td>0.09</td>
<td>0.09</td>
<td>0.02</td>
</tr>
</tbody>
</table>

such reports are available for the combined use of T. harzianum and P. chlamydosporia for the production of rooted plants of black pepper, except for the individual use of T. harzianum against foot rot and P. chlamydosporia against slow decline diseases. The present study recommend soil solarisation along with the use of Trichoderma + Pochonia combination or combination of Streptomyces strains Act 2+9 for the production of healthy rooted planting material. The effect of solarized potting mixture on growth of black pepper rooted cuttings was reported earlier (Thankamani et al. 2007). Since both Phytophthora and nematodes are serious threats of black pepper, the treatment combinations are made in such a way that the combination contain one antagonist against Phytophthora and another antagonist against nematode. Similar work was reports for the use of consortia in planting material production. Consortia of P. fluorescens + T. harzianum + Paecilomyces lilacinus were used for the production of tomato seedlings for combating nematode infection (Mukhtar 2013) and P. fluorescens + T. harzianum for the production of nematode free papaya seedlings (Rae 2007). The promotive effects of Pseudomonas and Trichoderma were quite significant in growth promotion in tomato during nursery and crop growth stages (Kumar et al. 2007). Similarly,
Mukhtar (2013) reported the biocontrol potential of *Pasteuria penetrans*, *P. chlamydosporia*, *P. lilacinus* and *T. harzianum* against *Meloidogyne incognita* in okra.

**Acknowledgements**

The authors are thankful to Department of Agriculture for funding, Mr. George Nedumkallel, farmer at Omasserry, Kerala State for active cooperation and involvement in supporting the nursery trial, Mr. K. Jayarajan, Chief Technical Officer for statistical analysis of the data and Mr. Vishnu P T for constant support in monitoring of the nursery.

**References**

Kumar Sunil, Arya M C & Singh Ranjit 2007 Efficiency of *Pseudomonas fluorescens* and *Trichoderma harzianum* as bio-enhancers in tomato at high altitude in central Himalayas. Indian J. Crop Sci. 2: 79–82.


Mukhtar T, Hussain M A & Kayani M Z 2013 Biocontrol potential of *Pasteuria penetrans*, *Pochonia chlamydosporia*, *Paecilomyces lilacinus* and *Trichoderma harzianum* against *Meloidogyne incognita* in okra. Phytopathologia mediterranea 52: 66–76.


Diversity in floral characters of monoecious nutmeg (*Myristica fragrans* Houtt.)

S Aarthi*, J Rema, B Sasikumar, K V Saji & M Anandaraj

ICAR-Indian Institute of Spices Research, Kozhikode-673 012, Kerala.
*E-mail: aarthichandru@gmail.com

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Abstract

Floral diversity in monoecious type nutmeg reveals three types of flowers namely; pistillate, staminate and hermaphrodite flowers. The structure of hermaphrodite flowers in nutmeg is reported for the first time. The flowers are borne on the leaf axil, flowering habit of the three types of flowers are seen in cymes as well as solitary in the same tree. The gynoecium consists of single ovary with bifid stigma in the pistillate flowers and the androecium of staminate flowers has adnate 7-13 anthers. Intra flower variability is evident in case of hermaphrodite flowers. In hermaphrodite flower the androecium ranges with 1-4 anthers, in the form of fused filament or free filament or both; some anthers are fused with the gynoecium. Besides remnants of the stamen in the developed fruits of hermaphrodite flowers, staminodes are also observed in the flowers. Colour of all the three types of flowers are light creamy yellow, with thick gamosepalous perianth which bursts as bilobed, trilobed, tetralobed and pentalobed lobes during anthesis. Analysis of variance of trees for flower types and floral attributes like flower type, number of anthers, length of anthers, length of filaments and length of ovary are found to be highly significant. High coefficient of variation is recorded for hermaphrodite (183.84%) and pistillate (171.71%) flowers. The pollen viability of the hermaphrodite flowers are found less as compared to the staminate flowers. The percentage of pollen viability is 79.74 and 90.77 in hermaphrodite and male flower respectively. In the population studied, the occurrence of hermaphrodite flower in monoecious tree ranged from 0 - 10%.

**Keywords:** hermaphrodite flowers, intra floral variability, monoecious, *Myristica fragrans* Houtt., pollen viability

Nutmeg (*Myristica fragrans* Houtt.) (Myristicaceae) is an important tree spice, yielding two spices, namely, the nutmeg (dried seed) and the mace (dried aril surrounding the seed). Nutmeg is hitherto considered to be predominantly dioecious in nature (Flach 1966). However, of late this concept is undergoing a paradigm shift, as monoecious trees are being reported often across the country (Krishnamoorthy *et al.* 1996; Krishnamoorthy *et al.* 2012; Rema *et al.* 2014). Though some literature is available on the reproductive biology of dioecious nutmeg (Armstrong & Drummond 1986), there is no report available for monoecious nutmeg trees.
Here we attempt to study the floral diversity of monoecious nutmeg.

The study on sex differentiation and variability in monoecious nutmeg was carried out at ICAR-Indian Institute of Spices Research (IISR), Kozhikode during June to September 2015. Total of 53 trees of 18 year old were selected randomly from the germplasm maintained at ICAR IISR and were studied for sex of tree from December 2014 onwards. The sex of flowers were recorded in all the trees at monthly interval. Out of the 53 trees of the selected population 6 trees are found to be monoecious. These 6 monoecious trees are used in the present study.

Details of 6 monoecious trees conserved at ICAR-IISR, Chelavoor:

<table>
<thead>
<tr>
<th>Plant number</th>
<th>Source</th>
<th>Nature of plant</th>
</tr>
</thead>
<tbody>
<tr>
<td>C- 1</td>
<td>ICAR-IISR, Experimental Farm, Peruvannamuzhi</td>
<td>Seedling</td>
</tr>
<tr>
<td>C- 9</td>
<td>ICAR-IISR, Experimental Farm, Peruvannamuzhi</td>
<td>Seedling</td>
</tr>
<tr>
<td>C- 21</td>
<td>ICAR-IISR, Experimental Farm, Peruvannamuzhi</td>
<td>Seedling</td>
</tr>
<tr>
<td>C- 43</td>
<td>ICAR-IISR, Experimental Farm, Peruvannamuzhi</td>
<td>Seedling</td>
</tr>
<tr>
<td>C -45</td>
<td>ICAR-IISR, Experimental Farm, Peruvannamuzhi</td>
<td>Seedling</td>
</tr>
<tr>
<td>C -54</td>
<td>Farmers field Ankola</td>
<td>Seedling</td>
</tr>
</tbody>
</table>

Within the 6 monoecious trees observations on inter and intra floral variability in monoecious for anther length, number of anthers, filament length, ovary length and flower opening type were recorded. Total of 100 flowers / month were collected from each tree and observations were recorded for variability during peak flowering season June – September, 2015.

Pollen viability test was done using Acetocarmine staining method. Pollen viability estimated using glycerolacetocarmine (Marks 1954). The pollen viability percentage determined as the ratio of the number of viable pollen to the total pollen number.

ANOVA for the floral attributes were done using SAS 9.3 software.

Based on the observations made on the phenological measurements inter and intra flower variability in monoecious nutmeg flowers are prominent. Three types of flowers are recorded in monoecious trees namely, pistillate, staminate and hermaphrodite flowers. The structure of hermaphrodite in nutmeg is reported for the first time. The flowers are borne on the leaf axil, flowering habit of the three types of flowers are seen in cymes as well as solitary in the same tree. All the types of flowers were light creamy yellow colour in monoecious tree, with thick gamosepalous perianth which bursts as trilobed or bilobed (less frequent) lobes during anthesis as reported earlier (Armstrong & Drummond 1986). However, addition to this tetralobed and pentalobed are observed rarely in monoecious trees (Fig. 1). Tetralobed lobes are reported in M. fatua, this type of lobe opening helps in more access for insects (pollinators) to enter the flower (Sharma & Armstrong 2013). The androecium of staminate flowers consisted of adnate 7-13 anthers, with average of 9.30 anthers and 4.09 and 3.18 mm of anther length and filament length respectively, androecium with 8-10 anthers were reported earlier by Armstrong & Drummond (1986) in staminate flowers. The gynoecium consists of single ovary with a mean length of 5.28 mm with bifid stigma in the pistillate flowers (Tables 1 & 2). Intra flower variability is evident in case of hermaphrodite flowers. In hermaphrodite flower the androecium ranges with 1-4 anthers, in the form of fused filament or free filament or both; some anthers are fused with the gynoecium (Fig. 2); the length of the anther varies from 3-6 mm. Besides remnants of the stamen in the developed fruits of hermaphrodite flowers, staminodes are also observed in the flowers (Fig.
Fig. 1. Variation in flower opening in monoecious tree flowers [a) Dimerous, b) Trimerous, c) Tetramerous and d) Pentamerous]

Fig. 2. Intra flower variability in number and length of anthers of hermaphrodite flowers

Fig. 3. Developed fruits from monoecious trees [a) from pistillate flowers, b) from hermaphrodite flowers (remnants of the stamen) and c) Microscopic view (20x)]
Table 1. Analysis of variance for flower types and floral attributes

<table>
<thead>
<tr>
<th>Tree No</th>
<th>Average number of flowers</th>
<th>Staminate</th>
<th>Pistillate</th>
<th>Hermaphrodite</th>
<th>No. of anthers</th>
<th>Length of anthers (mm)</th>
<th>Length of filament (mm)</th>
<th>Length of ovary (mm)</th>
<th>No. of anthers</th>
<th>Length of ovary (mm)</th>
</tr>
</thead>
<tbody>
<tr>
<td>C-1</td>
<td>100.00</td>
<td>0 (0.71)</td>
<td>0 (0.71)</td>
<td>8.88</td>
<td>4.10</td>
<td>2.94</td>
<td>0 (0.71)</td>
<td>0 (0.71)</td>
<td>0 (0.71)</td>
<td>0 (0.71)</td>
</tr>
<tr>
<td>C-9</td>
<td>92.66</td>
<td>5.66 (2.17)</td>
<td>1.66 (1.28)</td>
<td>9.42</td>
<td>5.50</td>
<td>3.82</td>
<td>6.00 (2.45)</td>
<td>2.25 (1.38)</td>
<td>5.20 (2.27)</td>
<td></td>
</tr>
<tr>
<td>C-21</td>
<td>83.00</td>
<td>11.00 (2.87)</td>
<td>7.33 (2.43)</td>
<td>8.86</td>
<td>3.10</td>
<td>2.98</td>
<td>5.59 (2.36)</td>
<td>2.23 (1.46)</td>
<td>5.23 (2.28)</td>
<td></td>
</tr>
<tr>
<td>C-43</td>
<td>100.00</td>
<td>0 (0.71)</td>
<td>0 (0.71)</td>
<td>8.98</td>
<td>4.06</td>
<td>3.48</td>
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<td>0 (0.71)</td>
<td>0 (0.71)</td>
<td>0 (0.71)</td>
</tr>
<tr>
<td>C-45</td>
<td>100.00</td>
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<td>0 (0.71)</td>
<td>10.92</td>
<td>3.92</td>
<td>3.18</td>
<td>5.80 (2.41)</td>
<td>2.20 (1.44)</td>
<td>5.40 (2.32)</td>
<td></td>
</tr>
<tr>
<td>C-54</td>
<td>50.66</td>
<td>41.00 (6.38)</td>
<td>8.33 (2.58)</td>
<td>8.76</td>
<td>3.88</td>
<td>2.70</td>
<td>5.79 (2.30)</td>
<td>2.22</td>
<td>5.28 (2.21)</td>
<td></td>
</tr>
<tr>
<td>Mean</td>
<td>87.72</td>
<td>19.22 (2.26)</td>
<td>5.77 (1.40)</td>
<td>9.30</td>
<td>4.09</td>
<td>3.18</td>
<td>5.79 (2.30)</td>
<td>2.22</td>
<td>5.28 (2.21)</td>
<td></td>
</tr>
<tr>
<td>CD (P&lt;0.05)</td>
<td>12.97**</td>
<td>NS</td>
<td>NS</td>
<td>0.30**</td>
<td>0.17**</td>
<td>0.15**</td>
<td>0.10**</td>
<td>NS</td>
<td>0.23**</td>
<td></td>
</tr>
<tr>
<td>CV (%)</td>
<td>8.31</td>
<td>8.20</td>
<td>10.40</td>
<td>12.09</td>
<td>4.04</td>
<td>4.94</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

Values in the parenthesis are transformed.
**Table 2.** Mean, range and CV% for floral traits in monoecious nutmeg

<table>
<thead>
<tr>
<th>Category</th>
<th>Characters</th>
<th>Mean</th>
<th>Range</th>
<th>CV%</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sex of flowers</td>
<td>Staminate</td>
<td>88.37</td>
<td>50-100</td>
<td>21.33</td>
</tr>
<tr>
<td></td>
<td>Pistillate</td>
<td>9.11</td>
<td>0-50</td>
<td>171.71</td>
</tr>
<tr>
<td></td>
<td>Hermaphrodite</td>
<td>2.74</td>
<td>0-15</td>
<td>183.84</td>
</tr>
<tr>
<td>Staminate</td>
<td>No. of anther</td>
<td>9.30</td>
<td>7-13</td>
<td>11.49</td>
</tr>
<tr>
<td></td>
<td>Length of anther (mm)</td>
<td>4.09</td>
<td>3-6</td>
<td>20.25</td>
</tr>
<tr>
<td></td>
<td>Length of filament (mm)</td>
<td>3.18</td>
<td>2-4</td>
<td>16.74</td>
</tr>
<tr>
<td>Pistillate</td>
<td>Length of ovary (mm)</td>
<td>5.42</td>
<td>5-6</td>
<td>8.28</td>
</tr>
<tr>
<td>Hermaphrodite</td>
<td>No. of anther</td>
<td>2.22</td>
<td>1-4</td>
<td>43.58</td>
</tr>
<tr>
<td></td>
<td>Length of ovary (mm)</td>
<td>5.31</td>
<td>4-6</td>
<td>9.54</td>
</tr>
</tbody>
</table>

**Table 3.** Pollen viability in staminate and hermaphrodite flowers of monoecious nutmeg

<table>
<thead>
<tr>
<th>Flower type</th>
<th>Number of pollens observed</th>
<th>Viable pollen</th>
<th>Sterile pollen</th>
<th>% of viability</th>
</tr>
</thead>
<tbody>
<tr>
<td>Staminate flower</td>
<td>152.66</td>
<td>145.00</td>
<td>7.67</td>
<td>94.77</td>
</tr>
<tr>
<td>Hermaphrodite flower</td>
<td>124.33</td>
<td>104.33</td>
<td>20.00</td>
<td>79.74</td>
</tr>
</tbody>
</table>


Air layering in cinnamon (*Cinnamomum verum* L.) under wet humid tropical conditions

A A Waman* & P Bohra

Division of Horticulture and Forestry, ICAR-Central Island Agricultural Research Institute, Port Blair-744 101, Andaman & Nicobar.

*E-mail: ajit.hort595@gmail.com

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Abstract

Cinnamon is a cross pollinated species and seed propagation has resulted in development of considerable variability in growth, yield and quality among the populations. Air layering has been considered as one of the efficient methods of multiplication in cinnamon. However, the success of layering varies greatly depending on the local environmental conditions. In the present investigation, air layering was performed at 20 days interval during rainy season (July 3rd to October 11th) of two consecutive years in Bay islands. Result revealed that first week of July was the most suitable time for air layering in cinnamon as it supported better rooting percentage (87.5%).

Keywords: Propagation; Season; Spice crop; Tropical islands

Cinnamon (*Cinnamomum verum* L.) or *dalchini* is one of the ancient tree spices grown in India. Though both bark as well as leaves are known to possess aromatic components mainly cinnamaldehyde and eugenol, the bark of this species is valued as a spice. It has been used in the form of dried bark, bark powder, oil and oleoresins. Cultivation of this spice in India is mainly confined to States of Andaman and Nicobar Islands, Kerala, Karnataka, North Eastern India and parts of Tamil Nadu and Maharashtra. Combined estimate of cinnamon and *tejpat* suggests that in India, it’s being grown on 2,770 ha area with about 5,050 t production (Indian Horticulture Database 2014). However, the production is not enough to meet the domestic demand and hence, cinnamon is being imported from other countries of the world (Indian Spices 2016). Increasing the productivity through development and adoption of improved technologies is a key factor in reducing the dependence on import.

Soil and climatic conditions of the Andaman and Nicobar Islands are well suited for the cultivation of cinnamon (Parthasarathy *et al.* 2009) and presently it is cultivated in about 150 ha yielding 40 t annually. Quality of cinnamon is assumed to be the finest in the islands compared with other parts of the country (Singh & Sankaran 2012). Large availability of interspaces in the coconut and arecanut plantations in the Andaman and Nicobar
Islands could be successfully utilized for its cultivation (Waman et al. 2016). However, existing plantations in the islands are of seedling origin. One can easily notice variations in cinnamon seedlings for leaf size, shape and colour of new flush, apart from the distinct chemotypes (Krishnamoorthy et al. 1988). Ergo, considerable variability is noticed in the yields and quality of the final produce and hence, vegetative propagation is of interest (Rema et al. 1997). The present report concerned an effort to identify the most appropriate time for carrying out air layering under island condition.

The present investigation was carried out in the Division of Horticulture and Forestry of ICAR- Central Island Agricultural Research Institute, Port Blair, Andaman and Nicobar Islands during 2015 and 2016. The islands exhibit typical tropical climate with average temperature of 18-31°C and annual rainfall of 3,100 mm distributed over May to December. Further, the relative humidity ranges between 60-90% in a year, while the average lies near to 70-80%.

For layering, healthy shoots of 25-30 cm length and 1.0-1.5 cm thickness were selected. Leaves and small branches near the ringing area on selected shoots were removed and two circular cuts were given to the shoots for removing a ring of bark of about an inch width. Commercial formulation of rooting hormone (Lipsa, Kolkata) was used for root induction and ringed portion was covered with soil: farmyard manure (1:1) before wrapping with polythene (20 cm × 20 cm). Experiment consisted of six treatments i.e. T<sub>1</sub>: layering on July 3, T<sub>2</sub>: layering on July 23, T<sub>3</sub>: layering on August 12, T<sub>4</sub>: layering on September 1, T<sub>5</sub>: layering on September 21 and T<sub>6</sub>: layering on October 11. Experiment was laid out in completely randomized design with 20 layers in each treatment. Various parameters were recorded at the time of separation (90 days after layering) and data was subjected to analysis of variance using Web Agri Statistical Package 2.0 (WASP 2.0, ICAR-RC for Goa, Ela, India).

Air layering was performed for six times during rainy season under island condition. Generally, rooting process is facilitated by the rains (Ranaware et al. 1995) and hence, the dry periods in the islands were avoided during the experimentation. Pooled analysis of two years data revealed that percentage root induction varied between 35.0% and 87.5% amongst the treatments studied (Table 1). Maximum rooting percentage was obtained in layers done on 3<sup>rd</sup> July (87.5%) followed by those done on 23<sup>rd</sup> July, while it was the lowest in layering performed on 21<sup>st</sup> September. Though rooting response varied considerably during different seasons, the mean number of primary roots per layer did not vary significantly (Table 1).

<table>
<thead>
<tr>
<th>Treatment</th>
<th>Rooting percentage</th>
<th>Number of primary roots layer&lt;sup&gt;a&lt;/sup&gt;</th>
<th>Mean length of primary root (cm)</th>
<th>Mean thickness of primary root (mm)</th>
</tr>
</thead>
<tbody>
<tr>
<td>July 3</td>
<td>87.5</td>
<td>5.8 a</td>
<td>5.52 b</td>
<td>2.74 a</td>
</tr>
<tr>
<td>July 23</td>
<td>57.5</td>
<td>4.2 a</td>
<td>7.37 a</td>
<td>2.33 a</td>
</tr>
<tr>
<td>August 12</td>
<td>47.5</td>
<td>4.2 a</td>
<td>5.18bc</td>
<td>2.04 a</td>
</tr>
<tr>
<td>September 1</td>
<td>50.0</td>
<td>3.7 a</td>
<td>8.53 a</td>
<td>2.17 a</td>
</tr>
<tr>
<td>September 21</td>
<td>35.0</td>
<td>5.3 a</td>
<td>3.60 c</td>
<td>2.15 a</td>
</tr>
<tr>
<td>October 11</td>
<td>43.2</td>
<td>3.8 a</td>
<td>4.00bc</td>
<td>2.20 a</td>
</tr>
</tbody>
</table>

*Means followed by same alphabet in a column do not differ significantly at 5% level of significance using least significant difference
Length of longest root varied significantly amongst the studied treatments (Table 1). Maximum length of primary root was observed in propagules obtained from cinnamon layered on 1st September (8.5 cm), which remained on par with layers of 23rd July. Similar to rooting percentage, length of root was found to be the lowest (3.6 cm) in layers of 21st September batch. Thickness of root ranged from 2.04 to 2.74 mm; however, the differences were non-significant amongst the treatments studied.

Similar to present communication, significant variations for rooting response and growth parameters have been reported from different agro-ecological regions of the country viz., West Bengal (Banerjee et al. 1982), Karnataka (Hegde et al. 1989) and Maharashtra (Ranaware et al. 1995). These reports have suggested the positive effects of rainy period on layering success; however, the raining season and pattern vary significantly in different regions and hence, location specific studies are required. As no reports are available for island conditions, present study was conducted. The prevalence of optimum microclimate during the part of the year would possibly have helped in maximizing the rooting success. Rema et al. (1997) reviewed different propagation methods in spice crops in which they supported the possible role of endogenous auxins, sugars and other biochemical constituents in variable rooting success during different seasons.

It could be concluded that first week of July is the most suitable time for carrying out air layering under Andaman and Nicobar Islands condition. This technique being easier, farmers can easily multiply superior types in large number. In addition, inputs required for the purpose are easily available at cheaper cost and hence, it could be a boon for on farm production of quality planting material in the Andaman Islands.

Acknowledgements

Authors are thankful to the Director of the Institute for providing necessary facilities for conducting the experiment.

References


IHD 2014 Indian Horticulture Database, National Horticulture Board, Gurgaon, India, 286 p.


Stability analysis for seed yield and yield attributing traits in fennel (*Foeniculum vulgare* Mill.)

S L Sawargaonkar*, A K Singh & S Sahu

*College of Agriculture & Research Station, Indira Gandhi Krishi Vishwavidyalaya, Raigarh-496 001, Chhattisgarh.*

*E-mail: shrikant.sawargaonkar@gmail.com*

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**Abstract**

Study was carried to evaluate phenotypic stability of 13 genotypes of fennel for seed yield and yield attributing traits. Out of 13 genotypes, three were checks (RF 205, RF 201 and local fennel) grown in Randomized Block Design (RBD) with three replications in each year. The significant genotypic differences were observed for all the characters suggesting differential responses of genotypes to the environmental changes. The results showed that among 13 genotypes RF 205 and Local fennel were found stable for seed yield plant⁻¹, FNL 69 for days to flower, FNL 70 and RF 101 for primary branches plant⁻¹, FNL 74 for umbels umbellet⁻¹ and FNL 71 for umbels plant⁻¹. The results revealed that some reliable predictions about G x E interaction as well as its unpredictable components contributed significantly in determining the stability of genotypes. The wider stability recorded by RF 205, RF 201 and local fennel during all the three years which will be useful for development of high yielding varieties of fennel.

**Keywords:** fennel, *Foeniculum vulgare* mill., stability, superiority

Fennel (*Foeniculum vulgare* mill.) is a highly cross pollinated and very important seed spice crop exhibiting 82.2% to 91.4% natural out crossing (Ramanujam *et al.* 1964). The hybridization of diverse genotypes followed by selection in segregating crop will be helpful in identifying heterozygous and heterogenous progeny. Insects plays an important role in maintaining heterogeneity in fennel crop. The existing varieties were developed using mass selection. Fennel seeds are having medicinal importance as carminative, cardiotonic, stimulant, vermicide and lactagogue (Lal 2014). The research efforts were for enhancing the productivity through developing high yielding inbreds and pure line varieties. India is the largest producer, consumer and exporter of spices and spice products. The total production of spices during 2013-14 was 5.9 MT from an area of 3.16 million ha (Annual Report 2014-15). Fennel had very little share to total production of spices. This yield gap can be minimized by growing suitable cultivars with appropriate packages of practices. For this development of high yielding and insect and pest resistant with good quality stable variety is essential. The newly variety must perform consistent performance across the
environments and over the years for high yield and other yield attributing and quality traits. Phenotypically stable genotypes are of great importance because environmental conditions vary from season to season. Wider adaptation to a particular environment and consistent performance of recommended genotypes is one of the main objectives in breeding programmes. A differential response of fennel genotypes when grown under different environments in the *Rabi* season has been reported by few scientists (Lal 2014; Drazic et al. 2014). Hence information on availability of stable high yielding varieties of Fennel is lacking. Keeping this in view, the present investigation was undertaken to determine the genotype x environment (year) interaction on stability parameters and to identify the stable and responsive genotypes for yield and yield contributing characters of fennel for Chhattisgarh.

The experimental materials consisted of 13 genotypes obtained from All India Coordinated Research Project (AICRP) on spices, Indian Institute of Spices Research (IISR), Kozhikode, Kerala, evaluated for stability of seed yield and yield attributing traits for the three years during 2012-13 to 2014-15 at the experimental farm of College of Agriculture and Research Station (CARS), Raigarh (21.9°N, 83.4°E, 215 m), Indira Gandhi Krishi Vishwavidyalaya (IGKV), Chhattisgarh. Out of 13 genotypes RF 205, RF 201 and local fennel were standard checks grown in Randomized Block Design (RBD) with three replications. Twenty plants in each row in plot size of 4 × 2.5 m, were maintained after thinning. The inter- and intra-row spacing was kept at 45 cm and 20 cm, respectively. The recommended package of cultural practices was followed to raise a good crop. In each plot, five competitive plants were identified randomly for recording data on days to 50% flowering, plant height (cm), primary branches plant⁻¹, umbels plant⁻¹, umblets umbel⁻¹ and seed yield plot⁻¹ (kg). The data recorded during three years were subjected to stability analysis according to the model proposed by Eberhart & Russel (1966) and three stability parameters mean (m), regression coefficient (bi) and the deviation from linearity (S²di) were estimated.
The genotypic differences were found to be highly significant for all the traits in each environment (year). The mean genotypic values from different year were subjected to pooled analysis. The mean sum of squares (MSS) due to genotypes (G) and environments (E) were significant for all the traits except for primary branches plant$^{-1}$ when tested against MSS due to genotype x environments. It revealed the non significant differential response of the varieties to the changing environments. The results were in close conformity to the findings of Lal (2008), Verma & Solanki (2015). The MSS due to G x E when tested against pooled error, were found highly significant for all the characters. Thus stability analysis was carried out for all the traits. The variance due to G x E were divided in to G x E (Linear) and due to pooled deviation (Non-linear). The G x E (Linear) mean squares were found significant for all the traits except primary branches plant$^{-1}$ indicating the presence of predictable components where as significance of pooled deviation for seed yield plot$^{-1}$, days to 50% flowering, plant height (cm), primary branches plant$^{-1}$ and umbels plant$^{-1}$ showed the presence of non-predictable components. These observations indicated that some reliable predictions about G x E interaction as well as its unpredictable components can be made for these traits. Hence, both components contributed significantly in determining the stability of genotypes (Lal 2008; Verma & Solanki 2015). Among 13 genotypes RF 205 and Local fennel were found stable for seed yield plant$^{-1}$, FNL 69 for days to flower, FNL 70 and RF 101 for primary branches plant$^{-1}$, FNL 74 for umbels umbel$^{-1}$ and FNL 71 for umbels plant$^{-1}$ (Table 2).

For days to flower, umbelets/umbel and umbels plant$^{-1}$ FNL 69 had recorded wider stability (above average mean, bi = 1 and S$^2$di = 0). The genotype FNL 70 was stable for plant height, primary branches plant$^{-1}$, umbelets umbel$^{-1}$ and umbels plant$^{-1}$ but had low seed yield, indicating its adaptation to stress environments (Lal 2008) while genotype FNL 71 recorded stability for plant height and umbels plant$^{-1}$. For umbels umbel$^{-1}$ FNL 72 had high stability across the environments of Raigarh. The genotypes FNL 69, FNL 70 and FNL 71 were among the top entries which had their mean umbels plant$^{-1}$ greater than the average of all the genotypes with regression coefficient (bi = 1) and non-significant deviation for regression (S$^2$di = 0). This indicated their high stability over the different environments of years (Lal 2008). The genotype FNL 68 and RF 101 had above average mean for umbels plant$^{-1}$, S$^2$di = 0 but the value of bi #1, indicating their adaptation to high input conditions. For umbels plant$^{-1}$ genotype RF 205 and FNL 67 had bi = 1 and S$^2$di = 0, but their mean was low, indicating their adaptation to stress environments (Table 3).

Table 2. Analysis of variance (mean squares) for phenotypic stability of yield and yield contributing characters in Fennel

<table>
<thead>
<tr>
<th>Source</th>
<th>df</th>
<th>Seed yield (kg ha$^{-1}$)</th>
<th>Days to 50% flow</th>
<th>Plant height</th>
<th>Prim. Branch</th>
<th>Umbels plant$^{-1}$</th>
<th>Umblets umbel$^{-1}$</th>
</tr>
</thead>
<tbody>
<tr>
<td>Total</td>
<td>38</td>
<td>48198.2</td>
<td>71.8</td>
<td>194.5</td>
<td>2.0</td>
<td>34.0</td>
<td>48.5**</td>
</tr>
<tr>
<td>Genotypes</td>
<td>12</td>
<td>49045.9*</td>
<td>25.4*</td>
<td>273.9**</td>
<td>3.2</td>
<td>35.4**</td>
<td>21.4**</td>
</tr>
<tr>
<td>ENV</td>
<td>2</td>
<td>507452.1*</td>
<td>1008.4**</td>
<td>1221.2**</td>
<td>1.7</td>
<td>145.1**</td>
<td>21.4**</td>
</tr>
<tr>
<td>Environment (Gen × Env)</td>
<td>26</td>
<td>47806.9*</td>
<td>25.**</td>
<td>157.8**</td>
<td>1.4</td>
<td>33.4**</td>
<td>61.1**</td>
</tr>
<tr>
<td>Environment Linear</td>
<td>1</td>
<td>1014905.6*</td>
<td>17**</td>
<td>2442.0**</td>
<td>3.4</td>
<td>290.3**</td>
<td>1218.5**</td>
</tr>
<tr>
<td>Gen. × Env Linear</td>
<td>12</td>
<td>17107.5*</td>
<td>25.1**</td>
<td>122.7**</td>
<td>0.3</td>
<td>44.7**</td>
<td>30.3**</td>
</tr>
<tr>
<td>Pooled Deviation</td>
<td>13</td>
<td>1752.6*</td>
<td>8.2*</td>
<td>14.5**</td>
<td>2.3**</td>
<td>3.2**</td>
<td>0.1</td>
</tr>
<tr>
<td>Pooled error</td>
<td>13</td>
<td>878.4</td>
<td>8.2</td>
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<td>0.1</td>
<td>0.1</td>
<td>6.8</td>
</tr>
<tr>
<td>S.N.</td>
<td>Genotypes</td>
<td>Seed yield (kg ha(^{-1}))</td>
<td>Days to flower</td>
<td>Plant height (cm)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>------</td>
<td>------------</td>
<td>-----------------------------</td>
<td>----------------</td>
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<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td>MEAN</td>
<td>bi</td>
<td>MSD</td>
<td>MEAN</td>
<td>B</td>
<td>MSD</td>
</tr>
<tr>
<td>1</td>
<td>FNL 67</td>
<td>470.8</td>
<td>0.9</td>
<td>-784.2</td>
<td>93.4</td>
<td>1.5</td>
<td>11.6**</td>
</tr>
<tr>
<td>2</td>
<td>FNL 68</td>
<td>374.9</td>
<td>0.9</td>
<td>-357.9</td>
<td>90.2</td>
<td>0.9</td>
<td>17.7**</td>
</tr>
<tr>
<td>3</td>
<td>FNL 69</td>
<td>425.2</td>
<td>1.0</td>
<td>1623.5</td>
<td>86.6</td>
<td>1.3**</td>
<td>-0.1</td>
</tr>
<tr>
<td>4</td>
<td>FNL 70</td>
<td>420.5</td>
<td>0.7</td>
<td>-796.2</td>
<td>87.5</td>
<td>1.1</td>
<td>5.2**</td>
</tr>
<tr>
<td>5</td>
<td>FNL 71</td>
<td>574.3</td>
<td>1.0</td>
<td>2262.9</td>
<td>84.3</td>
<td>0.8</td>
<td>10.3**</td>
</tr>
<tr>
<td>6</td>
<td>FNL 72</td>
<td>652.1</td>
<td>0.3</td>
<td>1070.8</td>
<td>87.8</td>
<td>0.1</td>
<td>1.1**</td>
</tr>
<tr>
<td>7</td>
<td>FNL 73</td>
<td>514.6</td>
<td>1.5</td>
<td>6542.8**</td>
<td>89.9</td>
<td>0.8</td>
<td>24.3**</td>
</tr>
<tr>
<td>8</td>
<td>FNL 74</td>
<td>587.6</td>
<td>1.6**</td>
<td>1211.1</td>
<td>91.0</td>
<td>1.6*</td>
<td>6.4**</td>
</tr>
<tr>
<td>9</td>
<td>FNL 75</td>
<td>503.4</td>
<td>1.8**</td>
<td>1845.3</td>
<td>94.4</td>
<td>0.7</td>
<td>8.1**</td>
</tr>
<tr>
<td>10</td>
<td>FNL 76</td>
<td>462.8</td>
<td>1.4</td>
<td>-668.1</td>
<td>92.0</td>
<td>1.1</td>
<td>0.8**</td>
</tr>
<tr>
<td>11</td>
<td>RF 101</td>
<td>673.4</td>
<td>0.3</td>
<td>655.5</td>
<td>89.1</td>
<td>1.1</td>
<td>15.4**</td>
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<tr>
<td>12</td>
<td>RF 205</td>
<td>808.4</td>
<td>0.7</td>
<td>-795.9</td>
<td>92.2</td>
<td>0.6</td>
<td>0.9**</td>
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<tr>
<td>13</td>
<td>Local check</td>
<td>696.3</td>
<td>0.9</td>
<td>-444.8</td>
<td>87.4</td>
<td>1.3</td>
<td>4.3**</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>S.N.</th>
<th>Genotypes</th>
<th>Prim branches</th>
<th>Umblets umbel(^{-1})</th>
<th>Umblets plant(^{-1})</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>MEAN</td>
<td>B</td>
<td>MSD</td>
</tr>
<tr>
<td>1</td>
<td>FNL 67</td>
<td>8.2</td>
<td>0.6</td>
<td>1.2**</td>
</tr>
<tr>
<td>2</td>
<td>FNL 68</td>
<td>8.6</td>
<td>1.8</td>
<td>0.0</td>
</tr>
<tr>
<td>3</td>
<td>FNL 69</td>
<td>9.2</td>
<td>1.5</td>
<td>1.6**</td>
</tr>
<tr>
<td>4</td>
<td>FNL 70</td>
<td>8.0</td>
<td>0.3</td>
<td>0.8**</td>
</tr>
<tr>
<td>5</td>
<td>FNL 71</td>
<td>6.9</td>
<td>-0.6</td>
<td>6.9**</td>
</tr>
<tr>
<td>6</td>
<td>FNL 72</td>
<td>7.1</td>
<td>0.8</td>
<td>0.7**</td>
</tr>
<tr>
<td>7</td>
<td>FNL 73</td>
<td>7.6</td>
<td>1.8</td>
<td>0.0</td>
</tr>
<tr>
<td>8</td>
<td>FNL 74</td>
<td>10.2</td>
<td>2.0</td>
<td>6.3**</td>
</tr>
<tr>
<td>9</td>
<td>FNL 75</td>
<td>7.1</td>
<td>2.6</td>
<td>1.8**</td>
</tr>
<tr>
<td>10</td>
<td>FNL 76</td>
<td>7.0</td>
<td>1.9</td>
<td>3.2**</td>
</tr>
<tr>
<td>11</td>
<td>RF 101</td>
<td>8.0</td>
<td>0.6</td>
<td>0.5**</td>
</tr>
<tr>
<td>12</td>
<td>RF 205</td>
<td>7.0</td>
<td>0.1</td>
<td>0.2**</td>
</tr>
<tr>
<td>13</td>
<td>Local check</td>
<td>6.8</td>
<td>-0.4</td>
<td>5.7</td>
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</tbody>
</table>
Table 4. Two way table showing stable and unstable genotypes for seed yield verses component characters

<table>
<thead>
<tr>
<th>Genotype</th>
<th>Stable genotypes</th>
<th>Stability parameters (bi/S2di) of yield components</th>
</tr>
</thead>
<tbody>
<tr>
<td>no</td>
<td>Days to flower</td>
<td>Plant height</td>
</tr>
<tr>
<td>12 RF 205 (C)</td>
<td>S/S</td>
<td>S/*</td>
</tr>
<tr>
<td>13 Local fennel (C)</td>
<td>S/*</td>
<td>S/S</td>
</tr>
</tbody>
</table>

Where, S=stable genotypes; S/*, */S and */*=unstable genotype; bi=regression coefficient; S2di=deviation from regression line; *: bi ≠1, S2di ≠0

Based on stability parameters genotypes RF 205 and Local fennel were classified as stable for seed yield plant⁻¹, which had their mean seed yield greater than the average of all the genotypes with unit regression coefficient (bi =1) and non-significant deviation from regression (S'di =0). This indicated their high stability over all the environments during 2012-13 to 2014-15. Lal (2008) and Verma & Solanki (2015) reported above average stability of Fennel genotypes for seed yield. The genotypes RF 101, FNL 72 and FNL 71 had above average mean, seed yield, regression coefficient of bi =1 but non-significant deviation from regression line (S'di=0). Hence, indicating its specific adaptability under good agronomic management practices.

Table 5. Superiority of fennel genotypes for yield over checks during 2012-13 to 2014-15

<table>
<thead>
<tr>
<th>E.N.</th>
<th>E.name</th>
<th>Yield (kg ha⁻¹)</th>
<th>% superiority</th>
<th>Disease reaction</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td>RF 101(NC)</td>
<td>Local check</td>
</tr>
</tbody>
</table>
| 2012-13
| 6     | FNL 67 | 578.1           | NO             | 2.75 MR         |
| 11    | RF 101 (c) | 599.7       | NO             | MR              |
| 12    | RF 205 (c) | 690.2       | NO             | MR              |
| 13    | FNL (LC)    | 562.6         | NO             | MR              |

| 2013-14
| 6     | FNL 67 | 686.9           | NO             | 6.36 MR         |
| 11    | RF 101 (c) | 777.1       | NO             | MR              |
| 12    | RF 205 (c) | 630.3       | NO             | MR              |
| 13    | FNL (LC)    | 918.4         | 25.2           | 2.5 MR          |
|      | FNL 74 | 914.9           | 24.7           | 2.1 MR          |
|      | FNL 75 | 872.0           | 18.9           | -2.7 MR         |
|      | FNL 73 | 807.3           | 10.0           | -9.9 MR         |
|      | FNL 71 | 770.4           | 5.0            | -14.0 MR        |
|      | RF 205 | 957.9           | 30.6           | 6.9 MR          |
|      | Local check | 896.2       | 22.2           | 0.0 MR          |
|      | RF 101 | 733.6           | 0.0            | MR              |

Where, No=No superiority; NC=National check
For the present study the meteorological parameters recorded with Indian meteorological standard week of 31 for first week of sowing, 37 for vegetative stage, 43 for flowering stage and 49 for physiological maturity stage respectively for 2012-13 to 2014-15 (Table 6). It was observed that during sowing period average optimum temperature of 27.6°C + 21.2°C were present over all the three years. At the same time average maximum humidity of 88.2% with minimum of 66.9% were recorded during first week of sowing. During vegetative stage of fennel crop mean maximum temperature of 24.9°C and minimum temperature of 15.2°C with humidity range of 87.2% to 60.7% recorded. At flowering time consistent maximum minimum temperature of 24.9°C-15.2°C respectively registered with average humidiy of 89.2% to 38.4% present. At physiological maturity mean maximum-minimum temperature of 32.0°C -15.0°C were registered while maximum humidity of 81.8% and minimum temperature of 37.3% were recorded. It was observed that at all the three years no rainfall during throughout the crop period.

The genotype FNL 67 had recorded 2.75 and 6.36 percent superiority for seed yield over local check during 2012-13 and 2013-14 respectively.

Table 6. Meterological parameters during crop growth period from 2012-13 to 2014-15

<table>
<thead>
<tr>
<th>Year</th>
<th>Temperature</th>
<th>Humidity</th>
<th>Rain fall (MM)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>(°C)</td>
<td>(%)</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Max</td>
<td>Min</td>
<td>Max</td>
</tr>
<tr>
<td>Seed germination week</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>2012-13</td>
<td>27.85</td>
<td>18.57</td>
<td>90.71</td>
</tr>
<tr>
<td>2013-14</td>
<td>27.86</td>
<td>23.86</td>
<td>86</td>
</tr>
<tr>
<td>2014-15</td>
<td>27</td>
<td>21.28</td>
<td>88</td>
</tr>
<tr>
<td>Average</td>
<td>27.6</td>
<td>21.2</td>
<td>88.2</td>
</tr>
<tr>
<td>Vegetative stage</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>2012-13</td>
<td>25.14</td>
<td>16.28</td>
<td>90.71</td>
</tr>
<tr>
<td>2013-14</td>
<td>24.29</td>
<td>13</td>
<td>80.28</td>
</tr>
<tr>
<td>2014-15</td>
<td>25.14</td>
<td>16.25</td>
<td>90.71</td>
</tr>
<tr>
<td>Average</td>
<td>24.9</td>
<td>15.2</td>
<td>87.2</td>
</tr>
<tr>
<td>Flowering stage</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>2012-13</td>
<td>25.42</td>
<td>11.42</td>
<td>88.71</td>
</tr>
<tr>
<td>2013-14</td>
<td>25.71</td>
<td>15.42</td>
<td>89.57</td>
</tr>
<tr>
<td>2014-15</td>
<td>25.42</td>
<td>11.42</td>
<td>88.71</td>
</tr>
<tr>
<td>Average</td>
<td>25.5</td>
<td>12.8</td>
<td>89.0</td>
</tr>
<tr>
<td>Physiological maturity</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>2012-13</td>
<td>32.71</td>
<td>14.71</td>
<td>83</td>
</tr>
<tr>
<td>2013-14</td>
<td>26.14</td>
<td>13.43</td>
<td>81.57</td>
</tr>
<tr>
<td>2014-15</td>
<td>37.14</td>
<td>16.71</td>
<td>80.71</td>
</tr>
<tr>
<td>Average</td>
<td>32.0</td>
<td>15.0</td>
<td>81.8</td>
</tr>
</tbody>
</table>
During 2014-15 five genotypes FNL 74, FNL 75, FNL 73, FNL 71, and FNL 76 recorded high heterosis for seed yield over RF 205 and RF 101, whereas FNL 74 & FNL 75 registered positive heterosis over local check.

It was observed that national check (RF 205) showed wider stability for seed yield, days to flower, primary branches plant$^{-1}$, umbels plant$^{-1}$ and umbelets umbel$^{-1}$ while local fennel had wider stability for seed yield (kg ha$^{-1}$), plant height, umbels plant$^{-1}$ and umbelets umbel$^{-1}$. While none of the test entries showed superior stable performance over the national and local checks during all the three years. This appears to be evidence for much greater genotype × environment interaction for the entries to be evaluated than for the pure lines. This is somewhat unexpected in view of the greater homeostasis in unfavorable environments usually found in heterozygous genotypes. This needs further study.

Acknowledgements

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References


Effect of irrigation methods and mulching on growth and yield parameters of chilli (*Capsicum annuum* L.) in arid condition

R L Bhardwaj, M M Sundria, S R Kumhar & N Kumar

*Agricultural Research Station, Agriculture University, Jodhpur-342 304, Rajasthan.*

*E-mail: rajubhardwaj3@gmail.com*

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**Abstract**

The research work was carried out to study the impact of various irrigation methods and mulching on plant growth, production and profitability of chilli cv. R.Ch. 1 at Agricultural Research Station, Mandor, Jodhpur during July, 2016 to February, 2017. The results of surface irrigation were compared with drip irrigation system under no mulch and in conjunction with plastic mulch. The results revealed that the crop was irrigated by drip irrigation on raise bed with 100 micron Linear Low Density Poly Ethylene plastic mulch (T8 treatment) exhibited significantly higher seedling survival at 15 and 30 days after transplanting (95.16% and 91.70%), highest plant height (47.10 cm at 45 DAT and 54.60 cm at harvest), highest number of branches (14.93) plant\(^{-1}\), maximum stem girth (2.32 cm) number of roots plant\(^{-1}\) (138.5), highest fruit set (38.47%), length of fresh fruit (12.56 cm), diameter of fruit (3.52 cm) and fresh weight of fruit\(^{-1}\) (8.42g) was observed. The maximum number of fruits plant\(^{-1}\) (125), highest yield plant\(^{-1}\) (1052.5g), yield ha\(^{-1}\) (337.63q) and premier fruit quality score (9.11) with maximum net return (Rs.326407.28) and benefit: cost ratio (3.41) was also reported in same treatment. Comparatively minimum time (15 hours) required for one hectare irrigation was also reported in drip irrigation on raise bed with plastic mulch. This led to lower population of white fly plant\(^{-1}\) (4.53), minimum weed infestation (1.53 weed m\(^{-2}\)), leaf curl (5.50%) and fruit rot (5.0%) incidence than other treatment combinations. The minimum growth, yield and profitability were reported in check basin method of irrigation without mulch (T\(_1\) treatment).

**Keywords:** *Capsicum annuum*, Day After Transplanting, leaf curl, surface irrigation, survival

Chilli (*Capsicum annuum* L.) is an important commercial vegetable cum spice crop of India belongs to the family Solanaceae. The production of chilli crop is affected adversely by moisture deficit. Productivity of the crop can be increased by adopting improved package of practices, particularly *in-situ* moisture conservation by mulching as well as high-tech irrigation especially drip irrigation with appropriate irrigation scheduling. Use of soil cover and mulching is also known to be beneficial chiefly through their influence on soil moisture conservation, solarization and control of weeds. Beneficial response of plants
to mulch includes early production, more yield and reduced insect and disease problems (Pattanaik et al. 2003). Linear Low Density Poly Ethylene (LLDPE) plastic films have been proved as better mulch because of their puncture resistance quality, thinness and lower cost (Panda 2004). Numerous experiments have reported the benefits of LLDPE mulch in several crops, but research is limited on response of chilli production in western Rajasthan by this method. Keeping this in background, the present study was undertaken to study the effect of different irrigation methods and mulching on chilli crop and compare the result with the conventional method of growing the crop under surface irrigation without mulch.

A field experiment was conducted at Agricultural Research Station, Mandor, Jodhpur (Rajasthan), India during kharif seasons of year 2016-17. The soil of experimental plot was of sandy loam texture with average pH range 8.5, having organic carbon 0.55%, available N 180 kg ha$^{-1}$, P 27.5 kg ha$^{-1}$ and K 250.0 kg ha$^{-1}$ during experimentation. The experiment was conducted in a completely randomized design having nine treatments comprising by different irrigation methods and mulching viz., $T_1$ = Check basin method, $T_2$ = furrow irrigation method, $T_3$ = Raise bed with trench method, $T_4$ = Flat bed with drip irrigation, $T_5$ = Flat bed + plastic mulch + drip irrigation, $T_6$ = Raise bed with drip irrigation, $T_7$ = Raise bed + organic mulch + drip irrigation, $T_8$ = Raise bed + plastic mulch + drip irrigation, $T_9$ = Sprinkler irrigation method. In well prepared field, transplanting of Chilli seedlings variety RCh 1 of 35-40 days old were planted in pair row method with a spacing of 45cm x 45 cm/90 cm (33,333 plant ha$^{-1}$) during last week of June. In check basin and sprinkler system of irrigation the bed size is 2 x 2 meter and in all other methods is 1 x 4 meters. The cultural practices of the crop were followed as per the recommendations. The organic material and LLDPE silver colour film of 100-micron thickness was used for mulching around the plant. The lateral lines of 12 mm diameter LLDPE pipes were laid along with crop rows.

The laterals were provided with inlet drippers of 8 litre hr$^{-1}$ discharge capacity. All the observations were taken from five randomly selected plant of each replication throughout the investigation period at appropriate time by adopting standard method for growth, development, fruiting behavior and yield.

Seedling survival per cent (after transplanting in main field at 15 DAT and 30 DAT) was recorded by following formula;

\[
\text{Survival percent} = \frac{\text{Total survival transplanted plants}}{\text{Total transplanted plants}} \times 100
\]

Plant height (at 45 DAT and at harvesting) was measured from soil surface upto the highest shoot tip by straightening all branches. Stem girth was measured 1 cm from the base of the stem using vernier calliper. Observation of number of branches, days taken to first flower initiation, duration of fruiting period and number of fruit plant$^{-1}$ was recorded by standard counting method. Number of roots, root length was measured by destructive method of uprooting the plants and taking measurement by standard method. Length of fresh fruits measured by scale and fruit diameter using vernier calliper and expressed in centimeter. Fruit set per cent was recorded by following formula;

\[
\text{Fruit set per cent} = \frac{\text{Total number of fruit set plant}^{-1}}{\text{Total number of flowers plant}^{-1}} \times 100
\]

Fruit weight was determined by weighing method at the time of harvesting and expressed in gram fruit$^{-1}$. The total fruit yield plant$^{-1}$ and hectare$^{-1}$ was calculated by weighing total marketable fruits and has been expressed in gram and quintal respectively. Further, the net return was calculated by subtracting cost of each treatment from gross return. The gross return was calculated from yield multiplied by average market rate during the period of investigation. The benefit cost ratio was calculated by dividing net return to total cost of cultivation. Benefit-Cost ratio and net profit were carried out to determine the economic feasibility of the crop using surface and drip irrigation as suggested by Tiwari et al. (1998a).
The seasonal system cost of drip irrigation system included depreciation, prevailing bank interest rate, and repair and maintenance cost of the system. The fixed cost of drip irrigation system was determined to be Rs 112,000 ha⁻¹. The useful life of drip system was considered to be 10 years. The system cost was evaluated by distributing the fixed cost of system over life period of drip irrigation set. For calculating depreciation, the life of the drip irrigation set and 10% junk value was considered. The interest was calculated on the average of investment of the drip irrigation set taking into consideration the value of the set in the first and last year @10% per annum. Cost of repairs and maintenance of set is @2% of initial cost of the drip irrigation set per year. The cost of cultivation includes expenses incurred in land preparation, interculture operation, fertilizer, crop protection measures, irrigation water and harvesting with labour charges. Therefore, total seasonal cost was worked as: depreciation, interest, repairs and maintenance cost of set + cost of cultivation + cost of mulch. The income from produce was calculated using prevailing average market price of capsicum @ Rs 1250 q⁻¹. Disease incidence (leaf curl and fruit rot) and quality of fruits was measured by visual inspection (Five member team of crop experts and plant pathologist). White fly population plant⁻¹ and weed infestation meter⁻² was calculate by simple counting method. The time required for irrigation was calculated as per actual required time of irrigation of specified area by different methods of irrigation. To test the significance of variance of data obtained from crop growth, yield and economics of variance technique for completely randomized design was done by standard procedure prescribed by Panse & Sukhatme (1985). Significance of difference among the treatments effect was tested by 'F' test and critical difference (CD) was calculated, wherever the results were significant.

The results revealed that, the irrigation methods and mulching are significantly influenced growth attributes at all the growth stages (Table 1). The maximum seedling survival per cent at 15 DAT (95.10%) and 30 DAT (91.70%) was recorded in T₈ treatment, which was significantly superior to other treatment but at par with T₆ and T₇ treatments. The maximum survival per cent of seedling in T₈ treatment might be due to more favourable moisture condition for seedling transplanting and re-establishment of roots than others. The height of plant under treatment T₈ (47.10 cm at 45 DAT) and treatment T₆ (62.60 cm at harvest) was found highest among all other treatments and is 67.19% and 13.40% higher than the T₁ treatment. About to number of branch plant⁻¹, maximum value was recorded in treatment T₈ (14.93) followed by treatment T₇ (12.53) and the lowest value was in treatment T₁ (7.17). Maximum stem girth at harvest (2.36 cm) and highest number of roots plant⁻¹ (138.50) were observed in T₈ treatment whereas longest root system (10.50 cm) was observed in T₃ treatment. The minimum stem girth (1.68 cm) and the number of roots plant⁻¹ (53.57) were observed in T₁ treatment whereas shortest root system (7.97 cm) was observed in T₉ treatment. The higher available moisture status in soil favourably influences the uptake of nutrients which maintains the cell turgidity, cell elongation, photosynthesis and respiration at optimum level, leading to favourable growth and development of plant in terms of plant height, number of branches plant⁻¹, stem girth and number of root plant⁻¹ in the present study. The highest increase in vegetative growth in drip irrigation with mulching might be due to the availability of soil moisture as well as favourable temperature at optimum level for plant growth development (Pattanaik et al. 2003; Paul et. al. 2013). The lowest value of vegetative growth in T₁ might be because of unfavourable moisture regime (moisture stress or excess moisture) in the soil through surface irrigation and competition of weeds for nutrients (Pattanaik et al. 2003; Agrawal & Agrawal 2005). The increased growth attributes might have supplied water and nutrients in adequate proportion, which resulted in triggering the production of plant growth hormone, viz., indole acetic acid (IAA) and
higher number of leaves and roots throughout the cropping period (Sankar et al. 2008).

The drip irrigation in combination with mulch significantly increased the yield of chilli as compared to drip irrigation without mulch (Table 2) and surface irrigation methods. The minimum days (42.38) required for first flower initiation was reported in T9 treatment whereas the maximum days (51.39) was required in T5 treatment. Among various treatments, highest fruit set (38.47%), length of fresh fruit (12.56 cm), diameter of fruit (3.52 cm), duration of fruiting (71.38 days), fresh weight of fruit (12.5 g), maximum number of fruits plant-1 (125), highest yield plant-1 (1052.5 g) and yield ha-1 (337.63 q) was recorded under T8 treatment, whereas lowest yield (153.45 q ha-1) was recorded under T1 treatment. This might be due to water stress during the critical growth period and fruit development stage coupled with aeration problem in first few days immediately after irrigation. Another reason to get low yield by surface irrigation without mulch might be due to less availability of nutrients for crop growth due to leaching and high weed competition between the crops (Pattanaik et al. 2003). In drip irrigation system on raise bed with plastic mulch the water is applied at a low rate for a longer period at frequent intervals near the plant root zone through lower pressure delivery system, which increases the availability of nutrients near the root zone with a reduction in leaching losses and minimum weed competition. More nutrient availability, especially near the root zone might have increased the translocation of photosynthetese to storage organ of chilli resulting in an increased weight of fruits. This result corroborated the findings of Singh (2007), Sankar et al. (2008), Paul et al. (2013) and Kumar et al. (2016).

Irrigation methods and mulching also significantly influenced the gross return, net return and benefit cost ratio in chilli (Table 3). Maximum net profit of Rs. 326407.28 ha-1 with B: C ratio of 3.41 was recorded in T8 treatment followed by Rs 296192.61 ha-1 with B: C ratio of 3.11 in T5 treatment and lowest net profit of Rs 119007.80 ha-1 with a B: C ratio of 1.63 in T1 treatment (Table 3). It is observed that, the drip

**Table 1. Effect of different irrigation methods and mulching on vegetative parameter of chilli**

<table>
<thead>
<tr>
<th>Treatments</th>
<th>Seedling survival %</th>
<th>Plant height (cm)</th>
<th>Average number of branches plant-1</th>
<th>Stem girth at harvest (cm)</th>
<th>Length of root (cm)</th>
<th>No. of roots plant-1</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>15 DAT</td>
<td>30 DAT</td>
<td>45 DAT</td>
<td>At harvest</td>
<td></td>
<td></td>
</tr>
<tr>
<td>T1</td>
<td>78.17</td>
<td>73.93</td>
<td>28.17</td>
<td>55.20</td>
<td>7.17</td>
<td>1.68</td>
</tr>
<tr>
<td>T2</td>
<td>82.37</td>
<td>79.23</td>
<td>29.07</td>
<td>57.30</td>
<td>8.07</td>
<td>1.86</td>
</tr>
<tr>
<td>T3</td>
<td>86.97</td>
<td>83.83</td>
<td>32.07</td>
<td>61.70</td>
<td>8.27</td>
<td>1.88</td>
</tr>
<tr>
<td>T4</td>
<td>88.40</td>
<td>84.73</td>
<td>34.03</td>
<td>61.55</td>
<td>9.48</td>
<td>2.26</td>
</tr>
<tr>
<td>T5</td>
<td>90.10</td>
<td>86.33</td>
<td>38.53</td>
<td>61.70</td>
<td>10.28</td>
<td>2.33</td>
</tr>
<tr>
<td>T6</td>
<td>92.50</td>
<td>89.33</td>
<td>40.43</td>
<td>62.60</td>
<td>9.98</td>
<td>2.32</td>
</tr>
<tr>
<td>T7</td>
<td>93.40</td>
<td>90.90</td>
<td>41.97</td>
<td>53.20</td>
<td>12.53</td>
<td>2.30</td>
</tr>
<tr>
<td>T8</td>
<td>95.10</td>
<td>91.70</td>
<td>47.10</td>
<td>54.60</td>
<td>14.93</td>
<td>2.36</td>
</tr>
<tr>
<td>T9</td>
<td>85.20</td>
<td>80.10</td>
<td>34.90</td>
<td>56.70</td>
<td>9.73</td>
<td>2.00</td>
</tr>
<tr>
<td>S.Em. +</td>
<td>1.646</td>
<td>1.539</td>
<td>2.173</td>
<td>1.859</td>
<td>0.442</td>
<td>0.088</td>
</tr>
<tr>
<td>CD (P&lt;0.05)</td>
<td>4.871</td>
<td>4.556</td>
<td>6.432</td>
<td>5.504</td>
<td>1.327</td>
<td>0.260</td>
</tr>
</tbody>
</table>
Irrigation with mulched treatments T5, T7 and T8 gave better net return with higher B:C ratio ha⁻¹ than their corresponding treatments without mulching in conventional irrigation method. The highest net return (US$ 7098 ha⁻¹), incremental net return (US$ 1556 ha⁻¹), and incremental benefit-cost ratio (7.03) were found for 50% water application with straw mulch (Biswas et al. 2015). The results are in conformity with the findings of Singh (2007), Sankar et al. (2008) and Kumar et al. (2016). Apart from reducing water consumption, drip irrigation with mulching also helps in reducing cost of cultivation and improving productivity of crops as compared to the same crops cultivated under flood method of irrigation (Paul et al. 2013).

Irrigation time significantly pretentious by different irrigation methods. The minimum time required in irrigation (14.50 hours) in T5 treatment which closely followed by T8 treatment. Drip irrigation method with or without mulching required less irrigation time than without mulching in conventional irrigation method. There was significant effect of LLDPE mulch over drip irrigation system alone. Drip irrigation with LLDPE mulching (T5 & T8 treatment) saving irrigation time (21.40 hour ha⁻¹ and 21.35 hour ha⁻¹) upto 60 per cent by reducing water losses and increased irrigation efficiency. The increase in water saving per cent in trench method (T2), drip irrigation system alone (T4), drip irrigation system with LLDPE mulch (T5 & T8), drip irrigation with organic mulch (T7) and sprinkler system (T9) over conventional surface irrigation by check basin method (T1) was 38.8%, 46.6%, 60.0%, 54.4% and 41.9% respectively. The highest water use efficiency of 592 kg ha⁻¹ mm⁻¹ was obtained with 50 per cent water application under polyethylene mulch (Biswas et al. 2015). Drip irrigation with mulching helps to achieve yield gains of upto 100 per cent, water savings of upto 40-80 per cent, and associated fertilizer, pesticide, and labour savings over conventional irrigation systems in capsicum crop (Paul et al. 2013). Similar trend has been reported in water use efficiency for okra crop by Tiwari et al. (1998a) and for tomato crop by Singh (2007).

### Table 2. Effect of different irrigation methods and mulching on yield and yield attributes of chilli

<table>
<thead>
<tr>
<th>Treatments</th>
<th>First flower initiation (DAT)</th>
<th>Fruit set %</th>
<th>Length of fresh fruit (cm)</th>
<th>Diameter of fresh fruit (cm)</th>
<th>Diameter of fresh fruit (cm)</th>
<th>Weight of single fresh fruit (gm)</th>
<th>No. of fruits plant⁻¹</th>
<th>Yield plant⁻¹ (gm)</th>
<th>Yield ha⁻¹ (q)</th>
</tr>
</thead>
<tbody>
<tr>
<td>T1</td>
<td>45.70</td>
<td>17.23</td>
<td>10.75</td>
<td>2.87</td>
<td>52.70</td>
<td>7.63</td>
<td>60.33</td>
<td>460.34</td>
<td>153.45</td>
</tr>
<tr>
<td>T2</td>
<td>47.70</td>
<td>17.01</td>
<td>11.30</td>
<td>2.75</td>
<td>54.00</td>
<td>7.79</td>
<td>65.33</td>
<td>508.95</td>
<td>169.65</td>
</tr>
<tr>
<td>T3</td>
<td>47.40</td>
<td>21.31</td>
<td>11.35</td>
<td>2.78</td>
<td>56.10</td>
<td>7.82</td>
<td>68.33</td>
<td>534.37</td>
<td>178.12</td>
</tr>
<tr>
<td>T4</td>
<td>49.19</td>
<td>27.00</td>
<td>12.01</td>
<td>3.01</td>
<td>62.39</td>
<td>7.97</td>
<td>74.17</td>
<td>591.11</td>
<td>197.03</td>
</tr>
<tr>
<td>T5</td>
<td>51.39</td>
<td>31.81</td>
<td>12.19</td>
<td>3.14</td>
<td>68.59</td>
<td>8.23</td>
<td>114.17</td>
<td>939.59</td>
<td>313.19</td>
</tr>
<tr>
<td>T6</td>
<td>49.99</td>
<td>30.89</td>
<td>12.39</td>
<td>3.22</td>
<td>60.39</td>
<td>8.26</td>
<td>89.17</td>
<td>736.52</td>
<td>245.50</td>
</tr>
<tr>
<td>T7</td>
<td>46.58</td>
<td>36.74</td>
<td>12.30</td>
<td>3.28</td>
<td>66.38</td>
<td>8.36</td>
<td>95.00</td>
<td>794.20</td>
<td>251.53</td>
</tr>
<tr>
<td>T8</td>
<td>48.38</td>
<td>38.47</td>
<td>12.56</td>
<td>3.52</td>
<td>71.38</td>
<td>8.42</td>
<td>125.00</td>
<td>1052.50</td>
<td>337.63</td>
</tr>
<tr>
<td>T9</td>
<td>42.38</td>
<td>16.27</td>
<td>11.57</td>
<td>3.10</td>
<td>47.58</td>
<td>7.90</td>
<td>75.00</td>
<td>592.50</td>
<td>184.30</td>
</tr>
<tr>
<td>S.Em. +</td>
<td>1.247</td>
<td>1.570</td>
<td>0.201</td>
<td>0.059</td>
<td>1.247</td>
<td>0.133</td>
<td>1.302</td>
<td>9.913</td>
<td>8.257</td>
</tr>
<tr>
<td>CD (P&lt;0.05)</td>
<td>3.693</td>
<td>4.649</td>
<td>0.595</td>
<td>0.176</td>
<td>3.693</td>
<td>0.395</td>
<td>3.854</td>
<td>29.337</td>
<td>24.437</td>
</tr>
</tbody>
</table>
Occurrence of chilli leaf curling and fruit rot was detected throughout the investigation period. The best performance, with a marked reduction in leaf curling and fruit rot and improve fruit quality was observed in drip irrigation with LLDPE mulching (Table 3). The minimum incidence of fruit rot (5.0%), leaf curing (5.5%) and highest quality score of fruit (9.11) was observed in T8 treatment which was closely followed by T7 and T5 treatments whereas maximum incidence of fruit rot (20.02%) and leaf curling (15.84%) was reported in check basin method of irrigation (T1 treatment) whereas minimum quality score (6.50) was observed in T9 treatment. Presence of white fly and weed was observed throughout the investigation period. The minimum population of white flies (4.53 plant\(^{-1}\)) and weed infestation (1.53 weed m\(^{-2}\)) was observed in T8 treatment whereas highest incidence of white flies (17.43 plant\(^{-1}\)) and weed infestation (30.03 weed m\(^{-2}\)) was observed in T1 treatment. This is due to the fact that in drip irrigation with mulching significantly reduced additional moisture level in field environment which in turn increase quality of fruits and reduce disease infestation, white fly population as well as it also trim down weed seed germination, growth and development. The increase in quality of fruits was due to the effective utilization of applied nutrients, water and significantly reduced weed growth; disease incidence and increased rate of photosynthesis, sink capacity and accumulated more amounts of dry matter and finally increased quality of fruits and yield. Conventional surface irrigation methods without mulching provide favorable environmental condition for increase insect population and development of disease as well as germinate and develop high density weed plants. The beneficial effect of drip irrigation and black LLDPE mulch in capsicum, tomato and okra was also reported earlier by Horo et al. (2003); Singh (2007); Vankar & Shinde (2007), Bhardwaj & Sarolia (2012), Paul et al. (2013).

The combination of raised bed + drip irrigation system with LLDPE mulching is observed to be economical and cost effective as compared with conventional surface irrigation without mulching. Thus, the use of drip irrigation system either alone or in combination with

### Table 3. Effect of different irrigation methods and mulching on economics, fruit quality, water saving (%), insect-pest and weed infestation of chilli

<table>
<thead>
<tr>
<th>Treatments</th>
<th>Net Return</th>
<th>B:C ratio</th>
<th>Quality score of fruits</th>
<th>Water saving (%)</th>
<th>No. of white fly plant(^{-1})</th>
<th>Fruit Rot (%)</th>
<th>Leaf curl (%)</th>
<th>Weed Infestation</th>
</tr>
</thead>
<tbody>
<tr>
<td>T1</td>
<td>119007.80</td>
<td>1.63</td>
<td>7.24</td>
<td>36.30 (0.0%)</td>
<td>17.43</td>
<td>20.02</td>
<td>15.84</td>
<td>30.03</td>
</tr>
<tr>
<td>T2</td>
<td>135858.99</td>
<td>1.78</td>
<td>7.46</td>
<td>22.20 (38.8%)</td>
<td>13.73</td>
<td>18.02</td>
<td>12.34</td>
<td>17.43</td>
</tr>
<tr>
<td>T3</td>
<td>144550.55</td>
<td>1.85</td>
<td>8.04</td>
<td>22.45 (38.2%)</td>
<td>12.43</td>
<td>14.02</td>
<td>10.84</td>
<td>15.53</td>
</tr>
<tr>
<td>T4</td>
<td>170392.68</td>
<td>2.24</td>
<td>8.00</td>
<td>19.40 (46.6%)</td>
<td>11.60</td>
<td>10.82</td>
<td>9.50</td>
<td>12.40</td>
</tr>
<tr>
<td>T5</td>
<td>296192.61</td>
<td>3.11</td>
<td>8.50</td>
<td>14.50 (60.0%)</td>
<td>7.90</td>
<td>7.82</td>
<td>7.30</td>
<td>2.10</td>
</tr>
<tr>
<td>T6</td>
<td>225678.88</td>
<td>2.78</td>
<td>8.20</td>
<td>18.15 (50.0%)</td>
<td>10.40</td>
<td>8.82</td>
<td>8.60</td>
<td>10.10</td>
</tr>
<tr>
<td>T7</td>
<td>230913.36</td>
<td>2.77</td>
<td>8.50</td>
<td>16.55 (54.4%)</td>
<td>9.53</td>
<td>6.00</td>
<td>8.10</td>
<td>5.73</td>
</tr>
<tr>
<td>T8</td>
<td>326407.28</td>
<td>3.41</td>
<td>9.11</td>
<td>14.55 (60.0%)</td>
<td>4.53</td>
<td>5.00</td>
<td>5.50</td>
<td>1.53</td>
</tr>
<tr>
<td>T9</td>
<td>152772.53</td>
<td>1.97</td>
<td>6.50</td>
<td>21.10 (41.9%)</td>
<td>15.43</td>
<td>19.00</td>
<td>13.40</td>
<td>32.18</td>
</tr>
<tr>
<td>S.Em. +</td>
<td>0.001</td>
<td>0.005</td>
<td>0.162</td>
<td>1.247</td>
<td>0.346</td>
<td>0.568</td>
<td>0.227</td>
<td>0.394</td>
</tr>
<tr>
<td>CD (P&lt;0.05)</td>
<td>0.003</td>
<td>0.017</td>
<td>0.480</td>
<td>3.693</td>
<td>1.026</td>
<td>1.68</td>
<td>0.672</td>
<td>1.166</td>
</tr>
</tbody>
</table>
mulching, could increase the chilli yield quality of fruits and profitability. It also reduces white fly population, disease incidence (root rot and leaf curling) and minimise with crop weed competition. Drip irrigation with mulching increase water use efficiency by significant reduction in irrigation time ha⁻¹. It is concluded that the drip irrigation method with LLDPE mulching is suitable for chilli production in arid and semi arid condition of western Rajasthan.

References


Evaluation of stable and non shattering isabgol cultivar - Gujarat isabgol 4


Seed Spices Research Station, Sardarkrushinagar Dantiwada Agricultural University, Jagudan-382 710, Mahesana, Gujarat.

*E-mail: hirenpbg@yahoo.co.in

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Abstract

Isabgol growing area was surveyed during 2009-10 and selected genotypes were evaluated in preliminary evaluation trial (PET) during 2010-13 and in large scale varietal trial (LSVT) during 2013-15. It was found that JI-09-21 recorded better growth and yield characters and yield over check Gujarat Isabgol 3. The JI-09-21 did not shatter much even after water dripping and recommended for cultivation as Gujarat Isabgol 4.

Keywords: gujarat isabgol 4, isabgol, new variety, non shattering type

Isabgol (Plantago ovata Forsk) is a short duration, more remunerative and medicinally important crop of arid and semiarid regions. In India, it is largely grown in Gujarat, Rajasthan, Madhya Pradesh and Haryana. The area under Isabgol in India during 2014-15 is 1.09 lakh hectares. The production recorded 72 thousand MT with productivity of 660 kg ha⁻¹. In Gujarat, area mostly falls in Banaskantha, Kachchh and Patan districts with acreage of nine thousand hectare with production and productivity 5000 MT and 556 kg ha⁻¹, respectively during 2015-16 (Anonymous 2016a). During the last decade, area and production of isabgol has decreased to the tune of 343% and 281%, respectively due to problems of seed shattering. At the time of maturity, unseasonal rain or heavy dew leads to failure of the crop which is the fact for reduction of area under Isabgol (Anonymous 2016b). The objective of the study was to evolve non-shattering isabgol cultivars.

Isabgol growing area of Kachchh in Gujarat was surveyed during 2009-10 and subsequently genotypes were evaluated. First three years (2010-11 to 2012-13) 13 genotypes (JI-09-03, 07, 10, 13, 15, 16, 20, 21, 22, 23, 24, 25 and 26) with check (Gujarat Isabgol -03) were evaluated in preliminary evaluation – trial (PET). During 2013-14 and 2014-15, nine genotypes (JI-08-02, JI-09-07, 13, 16, 20, 21, 22, 24 and 25) along with check (Gujarat Isabgol -03) was tested. The trials were carried out at three different locations viz., Jagudan, Kholwada and Deesa in a randomized block design with three replications.

The stability analysis of variance and stability parameters viz., linear regression coefficient (bi) and deviation from regression (S²di) of genotype means over environment were computed as suggested by Eberhart & Russell (1966).

Shattering per cent was computed as suggested by Singh et al. (2005) and Chandra (1967). Five
plants in each replication were selected at maturity stage. Entire spike were dipped in water and then observed for seed shattering from selected plants. The percentage of seed shattering calculated by using a following formula.

\[
\text{Shattering per cent} = \left(\frac{\text{Expected seed yield} - \text{Seed wt. after threshing}}{\text{Expected seed yield}}\right) \times 100
\]

Expected seed yield= H.I. x Sun Dry wt. (kg)/100

Considering eight trials at three locations for five years, JI-09-21 was recorded higher (928 kg ha\(^{-1}\)) seed yield against 830 kg ha\(^{-1}\) of Gujarat Isabgol 3 (GI 3), which was 11.78% higher than GI-3 (Table 1). The new variety named as Gujarat Isabgol 4 was having more tillers plant\(^{-1}\) (6.0), more spikes plant\(^{-1}\) (22.7), seeds spike\(^{-1}\) (80.7), higher test weight (1.58 gm) and high swelling factor (11.4 cc g\(^{-1}\)) than that of check variety GI-3, due to these yield contributing traits GI-4 is having high seed yield potential (Table 3). The similar kind of results also obtained by Prajapati et al. (2011).

Gujarat Isabgol 4 recorded high mean with regression coefficient (bi) near unity and deviation from regression (\(S^{2}\)) around zero for seed yield, indicating GI-4 has average responsiveness and are highly stable over environments (Table 2).

The new culture has compact spike and did not separate easily even after dipping in water. Only 7.25% seeds shattered after dipping in water.

Table 1. The comparative yield performance (kg ha\(^{-1}\)) of JI-09-21 (GI 4) over different locations

<table>
<thead>
<tr>
<th>Year</th>
<th>Trial</th>
<th>Yield (kg ha(^{-1}))</th>
<th>IOC (%)</th>
<th>Rank</th>
<th>S.Em ±</th>
<th>CD</th>
<th>C.V. %</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>JI-09-21</td>
<td>GI-3</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Jagudan</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>2010-11</td>
<td>PET</td>
<td>1014</td>
<td>894</td>
<td>13.42</td>
<td>1/14</td>
<td>56</td>
<td>167</td>
</tr>
<tr>
<td>2011-12</td>
<td>PET</td>
<td>1078</td>
<td>948</td>
<td>13.71</td>
<td>1/14</td>
<td>49</td>
<td>150</td>
</tr>
<tr>
<td>2012-13</td>
<td>PET</td>
<td>907</td>
<td>863</td>
<td>5.10</td>
<td>2/14</td>
<td>51</td>
<td>155</td>
</tr>
<tr>
<td>2013-14</td>
<td>LSVT-II</td>
<td>946</td>
<td>874</td>
<td>8.24</td>
<td>1/10</td>
<td>52</td>
<td>155</td>
</tr>
<tr>
<td>Mean (5)</td>
<td></td>
<td>979</td>
<td>878</td>
<td>11.46</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Kholwada</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>2013-14</td>
<td>LSVT-II</td>
<td>878</td>
<td>769</td>
<td>14.17</td>
<td>1/10</td>
<td>52</td>
<td>154</td>
</tr>
<tr>
<td>2014-15</td>
<td>LSVT-II</td>
<td>852 *</td>
<td>759</td>
<td>12.25</td>
<td>1/10</td>
<td>31</td>
<td>91</td>
</tr>
<tr>
<td>Mean (2)</td>
<td></td>
<td>865</td>
<td>764</td>
<td>13.22</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Deesa</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Overall Mean (8 trials)....</td>
<td></td>
<td>928</td>
<td>830</td>
<td>11.78</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
</tbody>
</table>

Table 2. Stability Analysis for seed yield in Isabgol

<table>
<thead>
<tr>
<th>Variety</th>
<th>Yi (mean seed yield)(kg plot(^{-1}))</th>
<th>bi(reg. coeff.)</th>
<th>(S_{d}^{2})(mean Sq. dev.)</th>
</tr>
</thead>
<tbody>
<tr>
<td>JI-09-21(GI 4)</td>
<td>0.82</td>
<td>0.94</td>
<td>0.00</td>
</tr>
<tr>
<td>GI-3 (Ch)</td>
<td>0.42</td>
<td>0.80</td>
<td>0.00</td>
</tr>
<tr>
<td>Mean</td>
<td>0.39</td>
<td>-</td>
<td>-</td>
</tr>
</tbody>
</table>
Table 3. Comparative yield and quality attributes of JI-09-21 (GI 4)

<table>
<thead>
<tr>
<th>Character</th>
<th>Mean</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>JI-09-21 (GI 4)</td>
</tr>
<tr>
<td>Days to flowering</td>
<td>63</td>
</tr>
<tr>
<td>Days to maturity</td>
<td>102</td>
</tr>
<tr>
<td>Plant height (cm)</td>
<td>31.0</td>
</tr>
<tr>
<td>No. of tillers plant&lt;sup&gt;1&lt;/sup&gt;</td>
<td>6.0</td>
</tr>
<tr>
<td>No. of spikes plant&lt;sup&gt;2&lt;/sup&gt;</td>
<td>22.7</td>
</tr>
<tr>
<td>Spike length (cm)</td>
<td>4.7</td>
</tr>
<tr>
<td>No. of seeds spike&lt;sup&gt;1&lt;/sup&gt;</td>
<td>80.7</td>
</tr>
<tr>
<td>1000 grain weight (g)</td>
<td>1.579</td>
</tr>
<tr>
<td>Swelling factor (cc g&lt;sup&gt;-1&lt;/sup&gt;)</td>
<td>11.4</td>
</tr>
</tbody>
</table>

Table 4. Effect of water dipping on shattering of isabgol seed

<table>
<thead>
<tr>
<th>Variety</th>
<th>Without Dipping</th>
<th>H.I %</th>
<th>Dipping in Water (Average of four sample)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Dry wt. (kg)</td>
<td>Seed wt. (kg)</td>
<td>Dry wt. (initial) (kg)</td>
</tr>
<tr>
<td>GI 4 (JI-09-21)</td>
<td>1.027</td>
<td>0.175</td>
<td>17.03</td>
</tr>
<tr>
<td>GI 3</td>
<td>0.959</td>
<td>0.158</td>
<td>16.47</td>
</tr>
</tbody>
</table>

GI-4 recorded 89.67% less shattering than that of check variety, which revealed that Gujarat Isabgol 4 was non shattering in habit as compared to Gujarat Isabgol 3 which is prone to high seed shattering. The post dipped seed weight of GI 4, was 0.115 kg threshed seeds, which was 96% higher than that of the post dipped seed yield of GI-3 (0.037 kg threshed seeds) (Table 4). Hence, Gujarat Isabgol 4 recommended for cultivation.

Acknowledgments

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